

Quantitative assessment of cerebral metabolism and hemodynamics in small-for-gestational-age (SGA) newborns

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Background: Small-for-gestational-age (SGA) newborns represent approximately 10% of births worldwide and 45% of births in some countries. It has been suggested that SGA might cause learning difficulties and behavioral abnormalities in childhood, yet the neurobiological basis for this is poorly understood. In this study, we employed several advanced imaging techniques—including T2-relaxation-under-spin-tagging (TRUST) magnetic resonance imaging (MRI), and phase-contrast (PC) MRI—to quantify oxygen extraction fraction (OEF), global cerebral blood flow (CBF), and cerebral metabolic rate of oxygen (CMRO₂) to elucidate pathophysiological vulnerabilities of SGA neonates.

Methods: A total of 41 newborns were enrolled in this study, consisting of 29 SGA and 12 appropriatefor-gestational-age (AGA) neonates. The SGA group was further divided into subgroups with and without abnormalities on structural MRI, denoted as SGA-a (N=17) and SGA-n (N=12). TRUST and PC MRI were performed to determine OEF, CBF, and CMRO₂. Linear regression analyses were performed to examine physiological parameters' dependence on scan age, gender, and group. Similar analyses were conducted for birth weight and brain volume. Receiver operating characteristic (ROC) curves were used to test physiological parameters' ability to different diagnostic groups.

Results: Regression analysis revealed that CMRO₂ was significantly lower (P=0.04) in the SGA group relative to the AGA group. When further stratifying the SGA participants into SGA-a and SGA-n subgroups, the SGA-a subgroup was found to have the most pronounced physiological deficits, with a lower CMRO₂ (P=0.004) and lower CBF (P=0.007) than those in the AGA group. Conversely, CMRO₂ (P=0.40) and CBF (P=0.90) in the SGA-n subgroup were not different from those of the AGA group. Accordingly, CBF in the SGA-a group was significantly lower (P=0.01) than that of the SGA-n group and CMRO2 also showed a difference (P=0.09). Additionally, CMRO₂ (P=0.002) and CBF (P=0.04) showed an age-related increase during this early developmental period. In analyzing the SGA-a subgroup relative to the remaining neonates, the area under curve (AUC) values were 0.6, 0.6, 0.7, 0.8, and 0.5 for birth weight, OEF, CMRO₂, CBF, and brain volume, respectively. In analyzing the SGA-a subgroup relative to the SGA-n subgroup, AUC values were 0.5, 0.6, 0.7, 0.8, and 0.5 for birth weight, OEF, CMRO₂, CBF, and brain volume.

Conclusions: Structural damage in SGA-a neonates is associated with cerebral hemodynamic and metabolic deficits. SGA neonates with normal CBF and CMRO₂reveal minimal structural abnormalities. Physiological imaging may help identify SGA patients at high risk of developing irreversible brain damage.

Keywords: Small-for-gestational-age (SGA); T2-relaxation-under-spin-tagging (TRUST); phase-contrast (PC); magnetic resonance imaging (MRI); oxygen extraction fraction (OEF); oxygen saturation fraction in venous blood (Yv); cerebral blood flow (CBF); cerebral metabolic rate of oxygen (CMRO₂)

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Introduction

Small-for-gestational-age (SGA) newborns have birth weights and/or birth lengths greater than two standard deviations below established means or below the 10th percentile. SGA and intrauterine growth restriction (IUGR) are often used interchangeably in the literature and clinical workflow (1). The global incidence of SGA newborns has gradually increased in recent years, with a current worldwide prevalence of 9.7% (2), with the highest percentage recorded in South Asia (45%) (3). Compared with appropriate-for-gestational-age (AGA) newborns, intensive-care treatment, morbidity due to perinatal fetal distress and neonatal asphyxia, and perinatal mortality are more likely in SGA infants; they are also prone to hyperbilirubinemia, hypoglycemia, intracranial bleeding, anemia, and respiratory complications. SGA status is also known to influence the nervous system's development, resulting in learning difficulties and neurological/behavioral abnormalities in childhood (4,5). Although most SGA infants show catch-up growth in height and weight, an increased risk of vascular diseases in adulthood has also been reported (6,7). Additionally, it has been suggested that catch-up or postnatal accelerated growth in low birthweight neonates may adversely affect cognitive function (8). Therefore, understanding neonatal brain function, such as cerebral oxygen metabolism and hemodynamics, may provide important pathophysiological insights in SGA infants.

Physiological and functional brain imaging in neonates is particularly challenging due to small brain size, susceptibility to motion, and limited scan time. Computed tomography (CT) (9), positron emission tomography (PET) (10), near-infrared spectroscopy (NIRS) (11,12), and ¹³³xenon clearance (13) are common methods of measuring cerebral blood flow (CBF) and metabolism in adults, but these procedures either involve radiation exposure, present difficulties with visualizing deep brain tissues, or require exogenous tracers; thus, their application in neonatal brain imaging has been limited. Recent advances in magnetic resonance imaging (MRI) technologies have allowed non-invasive and quantitative measurement of brain hemodynamics and metabolism without the need for contrast agents. The brain's oxygen extraction fraction (OEF) can be determined using a T2-relaxation-underspin-tagging (TRUST) MRI technique (14), while global CBF can be measured using quantitative flow phasecontrast (PC) MRI. Therefore, OEF and CBF can be

combined to assess the global cerebral metabolic rate of oxygen (CMRO₂) (14). Importantly, the entirety of these physiological measurements can be completed in less than 5 minutes, making their application practical in the neonatal population (15,16). It should also be noted that arterial spin labeling (ASL), another important perfusion MRI technique, was considered but tends to have a lower signalto-noise ratio, longer scan time, and greater susceptibility to confounding factors such as the bolus arrival time.

In previous research, infants with hypoxic-ischemic encephalopathy (HIE) had lower CBF and CMRO₂ using the T2 prepared tissue relaxation inversion recovery (T2-TRIR) pulse sequence and ASL (17). Newborns with severe HIE had lower CBF and extracted less oxygen than those with moderate HIE using ASL and NIRS. A strong correlation was found between CMRO₂ and CBF in asphyxiated newborns with severe HIE (r=0.88) (18). Our previous work found newborns with white matter damage had lower oxygen consumption and CBF on PC MRI and TRUST MRI (15). Baenziger reported that CBF in SGA neonates was significantly higher than in AGA neonates (16.1±4.1 vs. 11.5±2.6 mL/100 g/min) using ¹³³xenon clearance (19). Few perinatal studies of cerebral oxygen metabolism and hemodynamics in SGA, especially grouping based on abnormal structure, have been reported.

In the present study, we used TRUST MRI and PC MRI to determine OEF, CBF, and $CMRO_2$ in a group of SGA newborns and compared them to a group of AGA neonates.

Methods

Participants

The Ethics Committee approved the study protocol of Shengjing Hospital of China Medical University (IRB2015PS28K). Written informed consent was waived because the data were collected as part of a clinically indicated MRI. Between July 2016 and December 2017, a total of 41 newborns were enrolled. Structural MRI pulse sequences were performed, including axial and sagittal T₁weighted MRI, axial T₂-weighted MRI, and diffusionweighted imaging (DWI). The exclusion criteria were congenital malformations, severe infection, or unusable MRI images. The participant was categorized as SGA if their birth weight was less than the 10th percentile for gestational age, according to the 2013 Fenton & Kim growth chart (20). The AGA participants were required to meet the birth weight criteria greater than the 10th percentile and the absence of structural MRI abnormalities. The participants were further divided into those with abnormal MRI (SGA-a) within the SGA group and those with normal MRI (SGA-n). Grouping of SGA neonates with or without brain injury was determined by the pediatrician based on MRI. There is often no difference between SGA and AGA infants' neurobehavioral development during the perinatal period (21); MRI is often used to observe neonatal brain damage and help determine injury type, injury severity, gray and white matter volume, and prognosis. Imaging helps pediatricians make an accurate diagnosis, prescribe helpful and safe treatment, and develop long-term follow-up observations based on SGA grouping. Rating of MRI for abnormalities was conducted by an experienced neuroradiologist (Y.Q., with 9 years of experience).

General MRI protocol

A 3T MRI system equipped with a phased array head coil (Intera Achieva; Philips Healthcare, Best, Netherlands) was used for all MRI studies. Before imaging, pediatrician sedated infants using chloral hydrate nasal feeding (50 mg/kg) following the standard clinical practice guidelines at our institution. All newborns were well-fed, visually monitored, and fitted with hearing protection during the scans.

The axial T₁-weighted spin-echo images were acquired with the following parameters: repetition time/echo time (TR/TE), 200 ms/2.3 ms; section thickness, 5 mm; the field of view (FOV), 180×150×89 mm³; matrix size, 224×162; and scan time, 34.4 s. Sagittal T₁-weighted spin-echo images used the following parameters: TR/TE, 250 ms/2.3 ms; section thickness, 5 mm; FOV, 230×230×107 mm³; matrix size, 256×250; and scan time, 33.0 s. Axial T₂-weighted imaging used the following parameters: TR/TE, 5,000 ms/80 ms; section thickness, 5 mm; FOV, 180×150×90 mm³; matrix size, 112×112; and scan time, 40.9 s. Axial echoplanar imaging (EPI) DWI was performed as follows: TR/TE, 3,500 ms/63 ms; section thickness, 5 mm; FOV, 180×180×89 mm³; b values 0 and 1,000 s/mm²; and scan time, 30.0 s.

Measurement of OEF

OEF can be determined by:

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$$EF = (Y_a - Y_v)/Y_a \times 100\%$$
 [1]

where Y_a and Y_v represent arterial and venous oxygenation, respectively. Y_a was measured peripherally

using a pulse oximeter on the toe. Y_v was measured using a TRUST MRI technique. TRUST MRI is based on the principle that blood T₂ has a calibratable relationship with oxygenation level; the sequence applies a spin labeling module to isolate pure venous blood signals, then employs a series of T₂-preparation pulses to modulate the MRI signal, the mono-exponential fitting of which yields blood T_2 . A hematocrit (Hct)-specific T_2 - Y_y calibration curve was used to convert venous T_2 to oxygenation, where by Hct was based on newborns' blood test results (22). The imaging slice of TRUST MRI was placed according to previously established procedures (14,15), parallel to the intercommissural lines with a 10-mm distance from the sinus confluence [postmenstrual age (PMA) >36 weeks] or directly through sinus confluence (PMA <36 weeks) (Figure 1). Other parameters included: slab thickness, 80 mm; four effective TEs (eTE) of 0, 40, 80, and 160 ms; TR, 3000 ms; inversion time (TI), 1,022 ms; FOV, 160×160×5 mm³; matrix size, 64×61; SENSE factor, 3; voxel size, 2.5×2.5×5 mm³; τCPMG, 10 ms; eTEs, three pairs; and total scan time, 72 s. Data processing of TRUST MRI followed previously established procedures (14-16). Global Y_v was combined with Y_a to provide an OEF estimation.

Measurement of CBF

Quantitative flow PC MRI was used to measure CBF. PC MRI employs bipolar gradients to encode flow velocity of blood in major arteries of the brain, specifically the left internal carotid artery (LICA), right internal carotid artery (RICA), left vertebral artery (LVA), and right vertebral artery (RVA), providing a measurement of total blood flow to the brain. The flow value can then be normalized using brain volume to obtain CBF in the units of mL/100 g/min. A time-of-flight (TOF) MR angiogram (MRA) was first performed to allow visualization of the arteries mentioned above (Figure 1). The TOF-MRA was conducted by positioning imaging slabs at the center of the epistropheus and stationing 60-mm saturation slabs above to suppress venous signals. The settings of TOF-MRA were: TR/TE, 20 ms/3.45 ms; flip angle, 18°; FOV, 90×90×20 mm³; voxel size, $0.8 \times 0.8 \times 2$ mm³; and scan duration, 23.7 s.

Based on the TOF-MRA images, four PC MRI scans were performed by placing the imaging slice perpendicular and centered on each target artery, respectively, according to previous procedures (14,15) (*Figure 1*). Those of the LICA and RICA was placed at the foramen magnum level, whereas the LVA and RVA imaging slices were placed

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Figure 1 Measurement of Yv and CBF using T2-relaxation-under-spin-tagging (TRUST) and phase-contrast (PC) MRI. (A,D) Axial T1weighted imaging (T_1WI); (B,E) sagittal T_1WI for positioning of TRUSTMRI study; views at effective TEs (eTEs) of 0, 40, 80, and 160 ms; mono-exponential fitting of signal intensity in targeted venous sinus (as function of eTE) yielding venous blood, T_2 then converted to Yv via calibration plot; (C,F) positioning for PCMRI of internal carotid arteries (left, LICA; right, RICA) and vertebral arteries (left, LVA; right, RVA), using maximal-intensity-projection (MIP) angiogram images; velocity maps of LICA, RICA, LVA, and RVA. (A,B,C) show 35.7-old newborn (birth weight, 1,780 g) in SGA-n group. There was normal signal intensity on T_1WI ; (D,E,F) as above of 35.1 week-old newborn (birth weight, 1,200 g) in SGA-a group. There were multiple areas of 35.7-week white matter damage enclosed in the periventricular areas. LICA, left internal carotid artery; RICA, right internal carotid artery; SGA, small-for-gestational-age.

immediately below epistropheus, away from the pivot points. Imaging parameters were: single slice; voxel size, $0.5 \times 0.5 \times 3$ mm³; FOV, $180 \times 180 \times 3$ mm³; maximum velocity encoding, 20 cm/s (non-gated); averages, 2; total scan (four arteries), 1.5 min.

Data processing of PC MRI followed the previously described procedures (14,16). Manual delineation of the LICA, RICA, LVA, and RVA was conducted based on each PC MRI's magnitude image, respectively, and the ROI masks were applied to the velocity maps to obtain flux in these vessels. The sum of fluxes in all vessels yielded the whole brain's blood flow rate (mL/min). To convert the value to CBF, brain volume was obtained by manually tracing the T_2 -weighted images and assuming a parenchymal density of 1.06 g/mL (23). CBF (mL/100 g/min) was then calculated by dividing the total flux by brain weight.

Calculation of CMRO₂

Once OEF and CBF were obtained, CMRO₂ was calculated based on Fick's principle (24,25):

 $CMRO_2 = CBF \times OEF \times Y_a \times C_a$ [2]

where the unit of CMRO₂ is μ mol O₂/100 g/min. C_a represents the oxygen-carrying capacity of blood and is assumed to be 897 μ mol O₂/100 mL for a Hct of 44% (26). Each neonate used a subject-specific C_a based on their Hct level.

Statistical analysis

Regression analysis was used to analyze the scan age and gender differences between SGA and AGA groups, SGA-a and SGA-n groups, SGA-a and AGA groups, and SGA-n and AGA groups, respectively, treating the groups as 0 and 1, and using scan age and gender as covariates. A linear regression analysis was used to examine physiological parameters' dependence on scan age, gender, and diagnostic group; the physiological parameter was assigned as the dependent variable, and scan age, gender, and the group were assigned as independent variables. The following coding was used in the regression analyses between pairs of groups: SGA group and AGA group were coded as 0 and 1, respectively; SGA-a and SGA-n as 1 and 0; SGA-a and

Table 1 Clinical characteristics of the 41 infants and their OEF, CMRO₂, CBF and brain volume in SGA (containing SGA-MRI abnormal and SGA-MRI normal) and AGA groups

Group	Ν	Scan age (weeks)	Birth age (weeks)	Birth weight (g)	OEF (%)	CMRO₂ (µmol/100 g/min)	CBF (mL/100 g/min)	Brain volume (mL)
SGA	29	37.5±3.2	33.5±4.2	1,811.5±612.2	37.0±14.4	33.6±12.2	13.8±5.3	304.0±82.7
SGA-a	17	37.5±2.8	33.7±4.3	1,808.3±570.7	38.5±17.3	30.2±10.1	11.5±4.5	299.0±93.3
SGA-n	12	37.4±3.6	33.3±4.0	1,815.9±666.7	34.9±9.1	38.4±13.3	17.0±4.7	311.2±64.2
AGA	12	36.6±3.7	35.5±3.9	2,777.5±1,117.1	31.4±4.8	40.1±11.4	16.0±3.8	311.5±81.1

Parameter values are reported as mean ± standard deviation. SGA, small-for-gestational-age; SGA-a, SGA neonates with abnormal structural MRI; SGA-n, SGA neonates with normal structural MRI.

AGA as 1 and 0; and SGA-n and AGA as 0 and 1. If scan age or gender was related to CMRO₂, CBF, or OEF, it was necessary to correct birth weight, CMRO₂, CBF, OEF, and brain volume for age and gender before group comparison. For example, corrected OEF was calculated using the formula:

Corrected OEF=OEF- $b_1 \times (age - age) - b_2$ $\times (gender - -gender)$ [3]

The coefficients b_1 and b_2 were based on the scan age and gender regression model results between SGA and AGA groups, SGA-a and SGA-n groups, SGA-a and AGA groups, and SGA-n and AGA groups, respectively, where similar statistical analysis had been reported (27). Corrected birth weight, brain volume, and other physiological parameters were compared using an independent-samples t-test between SGA and AGA, SGA-a and SGA-n, SGA-a and AGA, and SGA-n and AGA groups, separately. Birth weight, brain volume, and cerebral physiological parameters also independently underwent receiver operating characteristic (ROC) curve analysis to distinguish SGA-a from the remaining neonates. Birth weight, brain volume, and cerebral physiological parameters also independently underwent ROC curve analysis to distinguish SGA-a from SGA-n neonates. These statistical analyses used the software SPSS (v21; IBM Corp, Armonk, NY, USA). A P value of 0.05 or less was considered statistically significant.

Results

Participant characteristics

Demographic and clinical characteristics of group participants and their OEF, CMRO₂, CBF, and brain volumes are summarized in *Table 1*. The SGA-n group included 12 neonates (male, 11; female, 1). The SGA-a group included 17 neonates (male, 10; female, 7); five displayed hypoxic/ischemic brain injuries, and five had suffered from premature rupture of membranes (PROM). Their MRI abnormalities included: eight cystic lesions, five instances of white-matter damage, three subarachnoid hemorrhages, and one cerebral infarction (*Figure 2*). The SGA group included the sum of SGA-n and SGA-a participants, thus consisted of 29 subjects in total. The AGA group included 12 neonates (male, 9; female, 3). These subjects received clinically indicated MRI due to certain clinical signs/symptoms, umbilical cord blood gas analysis, or other routine blood tests at birth; however, no MRI abnormalities were observed, presumably excluding any nervous system disease.

Dependence of cerebral bemodynamics and metabolism on diagnostic groups

Figure 1 shows TRUST and PC MRI data from two representative subjects. We first compared the physiological measures of SGA and AGA neonates (*Table 2*). CMRO₂ was significantly lower (P=0.04) in the SGA group relative to the AGA group, but there was no significant difference in CBF (P=0.12) or OEF (P=0.21). Additionally, CMRO₂ (P=0.002) and CBF (P=0.04) showed an age-related scan increase during this early developmental period. Birth weight, but not brain volume, was significantly different between the SGA and AGA neonates.

Next, we divided the SGA neonates into SGA-a and SGA-n subgroups. Compared to the AGA neonates, the SGA-a group had a lower CMRO₂ (P=0.004) and lowered CBF (P=0.007) than the AGA group. Conversely, CMRO₂ (P=0.40) and CBF (P=0.90) in the SGA-n group were not different from the AGA group. Accordingly, CBF in the

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Figure 2 T_1WI , T_2WI and DWI for SGA-a neonates. (A,B,C) A 34.4-week-old newborn (birth weight, 1,840 g; male) with white matter damage and increased signal intensity on T_1WI and DWI; (D,E,F) a 36.1-week-old newborn (birth weight, 1,125 g; male) with periventricular leukomalacia with multiple cystic lesions enclosed in bilateral centrum semiovale. (G,H,I) A 36.1-week-old newborn (birth weight, 1,140 g; male) with cerebral infarction with mixed signal intensity in the right basal ganglia on T_1WI , and increased signal intensity on T_2WI and DWI. T_1WI , T_1 -weighted imaging; DWI, diffusion-weighted imaging; SGA, small-for-gestational-age.

SGA-a group was significantly lower (P=0.01) than that in the SGA-n group, and CMRO₂ also showed a trend of difference (P=0.09) between the SGA-a and SGA-n groups. OEF results showed no statistically significant differences between SGA and AGA groups (P=0.21), SGA-a and SGA-n groups (P=0.52), SGA-a and AGA groups (P=0.19), and SGA-n and AGA groups (P=0.35), separately.

Regression analysis showed no significant differences in scan age and gender between SGA and AGA groups (P=0.45,

P=0.99), SGA-a and SGA-n groups (P=0.95, P=0.08), SGA-a, and AGA groups (P=0.58, P=0.48), and SGA-n and AGA groups (P=0.48, P=0.26), respectively.

Table 2 shows detailed results of coefficients (scan age: b₁; gender: b₂), 95% confidence interval (CI) of coefficients, and their P values. Group comparisons of corrected birth weight, CMRO₂, CBF, OEF, and brain volume for scan age and gender had similar results. The corrected CMRO₂ and corrected CBF showed significant differences between

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Table 2 Regree	ssion analysis betw	een SGA (cor	taining SG	A-a and SG	A-n) and AG	A groups				
	100000 1000000		P value		Co	efficients(b.	1,2)		95% CI of coefficier	nts
aroup	rarameter	Scan age	Gender	Group	Scan age	Gender	Group	Scan age	Gender	Group
SGA and	Birth weight	<0.001*	0.65	<0.001*	190.2	-84.0	1,136.3	141.0 to 239.4	-454.8 to 286.7	778.9–1,493.7
AGA groups	OEF	0.85	0.98	0.21	-0.1	0.1	-5.7	-1.4 to 1.1	-9.2 to 9.4	-14.6 to 3.3
	$CMRO_2$	0.002*	0.40	0.04*	1.8	-3.4	8.0	0.7 to 2.8	-11.4 to 4.7	0.3 to 15.8
	CBF	0.04*	0.18	0.12	0.5	-2.4	2.6	0.02 to 1.0	-5.9 to 1.2	-0.8 to 6.0
	Brain volume	<0.001*	0.27	0.39	15.3	-27.5	20.5	8.7 to 21.9	-77.3 to 22.3	-27.4 to 68.6
SGA-a and	Birth weight	<0.001*	0.98	0.85	150.1	-4.0	-15.6	100.1 to 200.3	-3,385.5 to 377.6	-187.9 to 156.7
SGA-n aroups	OEF	0.86	0.87	0.52	-0.2	-1.1	1.9	-2.0 to 1.7	-15.0 to 12.8	-4.3 to 8.3
	$CMRO_2$	0.05	0.85	0.09	1.4	-1.0	-4.0	-0.004 to 2.8	-11.6 to 9.6	-8.8 to 0.7
	CBF	0.24	0.56	0.01*	0.3	-1.3	-2.5	-0.2 to 0.9	-5.7 to 3.1	-4.6 to -0.6
	Brain volume	0.01*	0.48	0.85	12.1	-25.4	-2.9	2.7 to 21.6	-97.6 to 46.8	-35.6 to 29.7
SGA-a and	Birth weight	<0.001*	0.35	<0.001*	222.2	-202.0	-1,153.1	159.2 to 285.2	-635.6 to 231.7	-1,556.5 to -749.8
AGA groups	OEF	0.74	0.98	0.19	-0.3	-0.2	7.4	-2.0 to 1.5	-12.2 to 11.9	–3.9 to 18.6
	$CMRO_2$	0.004*	0.83	0.004*	1.8	0.8	-11.8	0.6 to 3.0	-7.3 to 9.0	-19.4 to -4.3
	CBF	0.15	0.98	0.007*	0.4	0.05	-4.9	-0.1 to 0.9	-3.7 to 3.7	-8.4 to -1.5
	Brain volume	0.0007*	0.28	0.42	17.4	-34.1	-23.9	8.1 to 26.8	-98.4 to 30.1	-83.7 to 35.8
SGA-n and	Birth weight	<0.001*	0.88	0.0003*	199.8	-52.1	1,131.7	126.5 to 273.2	-788.1 to 684.5	584.9 to 1,678.6
AGA groups	OEF	0.79	0.63	0.35	0.1	-2.1	-3.1	-0.8 to 1.0	-11.1 to 6.9	-9.7 to 3.6
		0.006*	0.45	0.40	2.0	-4.9	4.1	0.6 to 3.3	-18.1 to 8.4	-5.8 to 14.0
	CBF	0.009*	0.50	06.0	0.6	-1.5	-0.2	0.2 to 1.1	-6.1 to 3.1	-3.6 to 3.2
	Brain volume	<0.001*	0.58	0.44	16.2	-15.6	15.9	10.6 to 21.9	-72.6 to 41.6	-26.4 to 58.4
*, P<0.05. SG/	A, small-for-gesta	tional-age; St	GA-a, SGA	neonates	with abnorme	al structural	MRI; SGA-n	, SGA neonates with	normal structural MRI;	Cl, confidence interval.

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Group	Corrected birth weight (g)	Corrected OEF (%)	Corrected CMRO ₂ (µmol/100 g/min)	Corrected CBF (mL/100 g/min)	Corrected brain volume (mL)
SGA	1,761.7±404.9	37.0±14.1	33.1±11.4	13.7±5.1	300.2±73.4
AGA	2,897.7±636.8	31.3±4.6	41.1±8.0	16.3±3.1	320.7±37.9
t	-6.6	1.9	-2.2	-1.6	-0.9
Р	<0.001*	0.06	0.04*	0.11	0.38
SGA-a	1,798.5±333.6	38.6±16.8	30.2±9.5	11.7±4.6	301.6±86.0
SGA-n	1,829.8±445.9	34.7±8.7	38.3±12.1	16.8±4.2	307.5±48.0
t	-0.2	0.7	-1.9	-3.0	-0.2
Р	0.80	0.48	0.06	0.01*	0.84
SGA-a	1,732.3±364.0	38.6±16.7	29.4±9.5	11.3±4.6	294.2±86.8
AGA	2,885.2±593.6	31.2±4.7	41.2±7.9	16.3±3.0	318.2±34.3
t	-6.2	1.4	-3.4	-3.1	-1.0
Р	<0.001*	0.13	0.004*	0.007*	0.33
SGA-n	1,731.1±520.9	34.7±9.0	37.2±12.5	16.6±4.3	303.4±52.5
AGA	2,862.8±652.4	31.6±4.8	41.3±8.3	16.4±3.0	319.3±37.9
t	-4.7	1.0	-0.9	0.1	-0.9
Р	0.0003*	0.31	0.40	0.90	0.40

Table 3 Comparison of corrected values between groups and their subgroups

Parameter values are reported as mean ± standard deviation. *, P<0.05. SGA, small-for-gestational-age; SGA-a, SGA neonates with abnormal structural MRI; SGA-n, SGA neonates with normal structural MRI.

SGA-a and AGA groups (P=0.004, P=0.007). This suggested that the differences in physiological parameters (CMRO₂ and CBF) between SGA-a and AGA groups were related to brain injury, not scan age (*Table 3*).

ROC curves in identifying SGA with abnormal structural MRI

Figure 3A shows ROC curves for birth weight [area under the curve (AUC) =0.6, 95% confidence interval (CI): 0.5-0.8), OEF (AUC =0.6, 95% CI: 0.4-0.8), CMRO₂ (AUC =0.7, 95% CI: 0.5-0.9), CBF (AUC =0.8, 95% CI: 0.6-0.9), and brain volume (AUC =0.5, 95% CI: 0.3-0.7) used to distinguish SGA-a neonates from AGA and SGA-n neonates. *Figure 3B* shows ROC curves of birth weight (AUC =0.5, 95% CI: 0.3-0.7), OEF (AUC =0.6, 95% CI: 0.2-0.7), CMRO₂ (AUC =0.7, 95% CI: 0.5-0.9), CBF (AUC =0.8, 95% CI: 0.6-1.0), and brain volume (AUC =0.5, 95% CI: 0.3-0.8) used to distinguish SGA-a neonates from SGA-n neonates. The AUC values for CBF and CMRO₂ all indicated fair diagnostic utility for identifying SGA-a. It was also observed that physiological parameters yielded higher AUC values than brain volume or body weight.

Discussion

To our knowledge, the present study is the first to determine oxygen consumption and CBF in SGA neonates. Our data suggest that, while SGA newborns generally have lower metabolism and CBF, patients with structural abnormalities show particularly pronounced deficits in these physiological parameters. CMRO₂ and CBF provide a greater ability to identify SGA neonates with abnormal structural MRI, compared with existing birth weight or brain volume measures.

Physiological and clinical considerations

This study's major finding was that SGA neonates with normal structural MRI and AGA neonates have similar CMRO₂ and CBF; however, CMRO₂ and CBF were lower in SGA neonates with abnormal structural MRI. CBF augmentation in SGA infants with normal structural MRI



Figure 3 Comparison of ROC curves for five models (birth weight, OEF, CMRO₂, CBF, brain volume) to identify SGA newborns with abnormal structural MRI (denoted SGA-a). (A) ROC curves used to distinguish SGA-a neonates from AGA and SGA newborns with normal structural MRI (denoted SGA-n). (B) ROC curves used to distinguish SGA-a neonates from SGA-n neonates. CBF and CMRO₂all showed fair utility in diagnosing SGA-a. The physiological parameters had higher AUC than the morphological index of brain volume. ROC, receiver operating characteristic; SGA, small-for-gestational-age; OEF, oxygen extraction fraction; CMRO₂, cerebral metabolic rate of oxygen; CBF, cerebral blood flow.

may correspond with attempted catch-up growth. Perinatal and early postpartum periods are critical developmental phases for low-birthweight neonates (8). In preterm infants or fetuses, growing tissues are characterized by high plasticity and high cellular proliferation rates. Conversely, such processes may alter biochemical and hormonal pathways, increase disease susceptibility in later life, incur long-term neurological deficits, and adversely impact cognitive function (8,25).

Furthermore, premature and low-birthweight newborns have immature cerebrovascular systems. Once disruption to oxygen supply and metabolism occurs, CMRO₂ may decline accordingly, and the likelihood of brain injury is increased due to poor cerebrovascular function. These notions were supported by the significant differences in CMRO₂ and CBF observed between SGA neonates with abnormal structural MRI and AGA neonates. Thus, the hemodynamic and metabolic function may play a critical role in determining long-term, irreversible brain injury in SGA neonates.

Technical considerations

PC MRI and oxygenation MRI have previously been used to study OEF, CMRO₂, and CBF in healthy and hypoxic-ischemic encephalopathic (HIE) neonates (14,16) (Table 4). The physiological parameters measured in the AGA and SGA-n groups were similar to those reported in healthy infants (14), although CBF values appeared slightly higher in our study population (16.0±3.8 vs. 13.4±4.2 mL/100 g/min); differences in scan age, birth age, and birth weight may have contributed to some of the differences. Compared with the SGA-n group, SGA-a neonates had lower CMRO₂ values, which was consistent with results previously reported in HIE neonates (16); however, CBF values in our study population were lower than those in HIE neonates (11.5±4.5 vs. 17.5±6.3 mL/100 g/min). Aside from age, gender, and weight effects, the higher CBF reported in HIE neonates might also be related to therapeutic hypothermia (16). These findings support TRUST MRI and PC MRI's accuracy and reliability in assessing cerebral hemodynamics and metabolism of newborns.

Relationship between physiological parameters and age

We found that $CMRO_2$ and CBF increased with age, which was consistent with previous reports (14,15) and may reflect the increased energy demands of an important developmental period. Therefore, cerebral blood supply and oxygen metabolism appear to increase to match brain development's rapid rate.

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Study	Method	Number of subjects and condition	Scan age (weeks)	Ya (%)	OEF (%)	CMRO ₂ (µmol/100 g/min)	CBF (mL/100 g/min)
Shetty (16)	PC and TRUST	5 hypoxic ischemic encephalopathy	39±1.4	96.8±2.6	24.4±5	34.74±8.3	17.5±6.3
Liu (14)	PC and TRUST	10 healthy neonates	37.4±2.6	95.8±2.2	33.3±2.7	38.3±17.7	13.4±4.2
This study	PC and TRUST	12 SGA-n	37.4±3.6	95.0±2.0	34.9±9.1	38.4±13.3	17.0±4.7
		17 SGA-a	37.5±2.8	95.0±0.0	38.5±17.3	30.2±10.1	11.5±4.5
		12 AGA	36.6±3.7	95.0±3.8	31.4±4.8	40.1±11.4	16.0±3.8

Table 4 Comparison of physiological parameters vs. previously published data

Parameter values are reported as mean ± standard deviation. SGA, small-for-gestational-age; SGA-a, SGA neonates with abnormal structural MRI; SGA-n, SGA neonates with normal structural MRI; TRUST, T2-relaxation-under-spin-tagging; PC, phase-contrast.

Regression analysis showed no significant differences in scan age and gender between groups (P>0.05), suggesting that CMRO₂ and CBF's significant differences between groups were not related to scanning age or gender. *Table 3* confirmed this, showing similar differences between groups for corrected CMRO₂ and CBF.

AUC values of physiological parameters used in identifying SGA-a neonates

We observed that physiological parameters had higher AUC values than the morphological index of brain volume. Although T₁- and T₂-weighted scans readily identified structural abnormalities in SGA neonates, they could not provide information regarding these abnormalities' etiologies. The data showed that SGA-a neonates had lower CBF compared with SGA-n and AGA groups. Perinatal declines in cerebral blood supply and oxygen metabolism may trigger pathophysiological responses with longterm, irreversible consequences, preceding morphological changes. Our ROC curve suggests that CBF and CMRO₂ are moderately effective in identifying SGA newborns with abnormal structural MRI (*Figure 3A,B*).

In summary, TRUST and PC MRI are useful methods for assessing cerebral oxygen consumption and hemodynamics in SGA neonates. While SGA neonates with normal structural MRI have similar physiological parameters to AGA neonates, SGA neonates with abnormal structural MRI showed reducedCMRO₂and CBF. Physiological imaging may be useful in identifying SGA neonates at high risk of developing irreversible brain damage.

Limitations

It is widely established that localized measurements of

physiological parameters for localized brain injury are more sensitive than global measurements, the latter of which can lead to false-negative results in inter-group comparisons; this was a limitation of this study. Considering the CBF autoregulation, and neuroplasticity that occurs after brain injury, global measurement of physiological parameters also have reference values for diagnosis and therapy in the early stages. In the future, we will combine the methods of near infrared reflectance spectroscopy (NIRS) and neurite orientation dispersion and density imaging (NODDI) to measure physiological parameters and microstructural alteration in different areas (28), which might produce new findings.

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Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi. org/10.21037/qims-20-1040). The authors have no conflicts of interest to declare.

Ethical Statement: The Ethics Committee approved the study protocol of Shengjing Hospital of China Medical University (IRB2015PS28K). Written informed consent was waived on the basis that the data were collected as part

of a clinically indicated MRI.

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