NOTE: Please save this file locally before filling in the table, DO NOT work on the file within your internet browser as changes will not be saved. Adobe Acrobat Reader (available free here) is recommended for completion.

The ARRIVE guidelines 2.0: author checklist

The ARRIVE Essential 10

These items are the basic minimum to include in a manuscript. Without this information, readers and reviewers cannot assess the reliability of the findings.

Item		Recommendation	Section/line number, or reason for not reporting
Study design	1	For each experiment, provide brief details of study design including: a. The groups being compared, including control groups. If no control group has been used, the rationale should be stated. b. The experimental unit (e.g. a single animal, litter, or cage of animals).	Methods/Section"Animals, surgical procedures, and small interfering RNA injection"/Line 142-143 and 147-149 a single animal
Sample size	2	a. Specify the exact number of experimental units allocated to each group, and the total number in each experiment. Also indicate the total number of animals used. b. Explain how the sample size was decided. Provide details of any *a priori* sample size calculation, if done.	Methods/Section"Animals, surgical procedures, and small interfering RNA injection"/Line140, 142-143 and 147-149 1. To study the effect of Etv1 on myelination in vivo, an interference group and a control group were set up. At least 3 animals in each experimental group were used to verify
Inclusion and exclusion criteria	3	a. Describe any criteria used for including and excluding animals (or experimental units) during the experiment, and data points during the analysis. Specify if these criteria were established *a priori.* If no criteria were set, state this explicitly. b. For each experimental group, report any animals, experimental units or data points not included in the analysis and explain why. If there were no exclusions, state so. c. For each analysis, report the exact value of *n* in each experimental group.	Yes,Methods/Section"Quantification and statistical analysis"/Line 336-342. IIn order to ensure the authenticity of the experiment, we did not exclude the data of each experimental group. In addition, our experiments have standard procedures and Yes, Figure legends: Figure 1/Line 741-742; Figure 2/Line 755-756; Figure 3/Line 770, 773, 776, and 781; Figure 4/Line 792 and 797; Figure 5/Line 805 and 813; Figure /Line
Randomisation	4	a. State whether randomisation was used to allocate experimental units to control and treatment groups. If done, provide the method used to generate the randomisation sequence. b. Describe the strategy used to minimise potential confounders such as the order of treatments and measurements, or animal/cage location. If confounders were not controlled, state this explicitly.	Yes,Mice were randomly selected from 6-8 week old male mice with consistent body weight and assigned to the experimental and control groups We selected 6-8-week-old male mice of similar body weight for the experiment. The operation was performed by the same person. The control and experimental groups were housed in different cages (3 mice/cage), likewise, the mice were housed
Blinding	5	Describe who was aware of the group allocation at the different stages of the experiment (during the allocation, the conduct of the experiment, the outcome assessment, and the data analysis).	Methods/Section"Quantification and statistical analysis"/Line 340-342. The experiment was kept double-blind, that is, animal grouping, reagent injection, animal surgery, result evaluation and data analysis were performed by different experimenters.
Outcome measures	6	a. Clearly define all outcome measures assessed (e.g. cell death, molecular markers, or behavioural changes). b. For hypothesis-testing studies, specify the primary outcome measure, i.e. the outcome measure that was used to determine the sample size.	Yes, Methods/Section"SC differentiation assay"/Line 187-189; Methods/Section"SC migration assay "/Line 200-201 and Line 208-209; Methods/Section"SC proliferation Yes,Methods/Section"Quantification and statistical analysis"
Statistical methods	7	a. Provide details of the statistical methods used for each analysis, including software used. b. Describe any methods used to assess whether the data met the assumptions of the statistical approach, and what was done if the assumptions were not met.	Yes,Methods/Section"Quantification and statistical analysis" N/A
Experimental animals	8	a. Provide species-appropriate details of the animals used, including species, strain and substrain, sex, age or developmental stage, and, if relevant, weight. b. Provide further relevant information on the provenance of animals, health/immune status, genetic modification status, genotype, and any previous procedures.	Yes, Methods/Section"Animals, surgical procedures, and small interfering RNA injection"/Line 140 N/A
Experimental procedures	9	For each experimental group, including controls, describe the procedures in enough detail to allow others to replicate them, including: a. What was done, how it was done and what was used. b. When and how often. c. Where (including detail of any acclimatisation periods). d. Why (provide rationale for procedures).	Yes, Methods/Section"Animals, surgical procedures, and small interfering RNA injection"/Line 141-149 Methods/Section"SC differentiation assay"/Line 182-188; Yes,Methods/Section"Animals, surgical procedures, and small interfering RNA injection"/Line 141-149 Methods/Section"SC differentiation assay"/Line 182-188: Yes,Methods/Section"Animals, surgical procedures, and small interfering RNA injection"/Line 141-149 Methods/Section"SC differentiation assay"/Line 182-188: Yes,Methods/Section"Animals, surgical procedures, and small interfering RNA injection"/Line 141-149 Methods/Section"SC differentiation assay"/Line 182-188:
Results	10	For each experiment conducted, including independent replications, report: a. Summary/descriptive statistics for each experimental group, with a measure of variability where applicable (e.g. mean and SD, or median and range). b. If applicable, the effect size with a confidence interval.	Yes,Methods/Section"Quantification and statistical analysis" N/A

The Recommended Set

These items complement the Essential 10 and add important context to the study. Reporting the items in both sets represents best practice.

Item		Recommendation	Section/line number, or reason for not reporting
Abstract	11	Provide an accurate summary of the research objectives, animal species, strain and sex, key methods, principal findings, and study conclusions.	Yes, Abstract/Line 31-34, 35-46, and 47-57
Background	12	a. Include sufficient scientific background to understand the rationale and context for the study, and explain the experimental approach. b. Explain how the animal species and model used address the scientific objectives and, where appropriate, the relevance to human biology.	Yes, Introduction/Line63-74, 90-109 and 110-120 Yes, Introduction/Line66-70 and 119-120; Discussion/Line587-589
Objectives	13	Clearly describe the research question, research objectives and, where appropriate, specific hypotheses being tested.	Yes, , Introduction/Line110-120;
Ethical statement	14	Provide the name of the ethical review committee or equivalent that has approved the use of animals in this study, and any relevant licence or protocol numbers (if applicable). If ethical approval was not sought or granted, provide a justification.	Yes,Methods/Section"Animals, surgical procedures, and small interfering RNA injection"/Line 136-139
Housing and husbandry	15	Provide details of housing and husbandry conditions, including any environmental enrichment.	Yes,Methods/Section"Animals, surgical proced
Animal care and monitoring	16	a. Describe any interventions or steps taken in the experimental protocols to reduce pain, suffering and distress. b. Report any expected or unexpected adverse events. c. Describe the humane endpoints established for the study, the signs that were monitored and the frequency of monitoring. If the study did not have humane endpoints, state this.	Yes, Methods/Section"Animals, surgical procedures, and small interfering RNA injection"/Line 149-150 N/A N/A
Interpretation/ scientific implications	17	a. Interpret the results, taking into account the study objectives and hypotheses, current theory and other relevant studies in the literature. b. Comment on the study limitations including potential sources of bias, limitations of the animal model, and imprecision associated with the results.	Yes, Introduction/Line 90-109, 110-119; Results and Discussion Yes, Discussion/579-589
Generalisability/ translation	18	Comment on whether, and how, the findings of this study are likely to generalise to other species or experimental conditions, including any relevance to human biology (where appropriate).	Yes, this study can provide a candidate molecular target for clinical interventions in demyelinating diseases
Protocol registration	19	Provide a statement indicating whether a protocol (including the research question, key design features, and analysis plan) was prepared before the study, and if and where this protocol was registered.	A protocol was prepared before the study without registration
Data access	20	Provide a statement describing if and where study data are available.	Yes, Footnote/Line 602-603
Declaration of interests	21	a. Declare any potential conflicts of interest, including financial and non-financial. If none exist, this should be stated. b. List all funding sources (including grant identifier) and the role of the funder(s) in the design, analysis and reporting of the study.	Yes, Footnote/Line 605-607 Yes, Acknowledgments/Line 593-597

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