



Diffusion along the perivascular space as a potential biomarker for glioma grading and isocitrate dehydrogenase 1 mutation status prediction

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Background: The diffusion tensor image analysis along the perivascular space (DTI-ALPS) may have the potential to reflect glymphatic dysfunction in patients with glioma. The study aimed to determine the correlation of DTI-ALPS with glioma grade and isocitrate dehydrogenase 1 (*IDH1*) genotype and to then compare the ALPS index with other diffusion metrics.

Methods: In this study, 81 patients with glioma and 31 healthy controls underwent magnetic resonance imaging (MRI) examination. The ALPS-index, fractional anisotropy (FA), mean diffusivity (MD), and mean kurtosis (MK) were calculated. Comparisons were made between the left and right hemispheres and between patients and controls. *IDH1* status was compared after age adjustment. The diagnostic performance of each metric was assessed via receiver operating characteristic (ROC) analysis.

Results: In patients with glioma, the ALPS-index of the hemisphere ipsilateral to glioma was significantly lower than that of the hemisphere contralateral to glioma (1.417 ± 0.177 vs. 1.478 ± 0.165 ; $P=0.002$), and the bilateral ALPS-index values in patients were significantly decreased compared with those in healthy controls. The ALPS-index was significantly higher in lower-grade gliomas (LrGGs) than that in glioblastomas (GBMs) (1.495 ± 0.151 vs. 1.320 ± 0.159 ; $P<0.001$) and was significantly lower in *IDH1*-wild-type LrGGs than in *IDH1*-mutant LrGGs (1.400 ± 0.185 vs. 1.530 ± 0.123 ; $P=0.036$). FA, MD, and MK also showed significant differences between LrGGs and GBMs and between *IDH1*-mutant and *IDH1*-wild-type LrGGs ($P<0.05$). Furthermore, the combination of the ALPS-index with FA, MD, or MK, exhibited superior discrimination ability compared to each metric used alone. The ALPS-index combined with MD had the highest area under the curve (AUC) of 0.854 as compared to that of 0.614–0.807 for a single metric in glioma grading, while for *IDH1* mutation prediction, this combination had the highest AUC of 0.861 as compared to that of 0.707–0.778 for a single metric.

Conclusions: The reduced ALPS-index may reflect tumor-induced glymphatic system impairment, and the ALPS-index may be able to complement conventional diffusion metrics in glioma grading and *IDH1* genotyping.

Keywords: Glioma; diffusion magnetic resonance imaging (diffusion MRI); glymphatic system; diffusion tensor imaging

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Introduction

Gliomas are the most common primary tumors of the central nervous system (1) and have been traditionally classified into grades I–IV according to the World Health Organization (WHO) based on histopathologic features (2). Since 2016, the central nervous system tumor classification added the isocitrate dehydrogenase 1 (*IDH1*) genotype for classification (2). Numerous previous studies have shown that gliomas with the *IDH1* mutation have a better prognosis than do gliomas with the wild-type *IDH1* gene (3,4). Thus, accurate glioma grading and genotype prediction before surgery are necessary for the treatment and prognosis of gliomas.

The glymphatic system is a waste drainage system in the brain which functions via aquaporin-4 (AQP4) water channels in astrocytic end-feet (5). A few recent studies have examined the relationship between the function of the glymphatic system and gliomas (6–8). In rodent models of glioma, the outflow of cerebrospinal fluid was found to be reduced, with the dorsal meningeal lymphatic vessels being extensively remodeled (6,7). In addition, impaired glymphatic function potentially allows for the accumulation of toxic proteins, cytokines, and chemokines, which may further promote tumor growth (9,10). As gliomas exhibit varying degrees of malignancy and aggressiveness, they may in turn cause varying degrees of damage and repair to the glymphatic system. Accordingly, we hypothesized that there may be differences in glymphatic function across grades and across *IDH1* mutation statuses.

Diffusion tensor image analysis along the perivascular space (DTI-ALPS) method has been recently proposed for noninvasively detecting glymphatic system function alteration (11). The perivascular space is an important part of the glymphatic pathway, and its diffusivity in the direction of the in the periventricular white matter can indirectly indicate the state of glymphatic function. Thus, the ALPS-index has been applied as a potential biomarker for glymphatic system function in recent studies (12–14). The ALPS-index is a metric calculated at the level of the lateral ventricle body. It is based on the fact that the perivascular space lies largely orthogonal to the projection and association fibers in this area. In addition, DTI and diffusion kurtosis imaging (DKI) have been widely used for noninvasive glioma grading and identification of *IDH1* mutation status (15–18). Most conventional diffusion metrics are measured in the tumor parenchyma, but the ALPS-index is measured in the peritumor areas, which

may provide additional pathophysiological information complementary to these metrics for glioma diagnosis and prognosis.

Therefore, the purpose of this study was to comprehensively evaluate the glymphatic dysfunction in patients with glioma using the DTI-ALPS method, evaluate the role of ALPS-index in glioma grading and *IDH1* mutation status prediction, and compare the diagnostic efficacy of ALPS-index with conventional DTI- and DKI-derived metrics.

Methods

Participants

This retrospective study was conducted in accordance with the Declaration of Helsinki (as revised in 2013) and approved by the Institutional Review Board of Tongji Hospital of Tongji Medical College of Huazhong University of Science and Technology (No. TJ-IRB202303157). Written informed consent was obtained from all patients.

The image data of consecutive patients admitted to our hospital between March 2017 to July 2021 were retrospectively analyzed. The inclusion criteria were as follows: (I) histopathologically confirmed glioma; (II) patients who underwent preoperative DTI examination; and (III) no chemotherapy or radiotherapy before examination. The exclusion criteria were as follows: (I) magnetic resonance imaging (MRI) images with artifacts or poor quality; (II) unknown *IDH1* mutation status; (III) lesions extending across the midline and bilaterally occupying the cerebral hemispheres; and (IV) purely cystic glioma or glioma with severe destruction of the lateral ventricular structures. Ultimately, 81 patients and 31 healthy controls were recruited for analysis and matched in terms of age and gender. The screening process for participants is shown in *Figure 1*.

Data acquisition

All images were acquired from a 3.0 T MR scanner (Discovery MR750, GE HealthCare, Chicago, IL, USA) with a 32-channel head coil. The conventional MRI sequence included T1 fluid-attenuated inversion recovery (T1-FLAIR), T2 fast spin echo (T2-FSE), and T2-FLAIR and contrast-enhanced T1 (CE-T1). Diffusion MRI was performed under the following parameters: $b=0, 1,250, \text{ and } 2,500 \text{ s/mm}^2$; 25 gradient directions; repetition time =6,500

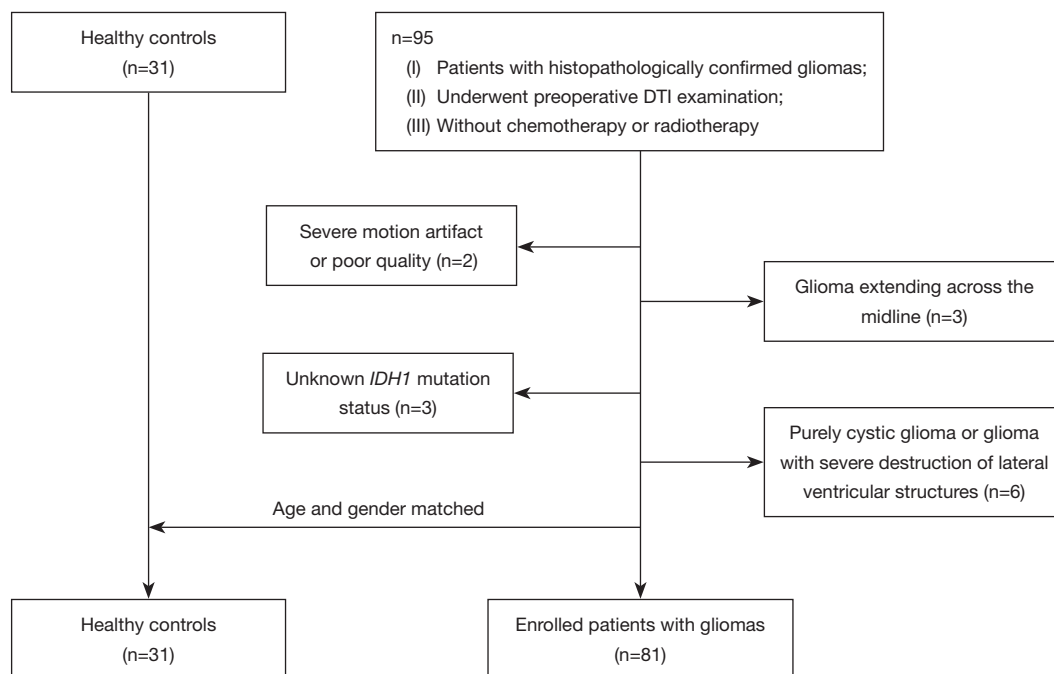


Figure 1 The screening process for participants. DTI, diffusion tensor imaging; *IDH1*, isocitrate dehydrogenase 1.

ms; echo time =85 ms; number of excitations =1, field of view =24×24 cm²; matrix size =128×128; and slice thickness =3 mm without slice spacing.

Data processing and analysis

Diffusion image preprocessing was performed using the TractoFlow pipeline. The diffusion imaging data were first denoised using Marchenko-Pastur principal component analysis (MP-PCA) (19) and corrected for Gibbs ringing artifacts using Mrtrix3 (<https://www.mrtrix.org>). Subsequently, correction for motion and eddy current artifacts, along with skull stripping, was performed using the FMRIB Software Library (FSL) (version 6.0.1, University of Oxford, Oxford, UK). N4 correction was performed with $b=0$ mm²/s using the Advanced Normalization Tools (ANTs) command (N4BiasFieldCorrection algorithm). The bias field was then applied to the whole set of diffusion imaging data. The DTI model was applied to fit fractional anisotropy (FA), mean diffusivity (MD), and diffusion tensors along the x-, y-, and z -axis by using FSL DTIFIT command with a single shell ($b=1,250$ s/mm²). The DKI model was applied to fit mean kurtosis (MK) with the DIPY Toolbox (<https://www.dipy.org>) with two shells ($b=1,250$ and $2,500$ s/mm²).

Two neuroradiologists (each with 5 years of experience) who were blinded to clinical data placed the regions of interest (ROIs) independently. According to anatomic images, neuroradiologists manually drew 3 to 6 circular ROIs with a 6-mm diameter on the enhanced tumor parenchyma (assumed to be tumor the core). For non-enhancing tumors, ROIs were placed based on T2-FLAIR hyperintensity. The peritumoral edema area corresponded to the regions within 1 cm of the peritumoral parts (hyperintense on FLAIR imaging but no enhancement on CE-T1-weighted imaging) (20). Necrotic, hemorrhagic, and cystic components were avoided. The measurements of FA, MD, and MK were calculated as the average values of the ROIs.

ROIs for DTI-ALPS analysis were placed at the layer of the lateral ventricle body by the two neuroradiologists mentioned above. In this region, the perivascular spaces (x-axis) nearly run perpendicular to the association (y-axis) and projection fibers (z-axis). The major differences in water molecule behavior between x-axis diffusivity in projection fibers (D_{xproj}) and association fibers (D_{xassoc}) and the diffusivity along the y-axis in the region of projection fibers (D_{yproj}) and z-axis diffusivity in the region of association fibers (D_{zassoc}) were derived from the perivascular space. Thus, the ALPS-index was calculated using the following

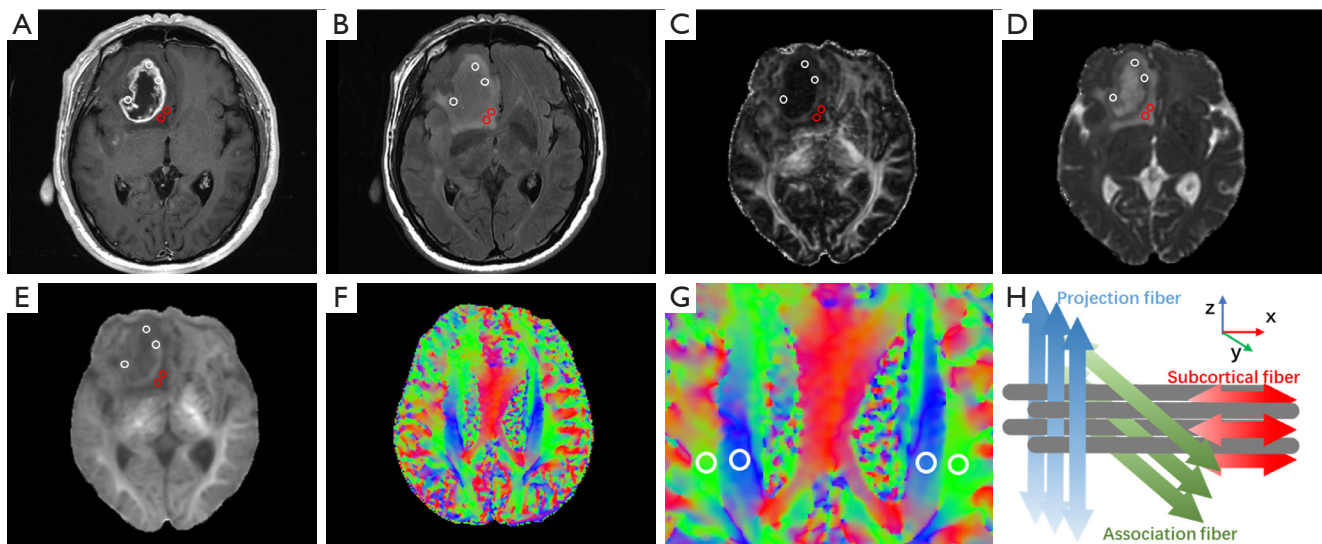


Figure 2 An example of ROI placement. (A,B) CE-T1 and T2-FLAIR. (C-E) On FA, MD, and MK maps, 3–6 ROIs were placed on the tumor parenchyma (white cycles) and peritumoral edema region (red cycles). (F-H) Color-coded V1 maps illustrating ROIs on projection fiber (blue area) and association fiber (green area) in bilateral periventricular regions. The red area indicates the subcortical fiber. (H) Schematic diagram showing the relationship between the direction of the perivascular space (gray cylinder) and the direction of the fibers. CE-T1, contrast-enhanced T1; T2-FLAIR, T2 fluid-attenuated inversion recovery; ROI, region of interest; FA, fractional anisotropy; MD, mean diffusivity; MK, mean kurtosis.

formula:

$$ALPS-index = \frac{\text{mean}(D_{xproj}, D_{xassoc})}{\text{mean}(D_{yproj}, D_{zassoc})} \quad [1]$$

Circular ROIs with a 5-mm diameter were drawn on the association fibers and projection fibers on both hemispheres at the level of the lateral ventricular body according to a color-coded V1 map. The direction of medullary veins is homologous at the level of the lateral ventricle body (21). Therefore, it is not difficult to place ROIs in the proper position. An example of ROI placement is shown in *Figure 2*.

Evaluation of tumor grade and IDH1 mutation status

The tumor grade was diagnosed according to the WHO 2016 classification of tumors (2). Although the fifth edition of the classification was recently released with some changes (22), our study was retrospective in nature and patients were included before the release of the new version, and thus WHO 2016 was still used in this study. The *IDH1* mutation status was determined with next-generation sequencing or immunohistochemistry.

Statistical analysis

All statistical analyses were performed with SPSS version 21 (IBM Corp., Armonk, NY, USA), MedCalc version 18 (MedCalc Software, Ostend, Belgium), and R version 4.1.3 (The R Foundation for Statistical Computing) software. Interobserver reliability was measured using the intraclass correlation coefficient (ICC), and the distribution normality of continuous variables was assessed with the Kolmogorov-Smirnov test. The chi-squared test and 1-way analysis of variance (ANOVA) were used to test the demographic characteristics.

The differences in ALPS-index between bilateral hemispheres were compared with the paired *t*-test. Since the diagnostic efficacy of FA, MD, and MK may be age-related (23,24) and since the ALPS-index was also found to be negatively correlated with age (25), the analysis of covariance (ANCOVA) with age adjustment was used to compare the differences in metrics between patients and healthy controls, between different grades of glioma, and between different *IDH1* genotypes. Bonferroni correction was used for multiple comparisons. To investigate whether the combination of metrics could provide better diagnostic

Table 1 Demographic, clinical, and pathological characteristics of patients

Characteristic	LrGG		GBM	Healthy control (N=31)	P value
	Grade II (N=28)	Grade III (N=17)	Grade IV (N=36)		
Age (years)	43.93±10.50	46.12±14.23	49.17±11.06	45.48±11.34	0.318
Gender (female)	12 (42.9)	11 (64.7)	23 (63.9)	18 (58.1)	0.139
Hemisphere (left)	17 (60.7)	12 (70.6)	16 (44.4)	–	0.168
<i>IDH1</i>					<0.001
<i>IDH1</i> (–)	6 (21.4)	6 (35.3)	27 (75.0)	–	
<i>IDH1</i> (+)	22 (78.6)	11 (64.7)	9 (25.0)	–	

Data are presented as mean ± standard deviation for continuous variables. Data are presented as numbers and frequencies for categorical variables. *IDH1*, isocitrate dehydrogenase 1; LrGG, lower-grade glioma; GBM, glioblastoma.

performance, a binary logistic regression analysis was conducted. Receiver operating characteristic (ROC) curves were used to calculate the area under the curve (AUC), sensitivity, specificity, and the cutoff value. The AUCs of metrics and combinations were compared using the DeLong test. Finally, the integrated discrimination improvement (IDI) indices were calculated to evaluate the added value of the combined models, with a positive IDI value indicating improved discrimination (26). $P < 0.05$ was considered to be statistically significant.

Results

Demographic characteristics of participants

The demographic and clinical characteristics of 81 patients with glioma and 31 healthy controls are summarized in *Table 1*. There were 45 lower-grade gliomas (LrGGs) (20 grade II astrocytomas, 8 grade II oligodendrogliomas, 13 grade III anaplastic astrocytomas, 4 grade III anaplastic oligodendrogliomas) and 36 glioblastomas (GBMs; grade IV). Age and gender were not significantly different between the patients and controls ($P = 0.318$ and $P = 0.139$, respectively). There were significant differences in *IDH1* mutation status ($P < 0.001$) but not in lesion distribution ($P = 0.168$) among patients with grade II–IV gliomas.

Repeatability of measurement

The interobserver variability of measurements is reported in *Table S1*. The ICCs of ALPS-index, FA, MD, and MK ranged from 0.809 to 0.972, showing excellent repeatability between the two observers (27). Thus, the final values

of all measurements were calculated by averaging two independent measurements.

Differences in the ALPS-index between cerebral hemispheres and between patients and healthy controls

The results of comparisons of the ALPS-index between the cerebral hemispheres and between patients and healthy controls are presented in *Figure 3*. There was no significant difference in the ALPS-index between the left and right hemispheres (1.574±0.173 vs. 1.581±0.167; $P = 0.604$) in healthy controls. In all patients with gliomas, the ALPS-index of the hemisphere ipsilateral to glioma was significantly lower than that of the contralateral hemisphere (1.417±0.177 vs. 1.478±0.165; $P = 0.002$). Furthermore, the ALPS-index of the hemisphere ipsilateral to the tumor (left-sided glioma: 1.437±0.181; right-sided glioma: 1.392±0.171) and even the ALPS-index values of the contralateral hemisphere (left-sided glioma: 1.498±0.165; right-sided glioma: 1.454±0.165) were significantly decreased in patients with either left-sided or right-sided gliomas compared to those of the corresponding hemisphere in healthy controls (all P values < 0.05). The ALPS-index of the hemisphere ipsilateral to the tumor showed no significant difference between patients with left-sided and right-sided glioma (1.437±0.181 vs. 1.392±0.171; $P = 0.258$). Therefore, in the subsequent analysis, we did not distinguish if the tumor was located in the left or the right hemisphere.

Correlation of metrics with glioma grade

The results of the differences in metrics between LrGGs and GBMs are shown in *Figure 4A*. For the tumor parenchyma,

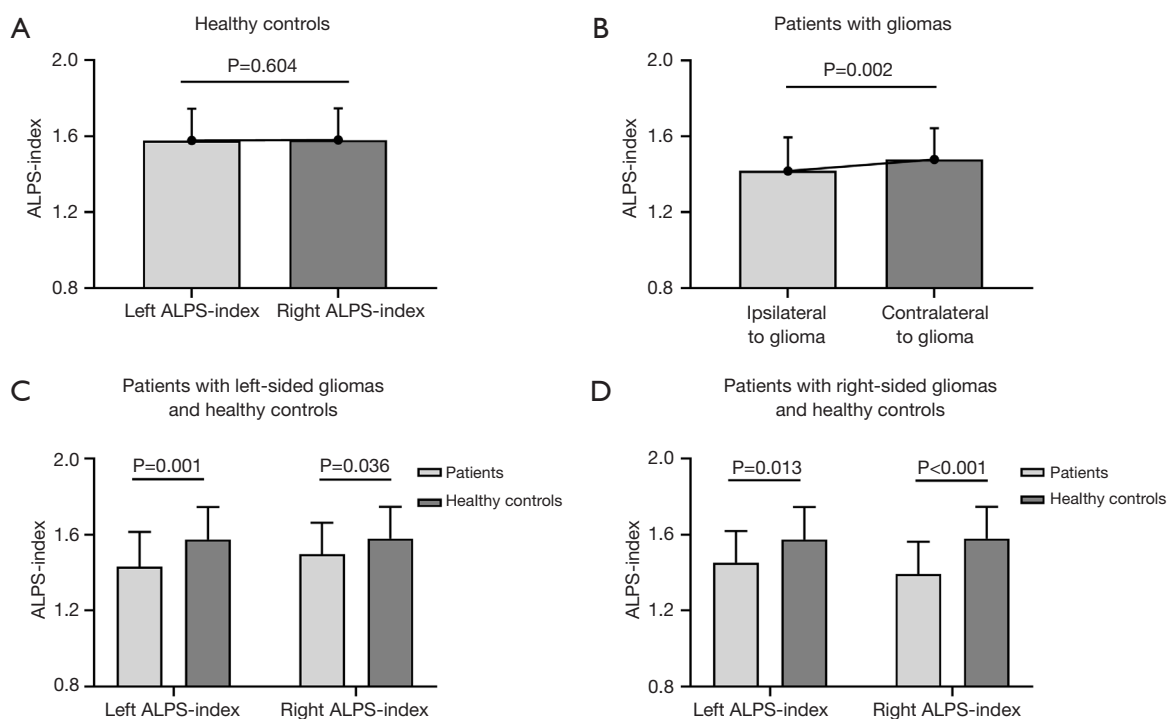


Figure 3 The ALPS-index in bilateral hemispheres in patients with gliomas and healthy controls. (A) Bilateral ALPS-index in healthy controls, (B) bilateral ALPS-index in patients with gliomas, (C) bilateral ALPS-index in patients with left-sided gliomas and healthy controls, and (D) bilateral ALPS-index in patients with right-sided gliomas and healthy controls. ALPS, analysis along the perivascular space.

LrGGs, as compared to GBMs, had a significantly higher ALPS-index (ALPS-index: 1.495 ± 0.151 vs. 1.320 ± 0.159 ; $P < 0.001$) and MD [(1.340 ± 0.373) vs. $(0.975 \pm 0.257) \times 10^{-3} \text{ mm}^2/\text{s}$; $P < 0.001$) but a significantly lower FA (0.123 ± 0.052 vs. 0.166 ± 0.103 ; $P = 0.021$) and MK (0.506 ± 0.155 vs. 0.686 ± 0.172 ; $P < 0.001$). The diffusion metrics in the peritumoral edema regions showed no significant difference between the LrGGs and GBMs [FA: 0.175 ± 0.068 vs. 0.191 ± 0.077 , $P = 0.309$; MD: (1.447 ± 0.245) vs. $(1.377 \pm 0.303) \times 10^{-3} \text{ mm}^2/\text{s}$, $P = 0.234$; MK: 0.504 ± 0.086 vs. 0.547 ± 0.110 , $P = 0.094$]. In addition, the differences in metrics for differentiating LrGGs from GBMs were further compared among grade II, III, and IV gliomas, as shown in Figure 4B and Table S2. The ALPS-index, MD, and MK were significantly different across the three groups.

Table 2 and Figure 5A, 5B show the results of ROC analyses for differentiating LrGGs from GBMs. In univariate analysis, MD showed the highest AUC of 0.807, followed by MK (0.794) and ALPS-index (0.794), and FA had a relatively low AUC of 0.614. The ALPS-index combined with FA, MD, or MK (Table S3) showed increased AUCs of 0.823, 0.854, and 0.854, respectively.

The combination of FA and ALPS-index significantly improved the diagnostic efficacy compared with FA alone ($P = 0.002$). However, the combination of MD and ALPS-index and the combination of MK and ALPS-index yielded no statistically significant improvement in diagnostic efficacy ($P = 0.259$ and $P = 0.102$, respectively). Although the Delong test showed no significant AUC difference for some combined models, significant improvement in discrimination (IDI: $P < 0.05$) could be obtained by adding the ALPS-index to other diffusion metrics. The ROC curves for the comparisons among grade II, III, and IV gliomas are shown in Figure S1, and their corresponding diagnostic characteristics are shown in Table S4.

Correlation of metrics with IDH1 mutation status

Table 3 displays the differences in metrics between the IDH1-mutant group and the wild-type group in both LrGGs and GBMs. In LrGGs, the ALPS-index and MD of tumor parenchyma were significantly lower in the IDH1 wild-type group than in the IDH1-mutant group (ALPS-index: $P = 0.036$; MD: $P = 0.019$), while the FA and MK of

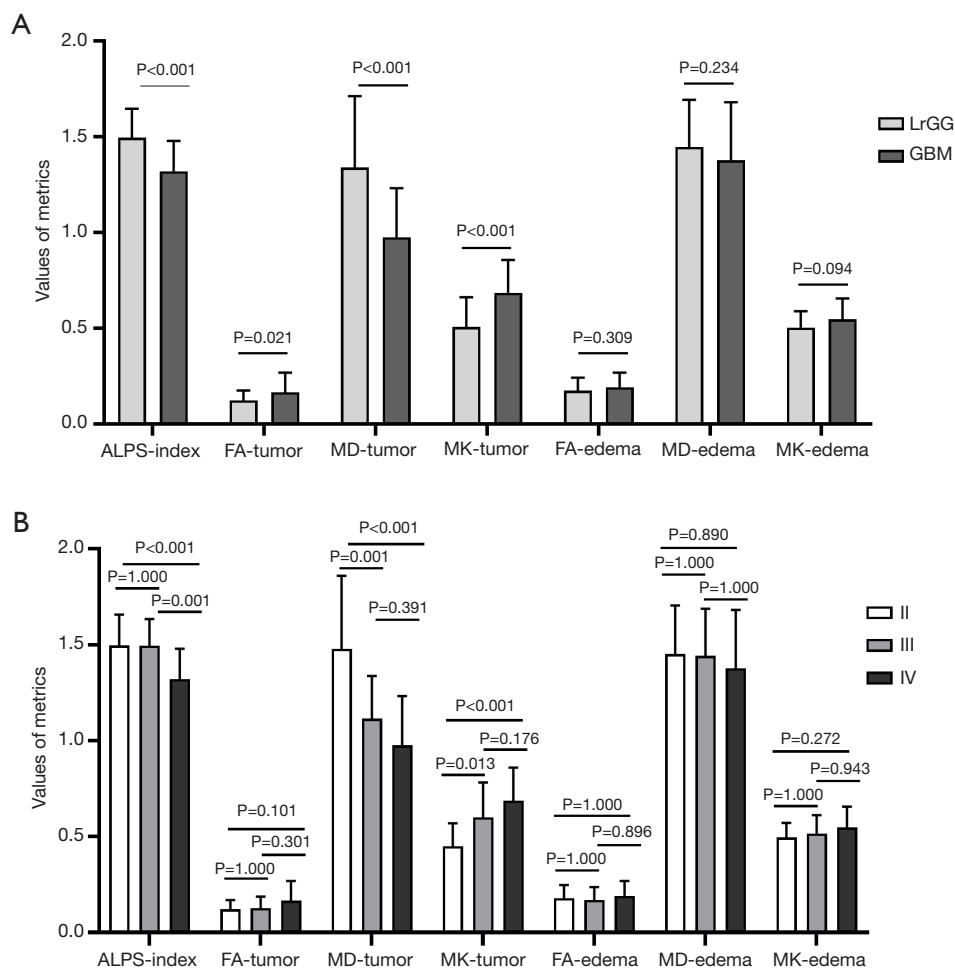


Figure 4 Metrics for glioma grading. (A) Bar charts of diffusion metrics comparing LrGGs and GBMs; (B) bar charts of diffusion metrics comparing grade II, III, and IV gliomas. MD is in units of 10^{-3} mm²/s. LrGG, lower-grade glioma; GBM, glioblastoma; ALPS, analysis along the perivascular space; FA, fractional anisotropy; MD, mean diffusivity; MK, mean kurtosis.

Table 2 Diagnostic performance of diffusion metrics in differentiating LrGGs from GBMs

Metric	AUC (95% CI)	Cutoff value	Sensitivity (%)	Specificity (%)	IDI
ALPS-index	0.794 (0.694–0.895)	1.381	72.22	80.00	–
FA	0.614 (0.486–0.741)	0.138	52.78	71.11	–
FA + ALPS-index	0.823 (0.731–0.915)	–	80.56	73.33	0.237 (P<0.001)
MD	0.807 (0.709–0.905)	1.054	72.22	86.67	–
MD + ALPS-index	0.854 (0.774–0.935)	–	83.33	80.00	0.123 (P=0.004)
MK	0.794 (0.696–0.893)	0.514	86.11	64.44	–
MK + ALPS-index	0.854 (0.772–0.935)	–	91.67	71.11	0.115 (P=0.005)

MD is in units of 10^{-3} mm²/s. LrGG, lower-grade glioma; GBM, glioblastoma; AUC, area under the curve; CI, confidence interval; IDI, integrated discrimination index; ALPS, analysis along the perivascular space; FA, fractional anisotropy; MD, mean diffusivity; MK, mean kurtosis.

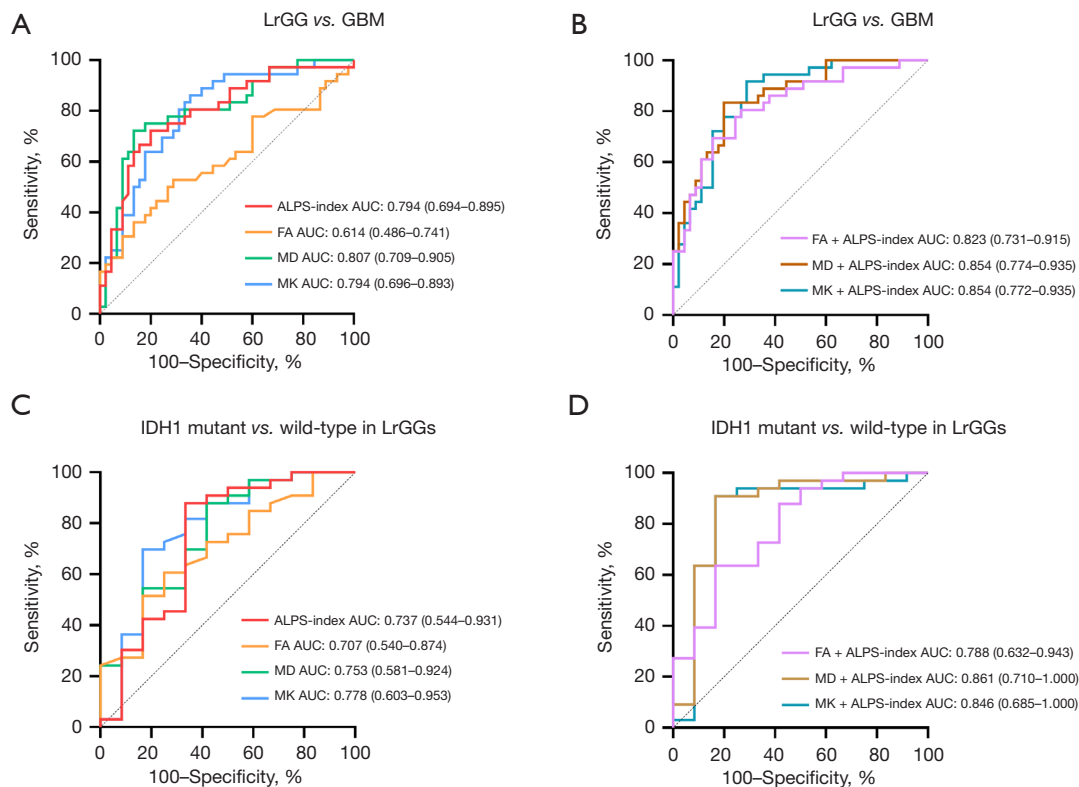


Figure 5 ROC curves of diffusion metrics in evaluating tumor grade and *IDH1* mutation status in LrGGs. ROC curves of single diffusion metrics in evaluating tumor grade (A), ROC curves of combined models in evaluating tumor grade (B), ROC curves of single metrics in evaluating *IDH1* mutation status in LrGGs (C), and ROC curves of combined models in evaluating *IDH1* mutation status in LrGGs (D). The data in parenthesis are shown as 95% CI. ROC, receiver operating characteristic; *IDH1*, isocitrate dehydrogenase 1; LrGG, lower-grade glioma; GBM, glioblastoma; ALPS, analysis along the perivascular space; FA, fractional anisotropy; MD, mean diffusivity; MK, mean kurtosis; CI, confidence interval.

the tumor parenchyma in the *IDH1*-wild-type group were significantly higher than those in the *IDH1*-mutant group (FA: $P=0.015$; MK: $P=0.003$). However, there was no significant difference in these metrics between *IDH1*-wild-type GBMs and *IDH1*-mutant GBMs (all P values >0.05). In addition, none of the peritumoral diffusion metrics showed a significant difference between *IDH1* mutation statuses in either LrGGs and GBMs.

The ROC curves for metrics in predicting *IDH1* mutation are shown in *Figure 5C, 5D*, and the corresponding AUCs, sensitivities, and specificities are summarized in *Table 4*. In univariate analysis, MK showed the highest AUC of 0.778, followed by MD (0.753), ALPS-index (0.737), and FA (0.707). The ALPS-index combined with FA, MD, and MK (*Table S5*) showed improved AUCs of 0.788, 0.861, and 0.846, respectively. Although the differences in AUCs were not significant ($P=0.397$, $P=0.133$, and $P=0.444$,

respectively), all improvements in discrimination were significant (IDI: all P values <0.05).

Correlation among the metrics

The ALPS-index was significantly correlated with MD ($\rho = 0.324$; $P=0.003$) and MK ($\rho = -0.382$; $P<0.001$). However, the correlation between the ALPS-index and FA was not significant ($\rho = -0.103$; $P=0.361$). Moreover, none of the diffusion metrics in peritumoral region were correlated with the ALPS-index (FA: $\rho = -0.181$, $P=0.131$; MD: $\rho = 0.049$, $P=0.685$; MK: $\rho = -0.200$, $P=0.095$). These results are shown in *Figure 6*.

Discussion

Our results demonstrated that in patients with gliomas,

Table 3 Comparisons between the *IDH1*-mutant group and the *IDH1*-wild-type group in LrGGs and GBMs

Metric	<i>IDH1</i> wild-type	<i>IDH1</i> mutant	P value
LrGG			
ALPS-index	1.400±0.185	1.530±0.123	0.036
FA-tumor	0.153±0.060	0.112±0.046	0.015
MD-tumor	1.103±0.313	1.426±0.359	0.019
MK-tumor	0.627±0.193	0.462±0.114	0.003
FA-edema	0.193±0.076	0.169±0.065	0.364
MD-edema	1.332±0.340	1.485±0.199	0.144
MK-edema	0.549±0.118	0.489±0.068	0.129
GBM			
ALPS-index	1.300±0.169	1.380±0.110	0.175
FA-tumor	0.174±0.111	0.142±0.075	0.448
MD-tumor	0.985±0.247	0.947±0.300	0.781
MK-tumor	0.685±0.170	0.690±0.187	0.999
FA-edema	0.194±0.081	0.183±0.071	0.736
MD-edema	1.421±0.279	1.249±0.350	0.186
MK-edema	0.535±0.096	0.579±0.143	0.274

Data are presented as mean ± standard deviation. MD is in units of 10^{-3} mm²/s. *IDH1*, isocitrate dehydrogenase 1; LrGG, lower-grade glioma; GBM, glioblastoma; ALPS, analysis along the perivascular space; FA, fractional anisotropy; MD, mean diffusivity; MK, mean kurtosis.

Table 4 Diagnostic performance of diffusion metrics for *IDH1* mutation status discrimination in LrGGs

Metric	AUC (95% CI)	Cutoff value	Sensitivity (%)	Specificity (%)	IDI
ALPS-index	0.737 (0.544, 0.931)	1.406	87.88	66.67	–
FA	0.707 (0.540, 0.874)	0.117	60.61	75.00	–
FA + ALPS-index	0.788 (0.632, 0.943)	–	63.64	83.33	0.160 (P=0.045)
MD	0.753 (0.581, 0.924)	1.093	87.88	58.33	–
MD + ALPS-index	0.861 (0.710, 1.000)	–	90.91	83.33	0.236 (P=0.002)
MK	0.778 (0.603, 0.953)	0.496	69.70	83.33	–
MK + ALPS-index	0.846 (0.685, 1.000)	–	90.91	83.33	0.146 (P=0.017)

MD is in units of 10^{-3} mm²/s. *IDH1*, isocitrate dehydrogenase 1; LrGG, lower-grade glioma; AUC, area under the curve; CI, confidence interval; IDI, integrated discrimination index; ALPS, analysis along the perivascular space; FA, fractional anisotropy; MD, mean diffusivity; MK, mean kurtosis.

the ALPS-index of the hemisphere ipsilateral to glioma was significantly lower than that of the contralateral hemisphere, and bilateral ALPS-index values in patients were significantly decreased compared with those in healthy controls. These findings may reflect the impairment of the

lymphatic system in patients with glioma. The ALPS-index showed good efficacy in glioma grading and predicting *IDH1* mutation status. In addition, our results showed that the ALPS-index improved diagnostic ability when added to other conventional diffusion metrics.

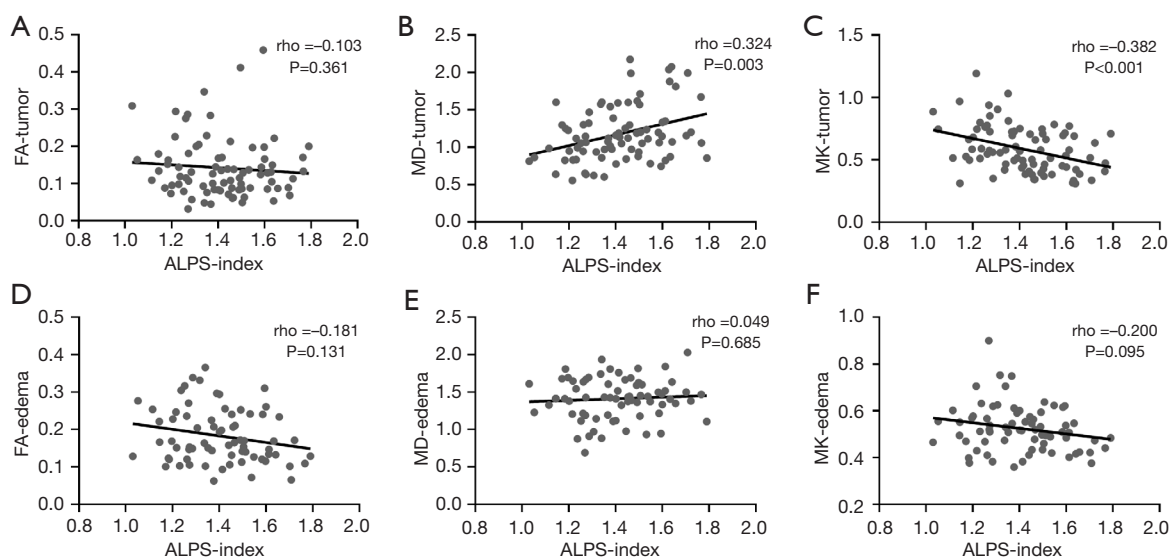


Figure 6 Scatter plots demonstrating the correlation between the ALPS-index and other diffusion metrics. The correlation between ALPS-index and FA of tumor parenchyma (A). The correlation between ALPS-index and MD of tumor parenchyma (B). The correlation between ALPS-index and MK of tumor parenchyma (C). The correlation between ALPS-index and FA of peritumoral edema (D). The correlation between ALPS-index and MD of peritumoral edema (E). The correlation between ALPS-index and MK of peritumoral edema (F). Rho is the Spearman correlation coefficient. MD is in units of $10^{-3} \text{ mm}^2/\text{s}$. ALPS, analysis along the perivascular space; FA, fractional anisotropy; MD, mean diffusivity; MK, mean kurtosis.

The ALPS-index reflects the diffusivity along the perivascular spaces of the medullary veins, which is an important part of the glymphatic system. Thus, a decreased ALPS-index value might partly indicate an impaired glymphatic system in patients with glioma, as observed in rodent experiments in which glioma could induce the reduction of cerebrospinal fluid outflow (6,8). It is also worth noting that the ALPS-index in the contralateral hemisphere of patients in our study was also significantly decreased compared with that in healthy controls. These findings indicate that the glioma might also induce glymphatic dysfunction in the contralateral hemisphere, with the glioma-induced global vasogenic edema potentially being the primary cause (28). In addition, some studies have proposed that gliomas can infiltrate into the contralateral normal-appearing white matter (29,30). However, due to the lack of histopathological confirmation, we cannot directly attribute the observed change to global vasogenic edema or tumor infiltration although this change of ALPS-index was observed in our study.

As it relates to lesion type, the ALPS-index of LrGGs was significantly higher than that of GBMs, which might suggest that LrGGs have better glymphatic function than do GBMs. In one experiment, glioma was found to induce

the remodeling of glymphatic pathways in glioma-bearing mice (7). Given that LrGGs exhibit lower aggressiveness and typically result in longer survival time (31), it is reasonable to assume that the glymphatic system of LrGGs may endure less-severe destruction and be capable of maintain function through more extensive remodeling. We also found that the *IDH1*-wild-type LrGGs had a lower ALPS-index as compared with the *IDH1*-mutant LrGGs, which suggests that the wild-type *IDH1* gene might induce more severe dysfunction of the glymphatic system. Toh *et al.* (32) observed similar results and speculated that reduced glymphatic function of *IDH1*-wild-type LrGGs may be associated with greater aggressiveness and faster tumor growth, thus making remodeling of the glymphatic pathway and compensation of function more difficult. However, the mechanism for this is unclear, and further animal or human studies are needed.

The changes in diffusion metrics in peritumoral regions may be induced by both increased water content and tumor infiltration (33). In our study, we also analyzed diffusion metrics in peritumoral edema, but no significant difference was found between groups, which is consistent with previous studies (20,23) and might be related to the small sample size and tumor heterogeneity of the study. Additionally,

nonsignificant correlations between ALPS and MD and MK were observed in our study. Since previous studies have found that AQP4 expression is negatively correlated with MD and positively correlated with MK (34,35), there might also be a potential correlation between ALPS-index and AQP4 expression in patients with glioma. The expression of AQP4 is upregulated as a reaction to glioma-associated edema (36,37), and the formation of peritumoral edema indirectly reflects the insufficient clearance of interstitial fluid and the disruption of the glymphatic system (32).

DTI- and DKI-derived metrics can be used for glioma grading and *IDH1* mutation status prediction. Although imaging features such as enhancement and edema are widely used in traditional neurosurgical estimation, the extent of enhancement is not always reliable for distinguishing glioma grades (38). Advanced diffusion MRI techniques offer clinically relevant physiological data that are not obtainable via conventional MRI, resulting in more accurate glioma diagnosis. GBMs often have pseudopalisading structures with obvious vascular endothelial hyperplasia, which may result in increased FA values (39). However, this increase may be confused with changes induced by peritumoral edema, which shows tends to have higher FA compared to tumor parenchyma (20,40). With increasing tumor grade, GBMs exhibit increased tumor cellularity and cytoplasm ratios, which restrict the diffusion of water molecules and decrease MD values (24,41). MK may better reflect the complexity of microstructures under non-Gaussian distribution in an organism (15), and a higher MK reflects higher tissue complexity, including increased cell density, hemorrhage, and necrosis. In contrast, a lower MK reflects a higher number of homogeneous nests and well-differentiated cells (42,43). In agreement with previous studies, we also found significantly higher FA, MK, and lower MD values in *IDH1*-mutant LrGGs compared to *IDH1*-wild-type LrGGs (43,44). Similar to GBMs, LrGGs in the in *IDH1*-mutant group were less aggressive than in the wild-type-*IDH1* group, the latter of which had a higher degree of microstructural complexity. In contrast, *IDH1* mutation can inhibit cellular proliferation, leading to fewer diffusion restrictions and less complexity (45,46). However, no metrics were significantly different between *IDH1* mutation statuses in GBMs possibly because GBMs have complex microstructures, perfusions, and other molecular changes besides *IDH1* mutation (47).

In ROC analyses, the ALPS-index demonstrated equivalent efficacy compared to conventional DTI- and DKI-derived metrics in diagnosing glioma grade and

IDH1 mutation status. It is worth noting that although significant differences in ALPS-index between groups might enhance the understanding of the disease progression pattern, there was some degree of overlap in the ALPS-index values between groups. This indicates that additional caution may be necessary when the ALPS-index is applied alone in individual diagnosis. Furthermore, a valuable finding was that the combination models further improved the discrimination abilities in glioma grading and *IDH1* genotyping, which may suggest that conventional DTI and DKI metrics mostly measured in the areas of tumor parenchyma may only reflect the change of intertumoral microstructure (43). Meanwhile, ALPS-index measured outside the tumor may reflect additional pathophysiological information (11,32) and may act complementarily to conventional diffusion metrics, with their combination more comprehensively reflecting the microenvironment change. Furthermore, as the image acquisition time of DTI is acceptable in clinical practice and automated calculation techniques are being developed (48,49), the application of ALPS-index may provide more convenience in clinic.

There are some limitations in this study. First, a single-center design was employed, and the sample size was small. It is reasonable to assume that the different tumor locations might have different impacts on the glymphatic system. However, our sample size was insufficiently large for analyzing tumor location in more refined subgroupings. Considering the imbalanced distribution of *IDH1* mutation status in different grades, we believe a larger sample size is needed in our future study. Second, although radiologists were experienced and ROIs were carefully drawn, subjectivity was inevitable, especially for GBMs with high heterogeneity. Generally, applying multiple ROIs can reduce the biases of subjective evaluation. Finally, although DTI-ALPS has been used in many studies such as Alzheimer disease (13), Parkinson disease (14,50), and normal pressure hydrocephalus (51) to evaluate the function of the glymphatic system, the relationship between the ALPS-index and gold standard contrast-based glymphatic imaging has not yet been substantially validated. Thus, the mechanism underlying the function of this deductive, model-based method should be further investigated.

Conclusions

A lower ALPS-index in patients with glioma may reflect the impairment of the glymphatic system, with these changes potentially being bilateral. In addition, the ALPS-index

Table 5 A synoptic table of main findings in this study

Metrics	Model	Potential pathophysiological implications	Correlation with tumor grade	Correlation with <i>IDH1</i> mutation status	Value in diagnosis
ALPS-index	DTI	Potentially correlated with the function of the glymphatic system	Gliomas can lead to decrease in the ALPS-index, and patients with GBMs have a lower ALPS-index than do patients with LrGGs	Patients with <i>IDH1</i> -wild-type gliomas have a lower ALPS-index than do patients with <i>IDH1</i> mutant gliomas	The ALPS-index could improve the diagnostic ability when added to other conventional diffusion metrics
FA		The measurement of the directionality of molecular motion, which can reflect the white matter integrity in brain	GBMs have a higher FA than do LrGGs	<i>IDH1</i> -wild-type gliomas have a higher FA	These conventional DTI- and DKI-derived metrics have been used in glioma grading and <i>IDH1</i> genotyping
MD		The rate of water molecules' diffusional motion, which correlates with tumor cellularity	GBMs have a lower MD than do LrGGs	<i>IDH1</i> -wild-type gliomas have a lower MD	
MK	DKI	Can provide the kurtosis of water diffusion under non-Gaussian distribution in an organism	GBMs have a higher MK than do LrGGs	<i>IDH1</i> -wild-type gliomas have a higher MK	

IDH1, isocitrate dehydrogenase 1; ALPS, analysis along the perivascular space; FA, fractional anisotropy; MD, mean diffusivity; MK, mean kurtosis; DTI, diffusion tensor imaging; DKI, diffusion kurtosis imaging; LrGG, lower-grade glioma; GBM, glioblastoma.

could provide peritumoral pathophysiological information and complement conventional diffusion metrics for glioma grading and *IDH1* genotyping. Thus, the ALPS-index has considerable potential as a novel imaging biomarker for gliomas. The key findings of this study are summarized in *Table 5*.

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Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <https://qims.amegroups.com/article/view/10.21037/qims-23-541/coif>). The authors report that this study received funding from the National Natural Science Foundation of China (Nos. U22A20354 and 81730049). The authors have no other conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are

appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013) and approved by the Institutional Review Board of Tongji Hospital of Tongji Medical College of Huazhong University of Science and Technology (No. TJ-IRB202303157). Informed consent was obtained from all individual participants.

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References

- Ricard D, Idbaih A, Ducray F, Lahutte M, Hoang-Xuan K, Delattre JY. Primary brain tumours in adults. *Lancet* 2012;379:1984-96.
- Louis DN, Perry A, Reifenberger G, von Deimling A, Figarella-Branger D, Cavenee WK, Ohgaki H, Wiestler OD, Kleihues P, Ellison DW. The 2016 World

- Health Organization Classification of Tumors of the Central Nervous System: a summary. *Acta Neuropathol* 2016;131:803-20.
3. Hartmann C, Hentschel B, Wick W, Capper D, Felsberg J, Simon M, Westphal M, Schackert G, Meyermann R, Pietsch T, Reifenberger G, Weller M, Loeffler M, von Deimling A. Patients with IDH1 wild type anaplastic astrocytomas exhibit worse prognosis than IDH1-mutated glioblastomas, and IDH1 mutation status accounts for the unfavorable prognostic effect of higher age: implications for classification of gliomas. *Acta Neuropathol* 2010;120:707-18.
 4. Turkalp Z, Karamchandani J, Das S. IDH mutation in glioma: new insights and promises for the future. *JAMA Neurol* 2014;71:1319-25.
 5. Iliff JJ, Wang M, Liao Y, Plogg BA, Peng W, Gundersen GA, Benveniste H, Vates GE, Deane R, Goldman SA, Nagelhus EA, Nedergaard M. A paravascular pathway facilitates CSF flow through the brain parenchyma and the clearance of interstitial solutes, including amyloid β . *Sci Transl Med* 2012;4:147ra111.
 6. Ma Q, Schlegel F, Bachmann SB, Schneider H, Decker Y, Rudin M, Weller M, Proulx ST, Detmar M. Lymphatic outflow of cerebrospinal fluid is reduced in glioma. *Sci Rep* 2019;9:14815.
 7. Hu X, Deng Q, Ma L, Li Q, Chen Y, Liao Y, et al. Meningeal lymphatic vessels regulate brain tumor drainage and immunity. *Cell Res* 2020;30:229-43.
 8. Xu D, Zhou J, Mei H, Li H, Sun W, Xu H. Impediment of Cerebrospinal Fluid Drainage Through Glymphatic System in Glioma. *Front Oncol* 2021;11:790821.
 9. He W, You J, Wan Q, Xiao K, Chen K, Lu Y, Li L, Tang Y, Deng Y, Yao Z, Yue J, Cao G. The anatomy and metabolome of the lymphatic system in the brain in health and disease. *Brain Pathol* 2020;30:392-404.
 10. das Neves SP, Delivanoglou N, Da Mesquita S. CNS-Draining Meningeal Lymphatic Vasculature: Roles, Conundrums and Future Challenges. *Front Pharmacol* 2021;12:655052.
 11. Taoka T, Masutani Y, Kawai H, Nakane T, Matsuoka K, Yasuno F, Kishimoto T, Naganawa S. Evaluation of glymphatic system activity with the diffusion MR technique: diffusion tensor image analysis along the perivascular space (DTI-ALPS) in Alzheimer's disease cases. *Jpn J Radiol* 2017;35:172-8.
 12. Zhang W, Zhou Y, Wang J, Gong X, Chen Z, Zhang X, Cai J, Chen S, Fang L, Sun J, Lou M. Glymphatic clearance function in patients with cerebral small vessel disease. *Neuroimage* 2021;238:118257.
 13. Hsu JL, Wei YC, Toh CH, Hsiao IT, Lin KJ, Yen TC, Liao MF, Ro LS. Magnetic Resonance Images Implicate That Glymphatic Alterations Mediate Cognitive Dysfunction in Alzheimer Disease. *Ann Neurol* 2023;93:164-74.
 14. Cai X, Chen Z, He C, Zhang P, Nie K, Qiu Y, Wang L, Wang L, Jing P, Zhang Y. Diffusion along perivascular spaces provides evidence interlinking compromised glymphatic function with aging in Parkinson's disease. *CNS Neurosci Ther* 2023;29:111-21.
 15. Jiang R, Jiang J, Zhao L, Zhang J, Zhang S, Yao Y, Yang S, Shi J, Shen N, Su C, Zhang J, Zhu W. Diffusion kurtosis imaging can efficiently assess the glioma grade and cellular proliferation. *Oncotarget* 2015;6:42380-93.
 16. Bai Y, Lin Y, Tian J, Shi D, Cheng J, Haacke EM, Hong X, Ma B, Zhou J, Wang M. Grading of Gliomas by Using Monoexponential, Biexponential, and Stretched Exponential Diffusion-weighted MR Imaging and Diffusion Kurtosis MR Imaging. *Radiology* 2016;278:496-504.
 17. Xiong J, Tan WL, Pan JW, Wang Y, Yin B, Zhang J, Geng DY. Detecting isocitrate dehydrogenase gene mutations in oligodendroglial tumors using diffusion tensor imaging metrics and their correlations with proliferation and microvascular density. *J Magn Reson Imaging* 2016;43:45-54.
 18. Gates EDH, Lin JS, Weinberg JS, Hamilton J, Prabhu SS, Hazle JD, Fuller GN, Baladandayuthapani V, Fuentes D, Schellingerhout D. Guiding the first biopsy in glioma patients using estimated Ki-67 maps derived from MRI: conventional versus advanced imaging. *Neuro Oncol* 2019;21:527-36.
 19. Veraart J, Novikov DS, Christiaens D, Ades-Aron B, Sijbers J, Fieremans E. Denoising of diffusion MRI using random matrix theory. *Neuroimage* 2016;142:394-406.
 20. Qiu J, Deng K, Wang P, Chen C, Luo Y, Yuan S, Wen J. Application of diffusion kurtosis imaging to the study of edema in solid and peritumoral areas of glioma. *Magn Reson Imaging* 2022;86:10-6.
 21. Okudera T, Huang YP, Fukusumi A, Nakamura Y, Hatazawa J, Uemura K. Micro-angiographical studies of the medullary venous system of the cerebral hemisphere. *Neuropathology* 1999;19:93-111.
 22. Louis DN, Perry A, Wesseling P, Brat DJ, Cree IA, Figarella-Branger D, Hawkins C, Ng HK, Pfister SM, Reifenberger G, Soffietti R, von Deimling A, Ellison DW. The 2021 WHO Classification of Tumors of the Central Nervous System: a summary. *Neuro Oncol*

- 2021;23:1231-51.
23. Van Cauter S, Veraart J, Sijbers J, Peeters RR, Himmelreich U, De Keyser F, Van Gool SW, Van Calenbergh F, De Vleeschouwer S, Van Hecke W, Sunaert S. Gliomas: diffusion kurtosis MR imaging in grading. *Radiology* 2012;263:492-501.
 24. Qi XX, Shi DF, Ren SX, Zhang SY, Li L, Li QC, Guan LM. Histogram analysis of diffusion kurtosis imaging derived maps may distinguish between low and high grade gliomas before surgery. *Eur Radiol* 2018;28:1748-55.
 25. Zhang Y, Zhang R, Ye Y, Wang S, Jiaerken Y, Hong H, Li K, Zeng Q, Luo X, Xu X, Yu X, Wu X, Yu W, Zhang M, Huang P. The Influence of Demographics and Vascular Risk Factors on Glymphatic Function Measured by Diffusion Along Perivascular Space. *Front Aging Neurosci* 2021;13:693787.
 26. Pencina MJ, D'Agostino RB Sr, D'Agostino RB Jr, Vasan RS. Evaluating the added predictive ability of a new marker: from area under the ROC curve to reclassification and beyond. *Stat Med* 2008;27:157-72; discussion 207-12.
 27. Cao T, Jiang R, Zheng L, Zhang R, Chen X, Wang Z, Jiang P, Chen Y, Zhong T, Chen H, Wu P, Xue Y, Lin L. T1 and ADC histogram parameters may be an in vivo biomarker for predicting the grade, subtype, and proliferative activity of meningioma. *Eur Radiol* 2023;33:258-69.
 28. Horváth A, Perlaki G, Tóth A, Orsi G, Nagy S, Dóczy T, Horváth Z, Bogner P. Biexponential diffusion alterations in the normal-appearing white matter of glioma patients might indicate the presence of global vasogenic edema. *J Magn Reson Imaging* 2016;44:633-41.
 29. Mehrabian H, Lam WW, Myrehaug S, Sahgal A, Stanisz GJ. Glioblastoma (GBM) effects on quantitative MRI of contralateral normal appearing white matter. *J Neurooncol* 2018;139:97-106.
 30. Horváth A, Perlaki G, Tóth A, Orsi G, Nagy S, Dóczy T, Horváth Z, Bogner P. Increased diffusion in the normal appearing white matter of brain tumor patients: is this just tumor infiltration? *J Neurooncol* 2016;127:83-90.
 31. Ostrom QT, Bauchet L, Davis FG, Deltour I, Fisher JL, Langer CE, Pekmezci M, Schwartzbaum JA, Turner MC, Walsh KM, Wrensch MR, Barnholtz-Sloan JS. The epidemiology of glioma in adults: a "state of the science" review. *Neuro Oncol* 2014;16:896-913.
 32. Toh CH, Siow TY. Factors Associated With Dysfunction of Glymphatic System in Patients With Glioma. *Front Oncol* 2021;11:744318.
 33. Okita Y, Takano K, Tateishi S, Hayashi M, Sakai M, Kinoshita M, Kishima H, Nakanishi K. Neurite orientation dispersion and density imaging and diffusion tensor imaging to facilitate distinction between infiltrating tumors and edemas in glioblastoma. *Magn Reson Imaging* 2023;100:18-25.
 34. Tan Y, Zhang H, Zhao RF, Wang XC, Qin JB, Wu XF. Comparison of the values of MRI diffusion kurtosis imaging and diffusion tensor imaging in cerebral astrocytoma grading and their association with aquaporin-4. *Neurol India* 2016;64:265-72.
 35. Behnam M, Motamedzadeh A, Aalinezhad M, Dadgostar E, Rashidi Noshabad FZ, Pourfridoni M, Raei M, Mirzaei H, Aschner M, Tamtaji OR. The role of aquaporin 4 in brain tumors: implications for pathophysiology, diagnosis and therapy. *Mol Biol Rep* 2022;49:10609-15.
 36. Mou K, Chen M, Mao Q, Wang P, Ni R, Xia X, Liu Y. AQP-4 in peritumoral edematous tissue is correlated with the degree of glioma and with expression of VEGF and HIF-alpha. *J Neurooncol* 2010;100:375-83.
 37. Yang L, Wang X, Zhen S, Zhang S, Kang D, Lin Z. Aquaporin-4 upregulated expression in glioma tissue is a reaction to glioma-associated edema induced by vascular endothelial growth factor. *Oncol Rep* 2012;28:1633-8.
 38. Togao O, Hiwatashi A, Yamashita K, Kikuchi K, Keupp J, Yoshimoto K, Kuga D, Yoneyama M, Suzuki SO, Iwaki T, Takahashi M, Iihara K, Honda H. Grading diffuse gliomas without intense contrast enhancement by amide proton transfer MR imaging: comparisons with diffusion- and perfusion-weighted imaging. *Eur Radiol* 2017;27:578-88.
 39. Liu X, Tian W, Kolar B, Yeane GA, Qiu X, Johnson MD, Ekholm S. MR diffusion tensor and perfusion-weighted imaging in preoperative grading of supratentorial nonenhancing gliomas. *Neuro Oncol* 2011;13:447-55.
 40. Flores-Alvarez E, Durand-Muñoz C, Cortes-Hernandez F, Muñoz-Hernandez O, Moreno-Jimenez S, Roldan-Valadez E. Clinical Significance of Fractional Anisotropy Measured in Peritumoral Edema as a Biomarker of Overall Survival in Glioblastoma: Evidence Using Correspondence Analysis. *Neurol India* 2019;67:1074-1081.
 41. Cha S. Update on brain tumor imaging: from anatomy to physiology. *AJNR Am J Neuroradiol* 2006;27:475-87.
 42. Hempel JM, Bisdas S, Schittenhelm J, Brendle C, Bender B, Wassmann H, Skardelly M, Tabatabai G, Vega SC, Ernemann U, Klose U. In vivo molecular profiling of human glioma using diffusion kurtosis imaging. *J Neurooncol* 2017;131:93-101.
 43. Zhao J, Wang YL, Li XB, Hu MS, Li ZH, Song YK, Wang JY, Tian YS, Liu DW, Yan X, Jiang L, Yang ZY, Chu JP.

- Comparative analysis of the diffusion kurtosis imaging and diffusion tensor imaging in grading gliomas, predicting tumour cell proliferation and IDH-1 gene mutation status. *J Neurooncol* 2019;141:195-203.
44. Xu Z, Ke C, Liu J, Xu S, Han L, Yang Y, Qian L, Liu X, Zheng H, Lv X, Wu Y. Diagnostic performance between MR amide proton transfer (APT) and diffusion kurtosis imaging (DKI) in glioma grading and IDH mutation status prediction at 3 T. *Eur J Radiol* 2021;134:109466.
45. Bralten LB, Kloosterhof NK, Balvers R, Sacchetti A, Lapre L, Lamfers M, Leenstra S, de Jonge H, Kros JM, Jansen EE, Struys EA, Jakobs C, Salomons GS, Diks SH, Peppelenbosch M, Kremer A, Hoogenraad CC, Smitt PA, French PJ. IDH1 R132H decreases proliferation of glioma cell lines in vitro and in vivo. *Ann Neurol* 2011;69:455-63.
46. Sun C, Zhao Y, Shi J, Zhang J, Yuan Y, Gu Y, et al. Isocitrate dehydrogenase1 mutation reduces the pericyte coverage of microvessels in astrocytic tumours. *J Neurooncol* 2019;143:187-96.
47. Xie Y, Li S, Shen N, Gan T, Zhang S, Liu WV, Zhu W. Assessment of Isocitrate Dehydrogenase 1 Genotype and Cell Proliferation in Gliomas Using Multiple Diffusion Magnetic Resonance Imaging. *Front Neurosci* 2021;15:783361.
48. Saito Y, Kamagata K, Uchida W, Takabayashi K, Aoki S. Improved reproducibility of diffusion tensor image analysis along the perivascular space (DTI-ALPS) index calculated by manual and automated methods. *Jpn J Radiol* 2023;41:1033-4.
49. Saito Y, Kamagata K, Andica C, Uchida W, Takabayashi K, Yoshida S, Nakaya M, Tanaka Y, Kamiyo S, Sato K, Nishizawa M, Akashi T, Shimoji K, Wada A, Aoki S. Reproducibility of automated calculation technique for diffusion tensor image analysis along the perivascular space. *Jpn J Radiol* 2023;41:947-54.
50. Chen HL, Chen PC, Lu CH, Tsai NW, Yu CC, Chou KH, Lai YR, Taoka T, Lin WC. Associations among Cognitive Functions, Plasma DNA, and Diffusion Tensor Image along the Perivascular Space (DTI-ALPS) in Patients with Parkinson's Disease. *Oxid Med Cell Longev* 2021;2021:4034509.
51. Yokota H, Vijayasarithi A, Cekic M, Hirata Y, Linetsky M, Ho M, Kim W, Salamon N. Diagnostic Performance of Glymphatic System Evaluation Using Diffusion Tensor Imaging in Idiopathic Normal Pressure Hydrocephalus and Mimickers. *Curr Gerontol Geriatr Res* 2019;2019:5675014.

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Table S1 The interobserver variability of diffusion metrics

Metric	ICC	95% CI	P value
ALPS-index ipsilateral to glioma	0.927	0.888–0.953	<0.001
ALPS-index contralateral to glioma	0.836	0.695–0.905	<0.001
FA-tumor	0.897	0.841–0.934	<0.001
MD-tumor	0.967	0.948–0.978	<0.001
MK-tumor	0.972	0.957–0.982	<0.001
Bilateral ALPS-index of healthy controls	0.954	0.905–0.978	<0.001
FA-edema	0.895	0.832–0.935	<0.001
MD-edema	0.809	0.694–0.881	<0.001
MK-edema	0.819	0.711–0.887	<0.001

ICC, intraclass correlation coefficient; CI, confidence interval; ALPS, analysis along the perivascular space; FA, fractional anisotropy; MD, mean diffusivity; MK, mean kurtosis.

Table S2 Comparisons among grade II, III, and IV gliomas with age adjustment

Metric	Grade II	Grade III	Grade IV	P	P _{II vs. III}	P _{II vs. IV}	P _{III vs. IV}
ALPS-index	1.495±0.161	1.495±0.139	1.320±0.159	<0.001	1.000	<0.001	0.001
FA-tumor	0.121±0.048	0.126±0.061	0.166±0.103	0.071	1.000	0.101	0.301
MD-tumor	1.477±0.382	1.114±0.222	0.975±0.257	<0.001	0.001	<0.001	0.391
MK-tumor	0.452±0.115	0.595±0.175	0.686±0.172	<0.001	0.013	<0.001	0.176
FA-edema	0.179±0.068	0.169±0.068	0.191±0.077	0.554	1.000	1.000	0.896
MD-edema	1.451±0.253	1.442±0.245	1.377±0.303	0.492	1.000	0.840	1.000
MK-edema	0.495±0.076	0.515±0.097	0.547±0.110	0.215	1.000	0.272	0.943

Data are presented as mean ± standard deviation. MD is in units of 10⁻³ mm²/s. ALPS, analysis along the perivascular space; FA, fractional anisotropy; MD, mean diffusivity; MK, mean kurtosis.

Table S3 Multivariable logistic regression analysis of diffusion metrics in evaluating tumor grade

Regression model	Variable	B	SE	P value
FA + ALPS-index	FA	7.145	3.361	0.033
	ALPS-index	-7.319	1.876	<0.001
MD + ALPS-index	MD	-3.829	1.185	0.001
	ALPS-index	-6.413	1.947	0.001
MK + ALPS-index	MK	5.947	1.950	0.002
	ALPS-index	-5.991	1.876	0.001

B, unstandardized coefficient; SE, standard error; FA, fractional anisotropy; ALPS, analysis along the perivascular space; MD, mean diffusivity; MK, mean kurtosis.

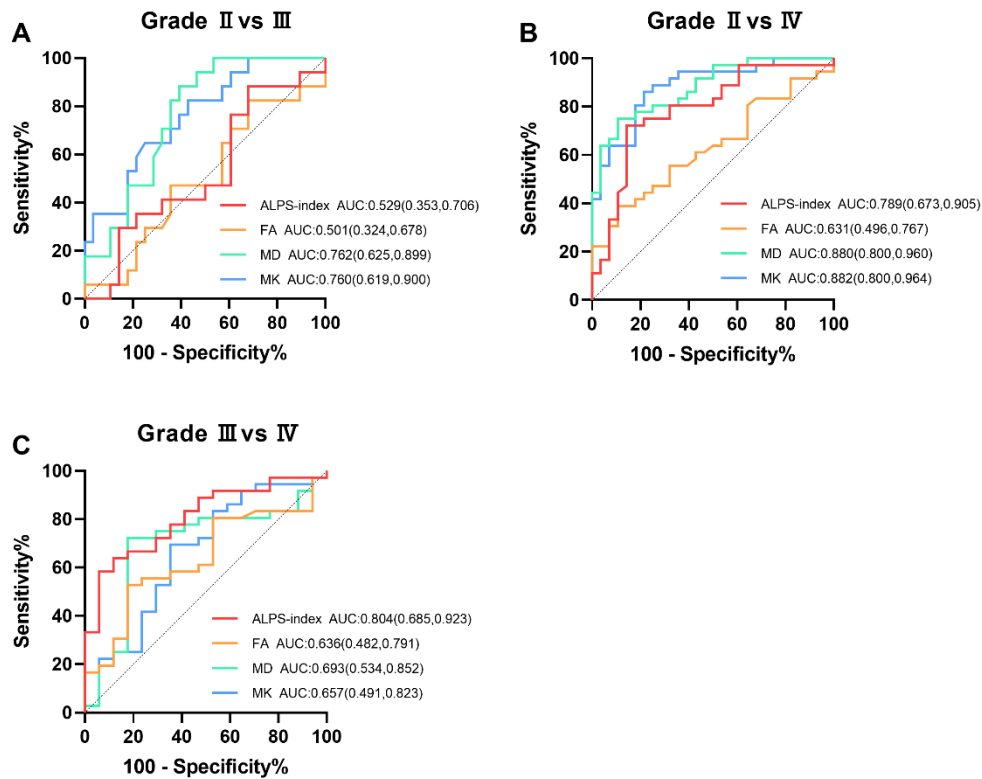


Figure S1 ROC curves of diffusion metrics in differentiating grade II from grade III gliomas (A), grade II from grade IV gliomas (B), and grade III from grade IV gliomas (C). The data in parenthesis are shown as 95% CI. ALPS, analysis along the perivascular space; FA, fractional anisotropy; MD, mean diffusivity; MK, mean kurtosis; ROC, receiver operating characteristic.

Table S4 Diagnostic performance of diffusion metrics in differentiating grade II from grade III gliomas, grade II from grade IV gliomas, and grade III from grade IV gliomas

Grade	Metric	AUC (95% CI)	Cutoff value	Sensitivity (%)	Specificity (%)
II vs. III	ALPS-index	0.529 (0.353, 0.706)	1.621	88.24	32.14
	FA	0.501 (0.324, 0.678)	0.142	82.35	32.14
	MD	0.762 (0.625, 0.899)	1.276	88.24	60.71
	MK	0.760 (0.619, 0.900)	0.508	64.71	75.00
II vs. IV	ALPS-index	0.789 (0.673, 0.905)	1.381	72.22	85.71
	FA	0.631 (0.496, 0.767)	0.174	38.89	89.29
	MD	0.880 (0.800, 0.960)	1.064	75.00	89.29
	MK	0.882 (0.800, 0.964)	0.514	86.11	78.57
III vs. IV	ALPS-index	0.804 (0.685, 0.923)	1.327	58.33	94.12
	FA	0.636 (0.482, 0.791)	0.142	52.78	82.35
	MD	0.693 (0.534, 0.852)	1.052	72.22	82.35
	MK	0.657 (0.491, 0.823)	0.579	69.44	64.71

AUC, area under the curve; CI, confidence interval; ALPS, analysis along the perivascular space; FA, fractional anisotropy; MD, mean diffusivity; MK, mean kurtosis.

Table S5 Multivariable logistic regression analysis of diffusion metrics in evaluating *IDH1* mutation status in LrGGs

Regression model	Variable	B	SE	P value
FA + ALPS-index	FA	-17.126	8.801	0.034
	ALPS-index	6.842	2.901	0.018
MD + ALPS-index	MD	3.734	1.721	0.03
	ALPS-index	6.369	1.933	0.028
MK + ALPS-index	MK	-7.273	2.977	0.015
	ALPS-index	5.728	2.875	0.046

IDH1, isocitrate dehydrogenase 1; LrGG, lower-grade glioma; B, unstandardized coefficient; SE, standard error; FA, fractional anisotropy; ALPS, analysis along the perivascular space; MD, mean diffusivity; MK, mean kurtosis.