

ALDOB represents a potential prognostic biomarker for patients with clear cell renal cell carcinoma

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Background: Previous studies have shown that aldolase B (ALDOB) might play controversial roles in multiple types of cancer, which could act as a cancer-promoting factor or a cancer-inhibiting factor depending on the subtype of the cancer. However, the role of ALDOB in clear cell renal cell carcinoma (ccRCC) patients has not been clearly elucidated. Therefore, this study aimed to comprehensively explore the expression level, prognostic value, functional enrichment, immune infiltration, and N6-methyladenosine (m6A) modification of ALDOB in ccRCC patients.

Methods: A total of 1,070 ccRCC tissues and 409 normal tissues from the Gene Expression Omnibus (GEO) database, The Cancer Genome Atlas (TCGA) database, and the ArrayExpress database were enrolled to evaluate the expression level and prognostic value of ALDOB in ccRCC. The Kaplan-Meier survival curves and the Log-Rank test were performed to assess the prognostic value. The univariate and multivariate Cox regression analysis were used to identify the independent prognostic predictors in ccRCC patients. In addition, R version 4.2.0 with its suitable packages were used to perform the functional enrichment analysis, immune infiltration analysis, and m6A methylation analysis. Statistical significance was set at the P value <0.05.

Results: The expression level of ALDOB was significantly down-regulated in ccRCC compared to normal tissue, and the ALDOB expression level was noticeably correlated with T stage, M stage, and histologic grade of patients with ccRCC. The survival analysis revealed that ALODB was the independent predictor of overall survival (OS), disease-specific survival (DSS), and progression-free survival (PFS) of ccRCC patients. In addition, the functional enrichment analysis showed that ALDOB and its related genes were mainly involved in the metabolism and metabolic pathways of multiple substances, including glycolysis, gluconeogenesis, and fatty acid degradation. Finally, the immune infiltration analysis and the m6A methylation analysis suggested that ALDOB was closely correlated with the infiltration abundance of immune cells and stromal cells in the tumor microenvironment and several types of m6A regulators in ccRCC.

Conclusions: As a potential prognostic biomarker for patients with ccRCC, downregulation of ALDOB was closely associated with the clinicopathological features, poor prognosis, immune infiltration, and m6A modification in ccRCC patients.

Keywords: Aldolase B (ALDOB); clear cell renal cell carcinoma (ccRCC); biomarker; prognosis

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Introduction

As one of the common malignancies of the urinary system, the incidence of renal cell carcinoma (RCC) worldwide has shown a yearly increase in recent years (1-3). Among the three main types of RCC, clear cell renal cell carcinoma (ccRCC) is the main pathological type of RCC with a worse prognosis compared to papillary RCC and chromophobe RCC, accounting for 70% to 80% of all RCC (4,5). In the clinic, the majority of ccRCC are detected incidentally via noninvasive imaging investigation, and the classic triad of flank pain, visible haematuria, and palpable abdominal mass is rare, that generally correlates with aggressive disease (6). Currently, due to the lack of powerful diagnostic tools and prognostic evaluation systems other than imaging modalities, it is difficult for clinicians to early diagnose and precisely predict the prognosis of ccRCC. Therefore, the development of effective molecular biomarkers and therapeutic targets becomes essential to improve the prognosis of patients with ccRCC.

Aldolase B, encoded by the *ALDOB* gene, is mainly involved in the conversion of fructose 1-phosphate to dihydroxyacetone phosphate and glyceraldehyde (7). Increasing evidence confirms that ALDOB not only

Highlight box

Key findings

• ALDOB represents a potential prognostic biomarker for patients with clear cell renal cell carcinoma.

What is known and what is new?

- ALDOB might play controversial roles in multiple types of cancer, which could act as a cancer-promoting factor or a cancer-inhibiting factor depending on the subtype of the cancer.
- The expression level of ALDOB was significantly down-regulated in ccRCC compared to normal tissue, and the ALDOB expression level was noticeably correlated with T stage, M stage, and histologic grade of patients with ccRCC. Furthermore, ALODB was the independent predictor of OS, DSS, and PFS of ccRCC patients.

What is the implication, and what should change now?

 ALDOB might be a novel prognostic biomarker and therapeutic target for ccRCC patients. The nomograms with more prognostic predictive power were constructed to predict the OS, DSS, and PFS of ccRCC patients by integrating T stage, N stage, M stage, histologic grade, and the expression level of ALDOB of ccRCC patients. Based on these nomograms, it would be helpful to assess the clinical outcome of ccRCC more accurately and provide more personalized prognostic assessment strategy for ccRCC patients. plays an important role in glycolysis and fructose metabolism, but is also aberrantly expressed in a variety of malignancies. And this abnormal expression pattern has been demonstrated to be highly associated with the clinicopathological features and prognosis of cancer patients (8-10). Tian et al. performed immunostaining to evaluate the expression level of the ALDOB protein in 172 patients with rectal adenocarcinomas treated with preoperative chemoradiotherapy followed by radical surgery (8). The results showed that overexpression of ALDOB was closely associated with clinicopathological factors of rectal adenocarcinoma patients, such as tumor advancement, lymphovascular invasion, and perineural invasion. And the survival analysis indicated that the high expression of ALDOB was significantly associated with worse disease-specific survival (DSS), which could be served as a prognostic biomarker for patients with rectal cancer (8). In addition, Li et al. used immunohistochemical staining to detect the protein expression level of ALDOB in 229 samples from patients with colorectal cancer who underwent primary tumor resection (9). The results showed that ALDOB overexpression was associated with poorer overall survival (OS) and disease-free survival (DFS) in patients with colorectal cancer. Further mechanistic study showed that ALDOB inactivation significantly inhibited the proliferation, migration, and invasive ability of colon cancer cells by participating in the regulation of the epithelialmesenchymal transition (9). Contrarily, in a study of gastric cancer, ALDOB was found to be down-regulated in tumor tissues compared with adjacent nontumor tissues (10). And the expression of ALDOB was closely related to the tumorinvasion depth, lymph node metastasis, distant metastasis, and tumor stage. Combined with the results of survival analysis, they revealed that ALDOB was an independent prognostic factor for OS in gastric cancer patients, suggesting that ALDOB could act as a novel molecular marker for gastric cancer patients (10). Therefore, based on the previous studies, ALDOB might play controversial roles in different types of cancers, which could act as a cancerpromoting factor or a cancer-inhibiting factor depending on the subtype of the cancers.

Although extensive researches have been carried out on the role of ALDOB in other malignancies, few researches evaluated the expression level of and prognostic value of ALDOB in ccRCC patients. In addition, no single study has explored the role and value of ALDOB in ccRCC patients. Therefore, the main purpose of this study is to comprehensively explore the expression level, prognostic

value, functional enrichment, immune infiltration, and N6methyladenosine (m6A) modification of ALDOB in ccRCC patients by integrating multiple datasets. This study not only attempts to elucidate the role and value of ALDOB in ccRCC, but also targets to develop a new prognostic biomarker and therapeutic target for ccRCC patients. We present the following article in accordance with the TRIPOD reporting checklist (available at https://tau. amegroups.com/article/view/10.21037/tau-22-743/rc).

Methods

Download and processing of original expression profiles

The gene expression profiles of GSE53757 (11), GSE53000 (12), GSE36895 (13), GSE15641 (14), GSE66272 (15), GSE68417 (16), GSE40435 (17), GSE12606 (18), GSE46699 (19), and GSE11151 (20), were obtained from the Gene Expression Omnibus (GEO) database, totally including 439 ccRCC tissues and 337 normal kidney tissues (Table S1). The RNA sequencing (RNA-seq) data of 530 ccRCC tissues and 72 normal kidney tissues with corresponding clinicopathological features and survival data were downloaded and standardized from The Cancer Genome Atlas (TCGA) database (TCGA-KIRC dataset) (21). Moreover, the E-MTAB-1980 dataset, which contained 101 ccRCC patients with RNA-seq and clinical data, was downloaded from the ArrayExpress database (22). Therefore, a total of 1,070 ccRCC tissues and 409 normal tissues was enrolled in this study. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013).

Identification and validation of differential expressed genes (DEGs)

Here, the DEGs between ccRCC and normal tissues were detected by the "limma" package in R software based on 439 ccRCC and 337 normal kidney tissues from ten GEO datasets. In this study, genes with P<0.05 and 1log₂(fold change)1>2 were considered as DEGs. The pivotal DEGs were acquired after the intersection of DEGs from ten GEO datasets by using the "UpSetR" package in R software. In addition, the expression levels of overlapping DEGs in 530 ccRCC and 72 normal kidney tissues from TCGA database were analyzed to further verify the results of the expression levels of pivotal DEGs obtained from the GEO datasets.

Prognostic value analysis of DEGs

The Kaplan-Meier survival curves were plotted, and the Log-Rank test was performed to assess the prognostic value of DEGs in ccRCC patients. And the primary outcomes of the survival analysis were OS, DSS, and progression-free survival (PFS). Here, hazard ratios (HRs) with 95% confidence intervals (CIs) were calculated to quantitatively estimate the prognostic value of DEGs. In addition, the univariate and multivariate Cox regression analyses were conducted to assess the influence on the survival of ccRCC patients within age, gender, T stage, N stage, M stage, AJCC stage, histologic grade, and DEGs expression level to further determine the independent predictors of OS, DSS, and PFS of ccRCC patients.

Expression analysis of ALDOB at mRNA and protein level

The mRNA expression level of ALDOB in ccRCC tissues and normal kidney tissues was analyzed and validated based on TCGA database as well as GEO database, respectively. Besides, the Clinical Proteomic Tumor Analysis Consortium (CPTAC) (https://proteomics.cancer.gov/programs/cptac) database was employed to investigate the protein expression level of ALDOB in normal tissues and tumor tissues (23). Furthermore, the Human Protein Atlas (HPA) (http:// www.proteinatlas.org/) database was utilized to visualize the immunohistochemistry images of ALDOB in ccRCC tissues and normal kidney tissues to verify the protein expression results of the CPTAC database (24). Finally, the AlphaFold Protein Structure Database (https://alphafold.ebi.ac.uk/) was used to predict the 3D protein structure of ALDOB (25).

Clinicopathological analysis of ALDOB

Based on the expression level of ALDOB and the clinicopathological data of ccRCC patients from TCGA database, the relationship between the mRNA expression level of ALDOB and T-stage, N-stage, M-stage, and pathological grade of ccRCC patients were determined. Furthermore, the logistic regression analyses were also used to validate the association of ALDOB and the clinicopathological characteristics of ccRCC patients from TCGA-KIRC dataset.

Construction and validation of nomograms

Based on the results of the Cox logistic regression analysis,

the prognostic predictors, including T stage, N stage, M stage, AJCC stage, pathological grade, and the expression level of ALDOB in ccRCC patients, were employed to construct the nomograms of 1-year, 3-year, and 5-year OS, DSS, and PFS of ccRCC patients from TCGA-KIRC dataset. Besides, the concordance index (C-index) was calculated by 1,000 bootstrap resamples, and the calibration curves were utilized to evaluate the predictive accuracy and reliability of the nomograms.

Protein-protein interaction network and functional enrichment analysis

In this work, a protein-protein interaction (PPI) network of ALDOB was built to predict the functional partners of ALDOB based on the Search Tool for the Retrieval of Interacting Genes/Proteins (STRING) database (26). Additionally, the Gene Ontology (GO) analysis and the Kyoto Encyclopedia of Genes and Genomes (KEGG) pathway analysis of ALDOB and its correlated genes were performed to reveal the potential functions and possible signaling pathways of ALDOB in ccRCC.

Immune infiltration analysis

Based on the "MCP-Counter" algorithm, this study analyzed the infiltration abundance of eight types of immune cells (T cells, CD8 T cells, cytotoxic lymphocytes, B lineage cells, natural killer cells, monocyte lineage, myeloid dendritic cells, and neutrophils) and two types of stromal cells (endothelial cells and fibroblasts) in ccRCC patients from the TCGA-KIRC dataset to explore the correlation between the expression level of ALDOB and the abundance of immune cells and stromal cells in the tumor microenvironment (TME) (27). Moreover, the "Estimate" package in R software was employed to calculate the Immune Score, Stromal Score, ESTIMATE Score, and Tumor Purity that respectively represented the immune component, stromal component, and the tumor purity for each ccRCC sample (28). Then, in order to determine whether ALDOB and ten types of immune cells and stromal cells could distinguish ccRCC patients, the "ConsensusClusterPlus" package in R software was used to perform consensus clustering analysis for subtyping the ccRCC patients from the TCGA-KIRC dataset. And the consensus matrix k value denoted the number of clusters. At the same time, the Kaplan-Meier survival curves and

the Log-Rank test were also performed to compare the prognosis between the two clusters. Finally, the "MCP-Counter" and "Estimate" algorithm were also used to evaluate the relative abundance of immune cells and TME within two different clusters.

m6A methylation analysis of ALDOB

Initially, the correlation between ALDOB and a total of 23 types of m6A regulators, including eight writers (METTL3, METTL14, RBM15B, RBM15, VIRMA, WTAP, CBLL1, and ZC3H13), two erasers (FTO and ALKBH5), and 13 readers (YTHDF1, YTHDF2, YTHDF3, YTHDC1, YTHDC2, HNRNPC, HNRNPA2B1, IGF2BP1, IGF2BP2, IGF2BP3, LRPPRC, FMR1, and ELAVL1) was examined to explore the potential mechanism for the abnormal expression pattern of ALDOB in ccRCC. Then, the least absolute shrinkage and selection operator (LASSO) regression was conducted for selecting and identifying a panel of genes within ALDOB and 23 m6A modulators via "glmnet and survival" package in R software based on TCGA-KIRC dataset. Next, the multivariate Cox regression analysis was applied to establish a prognostic signature and calculate Risk Scores for each ccRCC patients in TCGA-KIRC dataset. Afterward, 530 ccRCC patients from TCGA-KIRC dataset were classified into low-risk group and high-risk group according to the optimal cut-off value of the risk scores. Subsequently, the Kaplan-Meier survival curve was performed with the Log-Rank test to compare the prognosis between the two risk groups. Lastly, the time-dependent receiver operating characteristic (ROC) curve and the area under curve (AUC) values were used to analyze the sensitivity and specificity in evaluating the accuracy of the model that this study constructed.

Statistical analysis

The Mann-Whitney U test and Student's *t*-test was used to compare the differences between two groups. The Kruskal-Wallis H test and Dunn's test were performed to evaluate the relationships between clinicopathological features and the expression level of ALDOB. The Kaplan-Meier curves were depicted to assess the prognostic value of genes in ccRCC patients and the Log-Rank test was performed to assess the survival differences between two groups. The univariate and multivariate Cox regression analysis were



Figure 1 The flowchart of this study. ccRCC, clear cell renal cell carcinoma; GEO, Gene Expression Omnibus; DEGs, differential expressed genes; TCGA, The Cancer Genome Atlas; KIRC, kidney renal clear cell carcinoma; PPI, protein-protein interaction.

used to identify the independent prognostic predictors. The Pearson correlation test was conducted for the correlation analysis. In this study, the statistical analysis was performed using R software (version 4.2.0). And the figures were drawn with R software (version 4.2.0) and the corresponding online databases. Statistical significance was set at the P value <0.05 for all the analyses. The flow diagram of this study is presented in *Figure 1*.

Results

Identification and validation of DEGs

According to the set criteria of DEGs, this study analyzed the DEGs and depicted the volcano plots in each GEO dataset (Figure S1). As a result, a total of 24 DEGs was screened and determined after taking the intersection of DEGs of ten GEO datasets (Figure S2). Among them, 12 DEGs were up-regulated in ccRCC tissues, including CA9, ANGPTL4, HILPDA, NDUFA4L2, ENO2, NETO2, AHNAK2, SCARB1, LOX, APOC1, SCD, and C3. And 12 DEGs were expressed at significantly lower levels in ccRCC tissues, including KCNJ1, KNG1, HRG, UMOD, SFRP1, CLDN8, FGF9, ALDOB, ESRRG, MAL, EGF, and SCNN1A (Figure S2). In addition, this study verified the expression levels of 24 DEGs in 530 ccRCC tissues and 72 normal kidney tissues from the TCGA-KIRC dataset. As expected, the expression levels of 24 DEGs were same as those in ten GEO datasets (Figure 2).

Prognostic value of DEGs in ccRCC patients

In order to explore the prognostic value of DEGs in ccRCC patients, the univariate Cox regression analysis was first employed to perform the survival analysis based on the clinical data from TCGA-KIRC dataset. The results showed that among the 12 up-regulated genes, the expression levels of ANGPTL4, ENO2, NETO2, SCARB1, LOX, APOC1, and C3 were associated with OS, DSS, and PFS of ccRCC patients (Figure 3A). The ccRCC patients with high expression level of ANGPTL4, NETO2, and SCARB1 experienced a better prognosis, and ccRCC patients with high expression of ENO2, LOX, APOC1, and C3 genes had a significantly shorter survival time (Figure 3A). Moreover, among the 12 down-regulated genes, the expression levels of KNG1, SFRP1, CLDN8, ALDOB, ESRRG, MAL, EGF, and SCNN1A were correlated with OS, DSS, and PFS of ccRCC patients (Figure 3B). And the prognosis of ccRCC patients with high expression levels of SFRP1, CLDN8, ALDOB, ESRRG, MAL, EGF, and SCNN1A were better (Figure 3B). Subsequently, the multivariate Cox regression analyses were conducted to further identify the independent predictors of OS, DSS, and PFS in ccRCC patients. The forest plots showed that, only the expression level of ALDOB was closely related to OS (HR =0.866, 95% CI: 0.794-0.945, P=0.001), DSS (HR =0.830, 95% CI: 0.741-0.931, P=0.001), and PFS (HR =0.838, 95% CI: 0.764-0.919, P<0.001) in ccRCC

patients (*Figure 4*). And the ccRCC patients with higher expression level of ALDOB experienced a better prognosis, while those with lower expression level of ALDOB had a significantly worse prognosis. Furthermore, the prognostic value of ALDOB in ccRCC patients was also validated by multivariate Cox regression analysis in an independent cohort (E-MTAB-1980 dataset) (Figure S3). Therefore, ALDOB was identified as an independent factor for prognosis of ccRCC patients (*Figure 4*).

The expression level of ALDOB in ccRCC patients

In this analysis, the expression level of the ALDOB mRNA was investigated based on the ten GEO datasets. The results suggested that the mRNA expression level of ALDOB was significantly down-regulated in ccRCC tissues compared to that in the normal tissues (*Figure 5A*). Furthermore, the analysis of the mRNA expression level of ALDOB from TCGA-KIRC dataset indicated the similar expression patterns in the ccRCC and normal samples (Figure 5A), further validating the findings of the analysis performed using the GEO datasets. In addition to analyzing the expression level of ALDOB at mRNA level, the expression of ALDOB at the protein level was also analyzed by using the CPTAC database and the HPA database. The results show that the protein expression level of ALDOB in the 110 ccRCC samples was significantly lower than that in the 84 normal tissues from the CPTAC database (Figure 5B). Moreover, a lower protein expression level of ALDOB protein was also found in the ccRCC tissues than that in the normal tissues, as visualized by the representative immunohistochemical images from the HPA database (Figure 5C). The 3D structure of ALDOB predicted by the AlphaFold database is shown in Figure 5D. In summary, these results indicated that ALDOB is down-regulated in ccRCC at both transcriptional and translational levels.

Relationship between the expression level of ALDOB and the clinicopathological features of ccRCC patients

To determine the relationship between the expression of ALDOB and the clinicopathological features of ccRCC patients, the characteristics of the ccRCC patients from the TCGA-KIRC dataset were further analyzed. The results from the logistic regression analyses showed that the expression level of ALDOB was also closely related to the T stage, AJCC stage, and histologic grade of ccRCC patients (Table S2). Furthermore, the results showed a



Figure 2 The expression level of 24 DEGs in ccRCC patients from TCGA database. (A) Expression level of 12 up-regulated DEGs; (B) Expression level of 12 down-regulated DEGs. ***, P<0.001. DEGs, differential expressed genes; ccRCC, clear cell renal cell carcinoma; TCGA, The Cancer Genome Atlas.

A		OS			DSS			PFS	
Gene	HR (95% CI)		P value	HR (95% CI)		P value	HR (95% CI)		P value
CA9	0.619 (0.449-0.855)		0.002	0.686 (0.454 – 1.036)	i	0.054	0.828 (0.587–1.167)	H a ti	0.261
ANGPTL4	0.624 (0.441-0.885)	⊷⊷	0.003	0.571 (0.368-0.885)	⊷¦	0.004	0.716 (0.516-0.995)	H -	0.037
HILPDA	0.745 (0.529-1.050)		0.071	0.580 (0.371-0.906)	⊷ !	0.006	1.260 (0.875-1.814)	ı⊥ ●	0.187
NDUFA4L2	1.242 (0.905-1.705)	н е с	0.164	1.551 (1.037–2.319)	, 	0.022	1.677 (1.202-2.339)	i	0.001
ENO2	1.456 (1.066-1.989)	¦	0.013	1.559 (1.069–2.273)	¦⊷	0.02	1.662 (1.162-2.377)	╎┝━╋━━┥	0.002
NETO2	0.592 (0.421-0.831)	H .	0.001	0.557 (0.363-0.855)	H	0.003	0.681 (0.499-0.928)	H+	0.015
AHNAK2	0.823 (0.610-1.112)		0.213	0.863 (0.586-1.269)	Hain I	0.462	0.712 (0.503-1.008)	He i	0.077
SCARB1	0.672 (0.498-0.907)	⊨ ● → ¦	0.008	0.607 (0.415-0.886)	⊨ e ⊷¦	0.009	0.714 (0.521-0.979)	⊷ -{	0.032
LOX	1.471 (1.046-2.069)	·	→ 0.016	1.846 (1.199–2.842)	· • • • • • • • • • • • • • • • • • • •	→ 0.002	2.335 (1.617-3.371)	· • •	0.001
APOC1	1.507 (1.118-2.032)	i	• 0.01	1.698 (1.163-2.480)	· • • • • • • • • • • • • • • • • • • •	0.009	1.635 (1.195–2.237)	i	0.004
SCD	0.861 (0.612-1.212)	┝━╋┼╼┥	0.374	1.298 (0.867–1.944)	┝┼╼╾╾┥	0.185	1.327 (0.951–1.851)	┟╼╾╍	0.081
C3	1.407 (1.039-1.904)	└ ──●────	0.023	1.819 (1.168–2.833)	·	→ 0.002	1.751 (1.209-2.538)	¦ ⊷⊷⊶	0.001
В		OS			DSS			PFS	
Gene	HR (95% CI)		P value	HR (95% CI)		P value	HR (95% CI)		P value
KCNJ1	0.839 (0.623 – 1.129)	H.	0.25	0.594 (0.390-0.905)	H H H	0.033	0.643 (0.444-0.931)	i	0.009
KNG1	1.652 (1.179-2.313)	¦ ⊷	0.01	2.078 (1.357-3.182)	¦ ⊷⊷	0.006	1.569 (1.105-2.228)	· · · · •	0.025
HRG	0.677 (0.485-0.945)	He H	0.037	0.593 (0.389-0.903)	HH-	0.032	0.793 (0.570 – 1.102)	⊢ ≜ ∔4	0.186
UMOD	0.733 (0.543-0.989)	H 	0.038	0.690 (0.474-1.006)	H+-	0.052	0.748 (0.548-1.022)	H	0.074
SFRP1	0.627 (0.455-0.865)	⊨∎≕¦	0.002	0.583 (0.388-0.876)	⊷¦	0.005	0.611 (0.438-0.852)	Here (0.002
CLDN8	0.648 (0.468-0.897)	⊨•¦	0.018	0.580 (0.394-0.855)	⊨=-¦	0.012	0.651 (0.471-0.900)	Here !	0.016
FGF9	1.319 (0.976 – 1.783)	—	0.066	1.386 (0.947-2.029)		0.087	1.271 (0.928 – 1.742)		0.129
ALDOB	0.463 (0.344-0.623)	HH I	0.001	0.360 (0.235-0.552)	⊷ ¦	0.001	0.414 (0.303-0.566)	⊷ -	0.001
ESRRG	0.435 (0.311-0.607)	⊷	0.001	0.287 (0.189-0.438)	•	0.001	0.413 (0.284-0.600)	⊷	0.001
MAL	0.630 (0.454 - 0.875)	Here !	0.014	0.517 (0.352-0.760)	HHH I	0.001	0.559 (0.409-0.765)	H 1	0.001
EGF	0.720 (0.525-0.988)	⊢ •−-;	0.032	0.518 (0.351-0.766)	Here i	0.001	0.615 (0.441-0.857)	⊨e i	0.002
SCNN1A	0.694 (0.501-0.961)		0.019	0.628 (0.415-0.949)		0.016	0.721 (0.516-1.009)		0.004
		0 1 1 1 2 2 0			4 5	ò		0 6 1 0 1 6 2	0

Figure 3 Forest plots of univariate Cox regression analysis of DEGs in ccRCC patients from TCGA-KIRC dataset. (A) Forest plots of univariate Cox regression analysis of 12 up-regulated DEGs; (B) forest plots of univariate Cox regression analysis of 12 down-regulated DEGs. DEGs, differential expressed genes; ccRCC, clear cell renal cell carcinoma; OS, overall survival; DSS, disease-specific survival; PFS, progression free survival; TCGA, The Cancer Genome Atlas; KIRC, kidney renal clear cell carcinoma.

decreasing trend of ALDOB expression with an upgrade of T stage, M stage, AJCC stage, and histologic grade of ccRCC patients (*Figure 6*). And the ccRCC patients with T1 stage, metastasis, AJCC stage 1, and histologic grade 1 were found to have the higher expression level of ALDOB (*Figures 6B-6E*). To summarize, these results showed that ALDOB might be associated with tumor development and progression in ccRCC patients.

Construction and validation of nomograms

Based on the results of Cox logistic regression analyses, the prognostic predictors, including T stage, N stage, M stage, AJCC stage, histologic grade, and ALDOB, were used to establish the nomograms of 1-year, 3-year, and 5-year OS, DSS, and PFS of ccRCC patients from TCGA-KIRC dataset. These nomograms were used to provide clinically quantitative tools to predict the prognosis of the patients with ccRCC (*Figure 7A-7C*). The C-indices of the predicted OS, DSS, and PFS were 0.764 (0.737–0.790), 0.860 (0.839–0.881), and 0.823 (0.801–0.846), respectively, suggesting the excellent prognostic ability for the ALDOBbased nomograms. Moreover, the calibration plots showed good consistency between the predicted survival and observation survival of OS, DSS, and PFS (*Figures 7D-7F*), demonstrating that these nomograms performed well and were ideal models for predicting the prognosis for ccRCC patients. Furthermore, the nomogram and calibration plot of 1-year, 3-year, and 5-year OS of ccRCC patients were also established in the external cohort (E-MTAB-1980 dataset), which validated the good prognostic ability for the ALDOB-based model (Figure S3). Overall, ALDOB might be a novel prognostic biomarker for predicting the prognosis of ccRCC patients.

Functional enrichment analysis and protein-protein interaction network

Firstly, the correlation analysis was performed to explore the potential related genes that might interact with ALDOB.

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Figure 4 Forest plots of univariate and multivariate Cox regression analysis of factors affecting the survival of ccRCC patients from TCGA-KIRC dataset. (A) OS; (B) DSS; (C) PFS. ccRCC, clear cell renal cell carcinoma; OS, overall survival; DSS, disease-specific survival; PFS, progression free survival; TCGA, The Cancer Genome Atlas; KIRC, kidney renal clear cell carcinoma; TNM, tumor node metastasis; AJCC, American Joint Committee on Cancer.

The correlation circle plot and heatmap showed that the top ten positively correlated genes that interacted with ALDOB were *G6PC*, *CYP4A11*, *FUT6*, *PKLR*, *SLC22A6*, *AGMAT*, *CYP4A22*, *SLC22A7*, *AKR7A3*, and *SLC22A12* (*Figure 8A,8B*). Then, the GO and KEGG analysis pathway analysis of ALDOB and its correlated genes was carried

out to reveal the potential functions of ALDOB in ccRCC. The results showed that in ccRCC, ALDOB was mainly involved and participated in the small molecule catabolic process (Biological Process), apical part of cell (Cellular Component), organic anion transmembrane transporter activity (Molecular Function), and PPAR signaling pathway

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Figure 5 The mRNA and protein expression level of ALDOB in ccRCC patients. (A) ALDOB mRNA expression between normal tissues and tumor tissues from ten GEO datasets and TCGA-KIRC dataset; (B) ALDOB protein expression between normal tissues (n=84) and tumor tissues (n=110) from the CPTAC dataset; (C) Immunohistochemical staining of ALDOB expression in ccRCC tissue and normal tissue from HPA database; (D) 3D structure of ALDOB (AlphaFold database). ***, P<0.001; **, P<0.01; *, P<0.05. ALDOB, aldolase B; ccRCC, clear cell renal cell carcinoma; GEO, Gene Expression Omnibus; TCGA, The Cancer Genome Atlas; KIRC, kidney renal clear cell carcinoma; CPTAC, Clinical Proteomic Tumor Analysis Consortium; HPA, Human Protein Atlas.

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Figure 6 Association of ALDOB expression and clinicopathological features of ccRCC patients from TCGA-KIRC dataset. (A) Heatmap of the expression level of ALDOB and the clinicopathological features of ccRCC patients; (B) association of ALDOB expression and tumor (T) stage of ccRCC patients; (C) association of ALDOB expression and metastasis (M) stage of ccRCC patients; (D) association of ALDOB expression and AJCC stage of ccRCC patients; (E) association of ALDOB expression and histologic grade of ccRCC patients. ***, P<0.001, **, P<0.01, *, P<0.05. ALDOB, aldolase B; ccRCC, clear cell renal cell carcinoma; AJCC, American Joint Committee on Cancer; TCGA, The Cancer Genome Atlas; KIRC, kidney renal clear cell carcinoma; N, node.

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Figure 7 Nomograms and calibration plots for the prediction of the prognosis in ccRCC patients from TCGA-KIRC dataset. (A) Nomogram for the prediction of OS in ccRCC patients; (B) Nomogram for the prediction of DSS in ccRCC patients; (C) Nomogram for the prediction of PFS in ccRCC patients; (D) calibration plot of the nomogram for prediction of OS in ccRCC patients; (E) calibration plot of the nomogram for prediction of PFS in ccRCC patients; (F) calibration plot of the nomogram for prediction of PFS in ccRCC patients. ccRCC, clear cell renal cell carcinoma; OS, overall survival; DSS, disease-specific survival; PFS, progression free survival; TCGA, The Cancer Genome Atlas; KIRC, kidney renal clear cell carcinoma; TNM, tumor node metastasis.

(KEGG Pathway) (*Figure 8C*). Finally, by the order of the predicted scores, a PPI network of ALDOB was built based on the STRING database. And the result showed that the predicted interactive partners of ALDOB were TPI1, FBP1, PFKL, FBP2, and PFKP (*Figure 8D*).

Immune infiltration analysis of ALDOB

Based on the "MCP-Counter" algorithm, this study firstly analyzed the infiltration abundance of eight types of immune cells and two types of stromal cells in ccRCC patients from TCGA-KIRC dataset. We observed that the expression level of ALDOB was correlated with the abundance of most of immune cells and stromal cells (*Figure 9A*). We next calculated the immune score, stromal score, estimate score, and tumor purity of ccRCC patients. The results showed that the immune score and the estimate score were higher in the ALDOB-low cohort than in the ALDOB-high cohort (*Figure 9B*). However, the tumor purity showed the opposite difference (*Figure 9B*). Therefore, the above results showed the significant correlation between ALDOB and the immune infiltration in ccRCC. In order to determine whether ALDOB and ten types of immune cells and stromal cells could distinguish ccRCC patients, we next performed consensus clustering analysis. The results showed that



Figure 8 GO and KEGG pathway enrichment analysis and protein-protein interaction network of ALDOB in ccRCC patients. (A) Correlation circle plot for the top ten positively correlated genes with ALDOB in ccRCC from TCGA-KIRC dataset; (B) correlation heatmap for the top ten positively correlated genes with ALDOB in ccRCC from TCGA-KIRC dataset; (C) GO and KEGG pathway enrichment analysis of ALDOB and its correlated genes in ccRCC; (D) protein-protein interaction network of ALDOB. ***, P<0.001. GO, Gene Ontology; KEGG, Kyoto Encyclopedia of Genes and Genomes; ALDOB, aldolase B; ccRCC, clear cell renal cell carcinoma; TCGA, The Cancer Genome Atlas; KIRC, kidney renal clear cell carcinoma; BP, biological process; CC, cellular component; MF, molecular function.

ALDOB and immune cells could classify the ccRCC patients from TCGA-KIRC dataset into two distinct clusters (*Figure 10A-10C*). And the heatmap demonstrated the specific expression level of ALDOB and the infiltration abundance of immune cells and stromal cells among two different clusters (*Figure 10D*). We also observed that

the ccRCC patients in the cluster 1 were more prone to experience a shorter OS than the patients in the cluster 2 (HR =0.490, 95% CI: 0.360–0.660, P<0.001) (*Figure 10E*). Moreover, the B lineage, fibroblasts, estimate scores, and immune scores were elevated in the cluster 1, whereas the ccRCC patients in the cluster 2 had significantly



Figure 9 Immune infiltration analysis of ALDOB in ccRCC. (A) Correlation heatmap of the expression level of ALDOB with the abundance of immune cells and stromal cells in ccRCC patients from TCGA-KIRC dataset; (B) comparison between ALDOB-Low group and ALDOB-High group for tumor microenvironment scores in ccRCC. **, P<0.01, *, P<0.05. ALDOB, aldolase B; ccRCC, clear cell renal cell carcinoma; TCGA, The Cancer Genome Atlas; KIRC, kidney renal clear cell carcinoma; NK, natural killer.

higher levels of the natural killer (NK) cells, neutrophils, endothelial cells, and tumor purity scores (*Figure 10F,10G*). Taken together, these results confirmed that ALDOB strongly affects the immune infiltration in ccRCC.

m6A methylation analysis of ALDOB

The correlation between the expression levels of ALDOB and 23 m6A regulators was first investigated. The correlation heatmap showed that the expression level of



Figure 10 Molecular subtypes analysis of ccRCC patients from TCGA-KIRC dataset based on ALDOB and immune cells and stromal cells. (A-C) Consistency clustering plots showing distinct two subtypes of ccRCC patients; (D) heatmap of the infiltration abundance of eight types of immune cells (T cells, CD8 T cells, cytotoxic lymphocytes, B lineage cells, natural killer cells, monocyte lineage, myeloid dendritic cells, and neutrophils) and two types of stromal cells (endothelial cells and fibroblasts) in ccRCC patients; (E) Kaplan-Meier survival curve of two distinct clusters and OS; (F) comparison between two clusters for eight types of immune cells and two types of stromal cells; (G) comparison between two clusters for tumor microenvironment scores. ccRCC, clear cell renal cell carcinoma; ALDOB, aldolase B; TCGA, The Cancer Genome Atlas; OS, overall survival; KIRC, kidney renal clear cell carcinoma; CDF, cumulative distribution function; NK, natural killer.

ALDOB was significantly correlated with the expression levels of METTL14 (r=0.25, P<0.001), ALKBH5 (r=0.24, P<0.001), YTHDC1 (r=0.22, P<0.001), RBM15B (r=0.21, P<0.001), LRPPRC (r=0.20, P<0.001), IGF2BP2 (r=-0.31, P<0.001), and IGF2BP3 (r=-0.29, P<0.001) (Figure 11A). Then, the LASSO regression analysis with 10-fold crossvalidation was carried out to identify the prognostic factors that were significantly correlated with the prognosis of ccRCC patients based on ALDOB and 23 m6A regulators (Figure 11B,11C). As a result, a total of ten genes, including HNRNPA2B1, IGF2BP3, METTL3, IGF2BP2, WTAP, IGF2BP1, ALDOB, ZC3H13, METTL14, and LRPPRC, were identified and enrolled through the LASSO regression analysis. Therefore, this study established the prognostic model and calculated the risk scores for each ccRCC patient based on these ten genes (Figure 11D). The risk factor diagram showed the difference in the distribution of the expression level of ten genes in the high-risk cohorts and low-risk cohorts (Figure 11E). Subsequently, the results of the survival analysis indicated that, compared to the ccRCC patients in high-risk groups, the ccRCC patients in lowrisk groups experienced a better OS (HR =3.750, 95% CI: 2.780-5.070, P<0.001) (Figure 11F). Finally, the AUC value calculated based on the time-dependent ROC curve was high, indicating the prognostic accuracy of the model that this study constructed (Figure 11G).

Discussion

For localised ccRCC patients, surgery is still the only curative treatment (4). However, due to 20–30% of ccRCC patients experience tumor recurrence and metastasis after surgery, there is an urgent need to diagnose, monitor, and manage this cancer (29,30). Currently, apart from imaging investigations, the reliable and effective molecular biomarkers for ccRCC that suitable for clinical application have not yet been identified, and the effective risk stratification and screening systems have not been established. The lack of early diagnostic tools has added much pressure and difficulties to the early detection and early treatment of ccRCC. Consequently, development of effective molecular biomarkers is essential to improve the clinical management and reduce the mortality of ccRCC patients.

Initially, this study was devoted to screen and identify the DEGs in ccRCC. Through the intersection of DEGs from ten GEO datasets, a total of 24 DEGs was determined, including twelve up-regulated genes and twelve down-

regulated genes. Besides, the expression levels of the 24 DEGs in ccRCC tissues and normal kidney tissues were verified using the TCGA-KIRC datasets. Subsequently, the results of survival analysis showed that the expression levels of 15 genes were markedly correlated with OS, DSS, and PFS of ccRCC patients. On the basis of univariate Cox regression analyses, the results of multivariate Cox regression analysis showed that, among the 24 DEGs, only the expression level of ALDOB was an independent predictor of OS, DSS, and PFS of ccRCC patients. Therefore, ALDOB was identified as a pivotal gene in ccRCC.

Metabolic patterns of cellular processes have been reported to be associated with a variety of tumorigenesis (31). Recent developments in the field of cancers have led to an interest in the changes in the metabolic pathways that control tumor energy and biosynthesis, known as metabolic reprogramming, which was considered to be a core hallmark of cancer (32,33). Moreover, the alterations of the known renal cell carcinoma genes, such as VHL, is closely related to the reprogramming of metabolic pathways (34). A number of researchers have reported that the development of ccRCC is usually accompanied by the metabolic reprogramming of glycolysis, fatty acid metabolism, and the tricarboxylic acid cycle (35-37). Therefore, glycolysis plays a vital role in the metabolic network. The progress of glycolysis usually consists of ten main steps, involving the participation of multiple enzymes (38). The members of the aldolase family are the fourth enzyme in the progress of glycolysis, that consists of ALDOB, ALDOB, and ALDOC, encoding by three different genes (7). Aldolase B, also known as fructose bisphosphate aldolase B, is encoded by the ALDOB gene and is mainly expressed in the liver and kidney tissues (7). In vivo, aldolase B catalyzes the conversion of fructose 1-phosphate to dihydroxyacetone phosphate and glyceraldehyde (39). It has been demonstrated that ALDOB not only plays an important role in glucose and fructose metabolism, but is also differentially expressed in a variety of malignancies, such as rectal adenocarcinoma, colorectal cancer, and gastric cancer (8-10). However, few studies have been able to draw on any systematic research into the role of ALDOB in ccRCC patients. Therefore, the aim of this study was to clarify the expression level, prognostic value, functional enrichment, immune infiltration, and m6A modification of ALDOB in ccRCC patients. In this study, we first observed that the mRNA and protein expression level of ALDOB was down-regulated in ccRCC patients. Furthermore, the results showed a decreasing trend of



Figure 11 m6A methylation analysis of ALDOB in ccRCC. (A) Correlation heatmap of ALDOB and 23 m6A regulators in ccRCC patients from TCGA-KIRC dataset. (B,C) The LASSO coefficients profiles of ALDOB and 23 m6A regulators and LASSO regression with 10-fold cross-validation obtained ten prognostic genes using minimum lambda value. (D) Coefficients of selected genes and the risk score formula. (E) The distribution of the risk score and the survival status of each ccRCC patient. (F) Kaplan-Meier survival curve of the ccRCC patients in high-risk and low-risk groups (OS). (G) Time-dependent ROC curve of the prognostic model. ALDOB, aldolase B; ccRCC, clear cell renal cell carcinoma; LASSO, least absolute shrinkage and selection operator; TCGA, The Cancer Genome Atlas; OS, overall survival; ROC, receiver operating characteristic; KIRC, kidney renal clear cell carcinoma; TPR, true positive rate; FPR, false positive rate.

ALDOB expression with an upgrade of T stage, M stage, AJCC stage, and histologic grade of ccRCC patients, suggesting that ALDOB was strongly correlated with the clinicopathological factors of ccRCC patients and might strongly influence tumor development and progression in ccRCC.

With continuous researches, several studies evaluated the prognostic value of ALDOB in a variety of malignancies and delved into the molecular mechanisms of ALDOB in the development of cancers. In a prostate cancer study, Xia et al. showed that the ALDOB, as an independent risk factor for metastasis-free survival of prostate cancer patients, was significantly associated with the pathological grade and tumor stage of patients with prostate cancer (40). And the prostate cancer patients in high-risk group exhibited therapeutic resistance and immunosuppression. Further experiments in vitro showed that the expression of ALDOB increased significantly under the hypoxic conditions and promoted the proliferation and invasion ability of prostate cancer cells (40). Several studies have confirmed that ALDOB could also regulate the progression of malignancies by participating in various important signaling pathways, including PI3K/AKT/mTOR signaling pathway, GSK-3ß signaling pathway, and Wnt signaling pathway (41-43). In addition, it was shown that the expression level of ALDOB was up-regulated in the liver metastasis tumors of colon cancer, providing sufficient energy supply to tumor cells by enhancing fructose metabolism (44). And the loss of ALDOB decreased the proliferative rates of tumor cells in the liver. Interestingly, this study also showed that reducing dietary fructose significantly reduces liver metastatic growth (44). Also, a research by Li et al. pointed out that ALDOB could suppress hepatocellular carcinogenesis by directly binding and inhibiting the activity of glucose-6phosphate dehydrogenase, the rate-limiting enzyme in the pentose phosphate pathway, revealing a new mode of metabolic reprogramming in hepatocellular carcinogenesis due to the loss of ALDOB (45). Based on the above research background, this study further discussed the prognostic value of ALDOB in ccRCC patients. The results of multivariate Cox regression analysis showed that among the 24 DEGs, only ALDOB was an independent prognostic factor for OS, DSS, and PFS in ccRCC patients, and the ccRCC patients with higher expression level of ALDOB experienced a better prognosis, partly being attributed to the fact that the ccRCC patients with higher expression level of ALDOB had lower T stage, N stage, M stage, and histologic grade. Previously, some scholars

also explored the mechanism of ALDOB in ccRCC. Wang et al. found that the accumulation of fructose 1, 6-bisphosphate in cancer cells by the down-regulation of ALDOB maintained the redox homeostasis and promoted ccRCC growth (46). What's more, in a genetic assessment on frozen tissue samples of metastatic ccRCC, Nouhaud et al. used quantitative multiplex polymerase chain reaction of short fluorescent fragment method to detect the gene copy number variations on the 14 genes including ALDOB (47). The results showed that the most frequent losses were located on ALDOB, and the loss of ALDOB was associated with higher histologic grade and worse prognosis of metastatic ccRCC patients (47). In summary, ALDOB could play an important role in the development of ccRCC and influence the prognosis of ccRCC patients through multiple pathways and mechanisms.

Presently, the prognosis of ccRCC patients treated with radical nephrectomy or partial nephrectomy always depends on the anatomical, histological, clinical, and molecular factors (4). Among them, anatomical factors are reflected in the TNM classification, providing the most reliable information. And histological factors include RCC subtype, tumor grade, sarcomatoid features, vascular invasion, tumor necrosis, invasion of the collecting system, and perirenal fat. For the patients with localized and locally advanced ccRCC, the commonly used prognostic evaluation systems are the University of California, Los Angeles, Integrated Staging System (UISS) and Stage, Size, Grade, and Necrosis (SSIGN) scores (48,49), while for advanced or metastatic ccRCC, the prognostic risk models are the International Metastatic RCC Database Consortium (IMDC) and the Memorial Sloan Kettering Cancer Center (MSKCC) scoring systems (50). The results of this study showed that ALDOB was an independent prognostic factor in the prognosis of ccRCC patients. Therefore, in this study, the nomograms with more prognostic predictive power were developed to predict the OS, DSS, and PFS of ccRCC patients by integrating T stage, N stage, M stage, histologic grade, and the expression level of ALDOB of ccRCC patients. Based on these nomograms, it would be helpful to more accurately assess the clinical outcome of ccRCC and provide more personalized prognostic assessment strategy for ccRCC patients.

Subsequently, to determine the potential functions and interactive partners of ALDOB in ccRCC, this study first analyzed the potential related genes that might interact with ALDOB in ccRCC patients. Next, the GO and KEGG analysis of ALDOB and its related genes were performed.

The results showed that ALDOB and its related genes are mainly involved in the metabolism and metabolic pathways of multiple substances, including small molecule catabolic process, glycolysis, gluconeogenesis, and fatty acid degradation. Therefore, this study confirmed that the aberrant expression of ALDOB is potentially involved in the metabolic reprogramming in ccRCC patients and is closely associated with reprogramming of diverse metabolic pathways and metabolic networks in ccRCC. Besides, the results of PPI network analysis suggested that the predicted functional partners of ALDOB were TPI1, FBP1, PFKL, FBP2, and PFKP. TPI1 gene codes triosephosphate isomerase 1, that catalyzes the interconversion between dihydroxyacetone phosphate and D-glyceraldehyde-3phosphate in glycolysis and gluconeogenesis (51). FBP1 gene codes fructose-bisphosphatase 1, that catalyzes the hydrolysis of fructose 1,6-bisphosphate to fructose 6-phosphate in the presence of divalent cations, acting as a rate-limiting enzyme in gluconeogenesis (52). PFKL gene codes phosphofructokinase, that catalyzes the phosphorylation of D-fructose 6-phosphate to fructose 1,6-bisphosphate by ATP, which is the first committing step of glycolysis (53). FBP2 gene codes fructosebisphosphatase 2, that catalyzes the hydrolysis of fructose 1,6-bisphosphate to fructose 6-phosphate in the presence of divalent cations, and probably participates in glycogen synthesis from carbohydrate precursors, such as lactate (54). PFKP gene codes phosphofructokinase, that catalyzes the phosphorylation of D-fructose 6-phosphate to fructose 1,6-bisphosphate by ATP, which is the first committing step of glycolysis (55). Therefore, ALDOB and its functional partners are mainly involved in a variety of pathways such as fructose metabolism and glycolysis. Previous studies have revealed that the metabolic reprogramming in ccRCC is mainly focused on the glucose metabolism, tricarboxylic acid cycle, and fatty acid oxidation and synthesis (35-37). The findings of this study implied that the disorders of fructose metabolism might also exist in ccRCC and ALDOB-mediated fructose metabolism might drive metabolic reprogramming in ccRCC, which provides a potential direction for further research.

The TME refers to the cellular environment in which the tumor exists (56). Numerous studies have confirmed that TME plays an important role in tumor development, adaptation, and pathogenesis (57-59). Therefore, understanding how to regulate the balance of TME within and between different tumor-supporting and tumorsuppressing infiltrating immune cells will hopefully lead to potentially more effective therapy for patients with cancers. In addition to tumor cells, the TME also includes immune cells, stromal cells, blood vessels, extracellular matrix, and other signaling molecules (60). Accordingly, in this study, we elucidated the infiltration abundance of immune cells and stromal cells in ccRCC patients and determined the correlation between ALDOB and these cells. The result indicated that the expression level of ALDOB was correlated with the abundance of most immune cells and stromal cells. In terms of immune scores, we observed ccRCC patients in the ALDOB-low expression group had elevated immune scores than those in ALDOB-high expression group. These results implicated that ALDOB might regulate the infiltrating abundance of immune cells and stromal cells in TME, thus exerting a potential influence on the prognosis of ccRCC patients. To confirm this hypothesis, the consensus clustering analysis was further performed based on the expression level of ALDOB and the abundance of immune cells and stromal cells to subtype the ccRCC patients. The results indicated that the ccRCC patients in the cluster 1 had the higher abundance of fibroblasts. In the physiological conditions, the primary function of fibroblasts is to support tissue integrity and participate in the wound healing (57). However, in the cancerous conditions, dysregulated cancer associated fibroblasts (CAFs) could promote the inflammatory response by producing cytokines and chemokines to recruit immune cells (57). On the other hand, CAFs could inhibit the antitumor activity of antigenpresenting cells by down-regulating the co-stimulatory molecules in dendritic cells (DCs) and stimulating the differentiation of immature DCs to regulatory DCs with immunosuppressive activity (57). Thus, CAFs could directly or indirectly contribute to the tumor development by driving immune suppression. Therefore, the ccRCC patients in cluster 1 were more prone to poor prognosis, partly attributed to the higher abundance of fibroblasts. In contrast, we observed a higher abundance of NK cells in the cluster 2 patients. As a central population of innate lymphoid cells, NK cells play a crucial role in triggering host immune responses against the tumor growth, and the immunosurveillance of solid tumors (61). For many patients with solid tumors, the high level of tumor infiltrating NK cells in TME predicts a good prognosis (61). Similarly, the findings of this study revealed that the ccRCC patients in the cluster 2 experienced a better prognosis, possibly due to the higher abundance of NK cells in these patients. Taken together, this study suggested that ALDOB might involve in regulating the infiltration abundance of immune cells and stromal cells to change the immune components of the TME, which in turn affects the prognosis of ccRCC patients.

m6A modification is the most prevalent, abundant, and conserved post-transcriptional modification to control the gene expression in the eukaryotic cells (62). As one of the most common reversibly epigenetic modification, m6A modification is widespread on mRNA and noncoding RNAs, that having important effects on the stability, localization, translation, splicing, and transport of RNAs (63). Currently, numerous studies have shown that the abnormalities of m6A modification are found in the various malignancies, and are strongly associated with the progression, metastasis, prognosis, and drug resistance of patients with tumors (63-65). Generally, there are three types of proteins that modulate m6A modifications, namely writers, erasers, and readers. In some types of cancers, these three types of m6A regulatory proteins are commonly dysregulated and could involve in different signaling pathways to influence the progression and prognosis of cancer patients (65). Therefore, in this study, the correlation between ALDOB and 23 m6A regulators was first investigated. The results showed that ALDOB was closely associated with multiple m6A regulators. Then, a risk score formula based on the ALDOB and nine m6A regulators was constructed after the LASSO regression analysis. The results showed that the models we constructed could effectively predict the prognosis of ccRCC patients, and ccRCC patients in the high-risk group were more prone to poorer OS than those with lower risk scores. In summary, this study revealed the potential correlation between ALDOB and m6A modifications, and highlighted the importance of the prognostic impact of ALDOB and m6A modulators on ccRCC patients.

Although this study aimed to undertake a comprehensive analysis of the expression level, prognostic value, functional enrichment, immune infiltration, and m6A modification of ALDOB in ccRCC, it still inevitably comes with limitations. First, this study included as many datasets as possible to fully demonstrate the expression pattern of ALDOB. However, further studies based on the prospective clinical studies and vitro cellular experiments are needed to validate the expression level and explore the specific mechanism of ALDOB in ccRCC. Second, this study investigated the potential correlation between ALDOB and TME and m6A modification. However, these hypotheses are required to be confirmed by the basic experiments to clarify the possible regulatory mechanisms. Finally, despite the fact that this study included samples from 1,479 tissues from ccRCC patients, the conclusions of this study were based on bioinformatic analysis and retrospective publicly available databases, making the conclusions less convincing. Therefore, more genomic and metabolomics studies are needed to further confirm the role of ALDOB in fructose metabolism and metabolic reprogramming in ccRCC. It would be the goal and direction of our future research.

Conclusions

Taken together, the findings of this study showed that the down-regulated ALDOB in ccRCC patients was closely associated with the clinicopathological features, poor prognosis, and TME in patients with ccRCC. Therefore, ALDOB might be a novel prognostic biomarker and therapeutic target for ccRCC patients. This study not only revealed that ALDOB-mediated fructose metabolism might drive metabolic reprogramming in ccRCC, but also suggested a new perspective for the precision medicine in ccRCC patients.

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Footnote

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Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013).

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Supplementary

Table S1 The number of the ccRCC tissues and normal kidney tissues	in GEO datasets enrolled in this study
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Dataset	ccRCC tissues (n)	Normal tissues (n)	Total tissues (n)	Platform
GSE53757	72	72	144	GPL570
GSE53000	53	6	59	GPL6244
GSE36895	29	23	52	GPL570
GSE15641	32	23	55	GPL96
GSE66272	27	27	54	GPL570
GSE68417	29	14	43	GPL6244
GSE40435	101	101	202	GPL10558
GSE12606	3	3	6	GPL570
GSE46699	67	63	130	GPL570
GSE11151	26	5	31	GPL570
Total	439	337	776	-



Figure S1 Volcano plots of DEGs between ccRCC tissues and normal kidney tissues in each GEO dataset. (A) GSE53757; (B) GSE53000; (C) GSE36895; (D) GSE15641; (E) GSE66272; (F) GSE68417; (G) GSE40435; (H) GSE12606; (I) GSE46699; (J) GSE11151.



Figure S2 Upset plots of common DEGs between ccRCC tissues and normal kidney tissues of ten GEO datasets. (A) Common upregulated DEGs between ccRCC tissues and normal kidney tissues; (B) common down-regulated DEGs between ccRCC tissues and normal kidney tissues.



D	Univariate Cox regression			Multivariate Cox regression			
Characteristics	HR (95% CI)		P value	HR (95% CI)		P value	
Gender							
Male vs. Female	2.366 (0.703 - 7.967)		0.164				
Age		I I			1		
>60 vs. <=60 years old	2.703 (1.062 - 6.878)	⊢ •−−1	0.037	2.020 (0.746 - 5.471)	k ∎i	0.167	
T Stage		ļ					
T3+T4 vs. T1+T2	5.223 (2.268 - 12.027)	╎⊷╸───	< 0.001	1.773 (0.731 - 4.303)	, *•1	0.205	
N Stage							
N1 vs. N0	18.917 (7.173 - 49.892)	!	 < 0.001	5.798 (1.899 - 17.702)	¦•	→ 0.002	
M Stage		i			i		
M1 vs. M0	6.245 (2.626 - 14.849)	¦ ⊷•	< 0.001	2.343 (0.942 - 5.826)	⊢ ∎I	0.067	
Histology Grade		!			1		
G3+G4 vs. G1+G2	3.698 (1.628 - 8.399)	, 	0.002	1.550 (0.605 - 3.968)	, H O	0.361	
ALDOB		I I					
The level of ALDOB	0.153 (0.063 - 0.374)	•	< 0.001	0.343 (0.121 - 0.970)	•	0.044	
		0 5 10 15	20		6 5	10 15	

Figure S3 External validation of the prognostic value of ALDOB in the E-MTAB-1980 dataset from the ArrayExpress database (OS). (A) The Kaplan-Meier survival curve comparing the high and low expression of ALDOB in OS of ccRCC patients; (B) Nomogram for the prediction of the OS in ccRCC patients; (C) calibration plot for the prediction of the OS in ccRCC patients; (D) forest plots of univariate and multivariate Cox regression analysis of factors affecting the OS of ccRCC patients. TNM, tumor node metastasis.

Clinicopathological features	Low expression of ALDOB (n=265)	High expression of ALDOB (n=265)	Р
Age (years), mean ± SD	60.55±11.92	60.57±12.38	0.986
Gender, n (%)			0.036
Female	81 (15.3)	105 (19.8)	
Male	184 (34.7)	160 (30.2)	
T stage, n (%)			0.002
T1	116 (21.9)	155 (29.2)	
T2	42 (7.9)	27 (5.1)	
Т3	98 (18.5)	81 (15.3)	
Τ4	9 (1.7)	2 (0.4)	
N stage, n (%)			0.141
NO	126 (49.4)	113 (44.3)	
N1	12 (4.7)	4 (1.6)	
M stage, n (%)			0.109
M0	204 (40.8)	218 (43.6)	
M1	46 (9.2)	32 (6.4)	
AJCC stage, n (%)			0.005
Stage I	113 (21.3)	152 (28.7)	
Stage II	35 (6.6)	22 (4.2)	
Stage III	67 (12.6)	58 (10.9)	
Stage IV	50 (9.4)	33 (6.2)	
Histologic grade, n (%)			<0.001
G1	4 (0.8)	10 (1.9)	
G2	101 (19.3)	126 (24.1)	
G3	102 (19.5)	104 (19.9)	
G4	53 (10.2)	22 (4.2)	

Table S2 The association between the mRNA expression of ALDOB and the clinicopathological features of ccRCC patients from 7	ГСGA-KIRC
dataset	

TCGA, The Cancer Genome Atlas; KIRC, kidney renal clear cell carcinoma; T, tumor; N, node; M, metastasis; AJCC, American Joint Committee on Cancer.