



# Increased tissue water in patients with severe sepsis affects tissue oxygenation measured by near-infrared spectroscopy: a prospective, observational case-control study

Chin-Kuo Lin<sup>1,2^</sup>, Shaw-Woei Leu<sup>3</sup>, Ying-Huang Tsai<sup>4,5</sup>, Shao-Kui Zhou<sup>6</sup>, Chieh-Mo Lin<sup>1,2,7</sup>, Shu-Yi Huang<sup>1,7</sup>, Che-Chia Chang<sup>1</sup>, Meng-Chin Ho<sup>1</sup>, Wei-Chun Lee<sup>1</sup>, Min-Chi Chen<sup>8,9</sup>, Ming-Szu Hung<sup>1,10,11,12</sup>, Yu-Ching Lin<sup>1,11,12</sup>, Jhe-Ruei Li<sup>6</sup>, Bor-Shyh Lin<sup>6</sup>

<sup>1</sup>Department of Pulmonary and Critical Care Medicine, Chang Gung Memorial Hospital, Chiayi; <sup>2</sup>Graduate Institute of Clinical Medicine Sciences, College of Medicine, Chang Gung University, Taoyuan; <sup>3</sup>Department of Thoracic Medicine, Chang Gung Memorial Hospital, Taoyuan; <sup>4</sup>Department of Pulmonary and Critical Care Medicine, Chang Gung Memorial Hospital, Linkou; <sup>5</sup>Department of Respiratory Care, College of Medicine, Chang Gung University, Taoyuan; <sup>6</sup>Institute of Imaging and Biomedical Photonics, National Yang Ming Chiao Tung University, Tainan; <sup>7</sup>Department of Nursing, Chang Gung University of Science and Technology, Chiayi; <sup>8</sup>Department of Public Health, Biostatistics Consulting Center, College of Medicine, Chang Gung University, Guishan, Taoyuan; <sup>9</sup>Department of Hematology and Oncology, Chang Gung Memorial Hospital, Chiayi; <sup>10</sup>Department of Respiratory Care, Chang Gung Memorial Hospital, Chiayi; <sup>11</sup>Department of Medicine, College of Medicine, Chang Gung University, Taoyuan; <sup>12</sup>Department of Respiratory Care, Chang Gung University of Science and Technology, Chiayi

**Contributions:** (I) Conception and design: CK Lin, YH Tsai, BS Lin; (II) Administrative support: YH Tsai, MS Hung, YC Lin; (III) Provision of study materials or patients: CK Lin, SW Leu, CM Lin, SY Huang, CC Chang, MC Ho, WC Lee; (IV) Collection and assembly of data: CK Lin, JR Li, SK Zhou; (V) Data analysis and interpretation: CK Lin, MC Chen, BS Lin; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

**Correspondence to:** Bor-Shyh Lin, PhD. Institute of Imaging and Biomedical Photonics, National Yang Ming Chiao Tung University, No. 301, Gaotie 3rd Rd., Guiren Dist., Tainan City 71150. Email: borshyhlin@gmail.com or borshyhlin@nycu.edu.tw.

**Background:** Tissue oedema affects tissue perfusion and interferes with the monitoring of tissue oxygenation in patients with severe sepsis. However, the underlying mechanisms remain unclear. We used a wireless near-infrared spectroscopy (NIRS) device that transmits tri-wavelength light to quantify tissue haemoglobin (Hb) and water (H<sub>2</sub>O) content. We estimated tissue H<sub>2</sub>O in severe sepsis patients and healthy controls, compared their difference, and investigated the correlation of tissue H<sub>2</sub>O with systemic haemodynamics and its impact on tissue oxygenation.

**Methods:** Seventy-seven adult patients with new-onset severe sepsis admitted to the intensive care unit within 72 h and 30 healthy volunteers (controls) were enrolled. The NIRS device was placed on the participant's leg to estimate the relative tissue concentrations of oxy-Hb ([HbO<sub>2</sub>]), deoxy-Hb ([HbR]), total Hb ([HbT]), and H<sub>2</sub>O ([H<sub>2</sub>O]) at rest for three consecutive days. Two-sample *t*-test or Mann-Whitney U test, chi-square test, and generalised estimating equations (GEEs) were used for comparisons.

**Results:** In severe sepsis patients, the [H<sub>2</sub>O] in the anterior tibia was higher [mean (standard deviation, 95% confidence interval), 10.57 (3.37, 9.81–11.34) vs. 7.40 (1.89, 6.70–8.11)] and the [HbO<sub>2</sub>], [HbT], and tissue Hb oxygen saturation (StO<sub>2</sub>) were lower [0.20 (0.01, 0.20–0.20) vs. 0.22 (0.01, 0.22–0.23), 0.42 (0.02, 0.42–0.43) vs. 0.44 (0.02, 0.44–0.45), and 47.25% (1.97%, 46.80–47.70%) vs. 49.88% (1.26%, 49.41–50.35%), respectively] than in healthy controls in first-day measurements. GEE analysis revealed significant differences in [H<sub>2</sub>O], [HbO<sub>2</sub>], [HbT], and StO<sub>2</sub> between groups over three consecutive days (all P≤0.001). In addition, [HbO<sub>2</sub>] and StO<sub>2</sub> levels gradually decreased over time in the patient group. A negative correlation was observed between

<sup>^</sup> ORCID: 0000-0002-5940-3715.

$[H_2O]$  and  $[HbO_2]$  and  $StO_2$ , which became more obvious over time (day 1:  $r=-0.51$  and  $r=-0.42$ , respectively; both  $P<0.01$ ; day 3:  $r=-0.67$  and  $r=-0.63$ , respectively, both  $P<0.01$ ). Systolic arterial pressure was positively related to  $[H_2O]$  ( $r=0.51$ ,  $P<0.05$ , on day 1) but was not associated with tissue oxygenation parameters.

**Conclusions:** NIRS can be used to quantify tissue  $H_2O$ . Severe sepsis patients have increased tissue  $H_2O$ , which responds to changes in arterial blood pressure and affects tissue oxygenation.

**Keywords:** Sepsis; microcirculation; near-infrared spectroscopy (NIRS); tissue oxygenation; tissue oedema

Submitted Feb 11, 2022. Accepted for publication Jul 22, 2022.

doi: 10.21037/qims-22-127

View this article at: <https://dx.doi.org/10.21037/qims-22-127>

## Introduction

Severe sepsis is a life-threatening disease characterised by severe systemic inflammation, unstable haemodynamics, and multiple organ failure (1,2). Current treatment guidelines recommend fluid resuscitation to expand the intravascular volume and the use of vasopressors to achieve a target systemic arterial pressure level for the normalisation of unstable haemodynamics (3,4). However, excessive fluid administration, leading to tissue oedema and deterioration of tissue perfusion, remains a major concern (5). In addition, due to microcirculatory alterations in the presence of severe sepsis, loss of coherence between the macrocirculation and microcirculation occurs (6). Therefore, achieving the recommended systemic arterial pressure target may not guarantee an improvement in peripheral tissue perfusion and oxygenation (6-9). Sepsis with normal blood pressure may still show abnormal microcirculation (10,11). The primary function of the circulatory system is to deliver nutrients and oxygen to the organs and tissues and remove waste products. The ultimate goal of treating sepsis-related circulatory dysfunction is to restore microcirculation to provide adequate tissue perfusion and oxygenation to prevent eventual multiple organ failure (12,13). Therefore, monitoring tissue perfusion and oxygenation is essential for managing sepsis-related circulatory dysfunction.

Near-infrared spectroscopy (NIRS), a non-invasive and indirect method, has been developed and applied to measure tissue oxygenation (14). NIRS can detect the tissue content of haemoglobin (Hb) and myoglobin (Mb) in different oxygenation states (15). The estimation of tissue Hb oxygen saturation ( $StO_2$ ) is based on the differential absorption properties of oxy-Hb ( $HbO_2$ ) and deoxy-Hb (HbR) under different spectra of near-infrared wavelength light (16). In limb skeletal muscle at rest, light absorption is mainly from Mb, which contributes 50% to 70% of

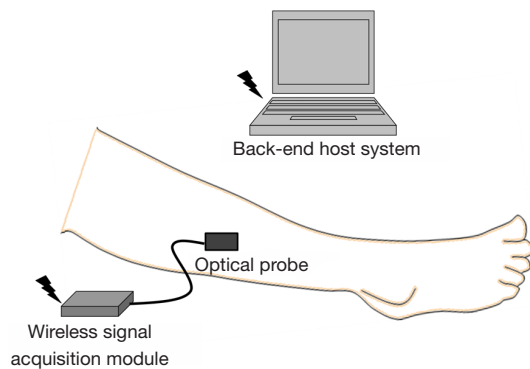
the total light absorption potential for NIRS in skeletal muscle (17). However, Hb rather than Mb can be altered by limb perfusion (18). In the presence of sepsis, the baseline thenar  $StO_2$  is low and changes in  $StO_2$  recovery after an ischaemic challenge are related to the survival of patients in the intensive care unit (ICU) (19). Low  $StO_2$  in early resuscitation is associated with poor outcomes and high mortality rates (20,21). However, sepsis-related tissue oedema caused by endothelial dysfunction and vascular leak has been determined to have a confounding effect on the assessment of  $StO_2$  with NIRS (22,23). To date, the impact of tissue water ( $H_2O$ ) on tissue  $HbO_2$  and HbR has not been comprehensively investigated.

In our previous study, we used NIRS to detect tissue Hb and  $H_2O$  in the extremities of children with Kawasaki disease. We found that tissue  $H_2O$  was significantly higher, but tissue Hb was lower in patients with Kawasaki disease than in healthy controls (24). In the present study, we applied NIRS to simultaneously detect and quantify regional tissue Hb and  $H_2O$  content in adult patients with severe sepsis and healthy volunteers. In addition, we compared the differences in regional tissue oxygenation and  $H_2O$  content between patients and healthy controls, as well as evaluated the relationships among systemic arterial pressure, regional tissue oxygenation, and  $H_2O$  content in patients with severe sepsis. We present the following article in accordance with the STROBE reporting checklist (available at <https://qims.amegroups.com/article/view/10.21037/qims-22-127/rc>).

## Methods

### *Near-infrared diffuse optical technique and wireless optical monitoring system*

The fundamental principle of the near-infrared diffuse optical technique is based on the unique absorption and



**Figure 1** Schematic diagram of the near-infrared spectroscopy system configuration and the position of the optical probe. The wireless optical monitoring system consists of an optical probe, a wireless signal acquisition module, and a back-end host system. The optical probe contains a tri-wavelength LED and a PD, which serve as the light emitter and receiver, respectively. The wireless signal acquisition module consists of several parts: an LED-driving circuit, a PD-sensing circuit, a microprocessor, and a wireless transmission circuit. It is designed to drive and switch the tri-wavelength LED, receive, amplify, and filter the optical signal obtained from the PD, and wirelessly transmit the optical signal to the back-end host system. The tri-wavelength light emitter provides a tri-wavelength light that penetrates through the tissue, and the light receiver receives the penetrated light. The optical signal received from the PD was digitised and transmitted to the back-end host system. A commercial laptop was used as the platform of the back-end host system and estimated the relative tissue concentrations of oxy-haemoglobin, deoxy-haemoglobin, and water from the change in the optical density corresponding to different wavelengths. LED, light-emitting diode; PD, photodiode.

scattering properties of different tissue components corresponding to different wavelengths of light (15,25–29). For red and near-infrared wavelength light (range, 600–1,200 nm), HbO<sub>2</sub>, HbR, Mb and H<sub>2</sub>O are the major absorbers in human tissue. Under the assumption that the Mb concentration during the short term is constant, we only monitor the change of HbO<sub>2</sub>, HbR, and H<sub>2</sub>O in this study. Based on the apparent difference in the absorption spectra of HbO<sub>2</sub>, HbR, and H<sub>2</sub>O corresponding to different wavelengths, we used a tri-wavelength light (700, 910, and 950 nm)-emitting diode to assemble a wireless optical monitoring system comprising an optical probe, a wireless signal acquisition module, and a back-end host system (Figure 1). The optical probe was positioned on the volar surface of the right leg of the patients and healthy

volunteers, at the level of the anterior tibial muscle, to estimate the relative tissue concentrations of HbO<sub>2</sub> ([HbO<sub>2</sub>]), HbR ([HbR]), total Hb ([HbT]), and H<sub>2</sub>O ([H<sub>2</sub>O]) from the change in the optical density corresponding to different wavelengths. For the wavelength  $\lambda$ , the optical density attenuation of red or near-infrared light in human tissue can be simply expressed as

$$OD(\lambda) = (\varepsilon_{HbO_2}(\lambda) \cdot [HbO_2] + \varepsilon_{HbR}(\lambda) \cdot [HbR] + \varepsilon_{H_2O}(\lambda) \cdot [H_2O]) \cdot d \cdot B(\lambda) \quad [1]$$

where  $d$  is the distance between the light source and detector,  $B(\lambda)$  is the differential path-length factor, and  $\varepsilon_{HbO_2}(\lambda)$ ,  $\varepsilon_{HbR}(\lambda)$ , and  $\varepsilon_{H_2O}(\lambda)$  denote the molar extinction coefficient of [HbO<sub>2</sub>], [HbR], and [H<sub>2</sub>O]. Then, [HbO<sub>2</sub>], [HbR], and [H<sub>2</sub>O] can be estimated by the approach of least-squares approximation, as followings,

$$\begin{aligned} [\{HbO_2\}, \{HbR\}, \{H_2O\}] &= \left( (E^T \cdot E)^{-1} \cdot E^T \cdot A \right) / d \\ \text{where } E &= \begin{bmatrix} \{\varepsilon_{HbO_2}(\lambda_1), \varepsilon_{HbR}(\lambda_1), \varepsilon_{H_2O}(\lambda_1)\}, \\ \{\varepsilon_{HbO_2}(\lambda_2), \varepsilon_{HbR}(\lambda_2), \varepsilon_{H_2O}(\lambda_2)\}, \\ \{\varepsilon_{HbO_2}(\lambda_3), \varepsilon_{HbR}(\lambda_3), \varepsilon_{H_2O}(\lambda_3)\} \end{bmatrix} \\ \text{and } A &= \left[ \frac{OD(\lambda_1)}{B(\lambda_1)}, \frac{OD(\lambda_2)}{B(\lambda_2)}, \frac{OD(\lambda_3)}{B(\lambda_3)} \right] \end{aligned} \quad [2]$$

StO<sub>2</sub> was calculated as

$$StO_2 = \frac{[HbO_2]}{[HbO_2] + [HbR]} \times 100\% \quad [3]$$

### Study design and participants

This prospective, observational case-control study was approved by the Institutional Review Board of Chang Gung Medical Foundation (approval No. 103-5357B). The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013) and was conducted in a 20-bed medical ICU of Chiayi Chang Gung Memorial Hospital from 27 November 2015 to 30 April 2019. Adult patients (age  $\geq 18$  years) who were transferred to the ICU from the emergency department and admitted within 72 h for new-onset severe sepsis were enrolled. Severe sepsis was defined as sepsis with sepsis-induced tissue hypoperfusion or organ dysfunction according to the 2012 Surviving Sepsis Campaign criteria (30). Patients with pregnancy, active pulmonary tuberculosis, or contact isolation were excluded from the study. Healthy volunteers who did not have any disorders were recruited among the students of the National Chiao Tung University as healthy controls. Informed consent was obtained from the healthy volunteers

and patients. Legal guardians provided informed consent to participate if the patient had cognitive impairment. After signing the informed consent form, the patients and healthy volunteers received a non-invasive NIRS device for use in estimations at rest. The estimated data were recorded after 3 min to stabilise the NIRS signals (31). The NIRS data were collected for 3 consecutive days or until the patient was transferred to an ordinary ward, discharged against medical advice, or died. We recorded the demographic and clinical data of all patients, including age, sex, aetiology of severe sepsis, acute physiology and chronic health evaluation II score on admission, systemic haemodynamic parameters, laboratory results, and ICU outcome. Patients with missing or incomplete NIRS data were excluded from the final analysis.

### Sample size calculation

The sample size estimate for the study was based on a comparison of repeated measures between patients and controls. Since this study was a pilot study and there was no previous applicable research to calculate the effect size, we considered a power of 0.95 at the 0.05 alpha level and an estimated effect size of 0.15 to calculate the sample size. Using G\*Power, a sample size of 70 per group was required. However, that was inflated by 20% for any potentially missing data, thus yielding a priori sample size of 84 per group.

### Statistical analyses

Continuous data were summarised as mean, standard deviation (SD), and 95% confidence interval (CI) or median and interquartile range (IQR), as appropriate. Categorical data were expressed using counts and percentages. A two-sample *t*-test was used to compare the differences between cases and controls for continuous variables once normality was demonstrated; otherwise, the nonparametric Mann-Whitney U test was performed. The chi-square or Fisher's exact test was performed for categorical variables. Pearson correlation coefficients were used to investigate pairwise relationships between continuous variables. As the relative tissue concentrations of Hb and H<sub>2</sub>O and tissue oxygenation were repeatedly measured at designated intervals over time, generalised estimating equations (GEEs), which consider the correlation within individuals, were employed to evaluate the differences in NIRS parameters between

groups. Statistical analyses were performed using SPSS version 22 (IBM Corp., Armonk, NY, USA). All tests were two-tailed with a significance level of 0.05.

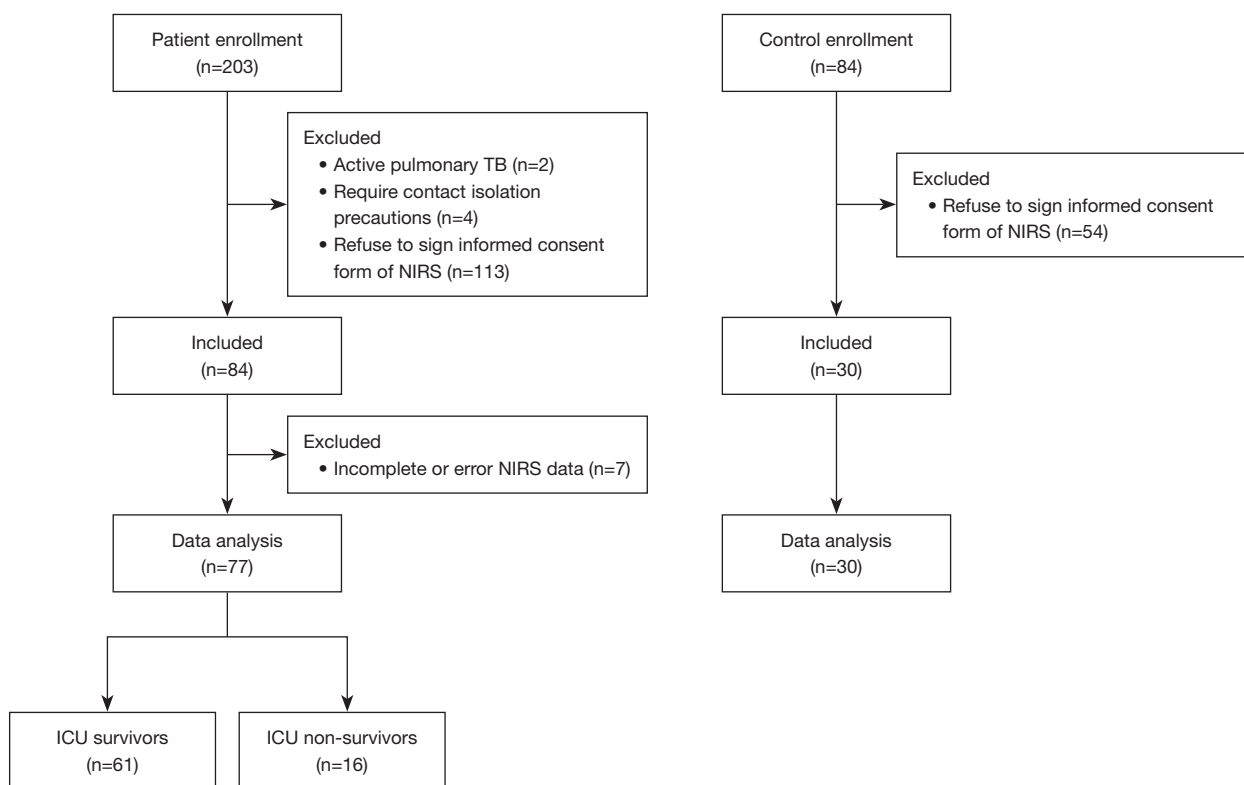
## Results

### *Demographic characteristics and clinical data of patients with severe sepsis*

A total of 203 consecutive patients were assessed for eligibility (Figure 2). Six patients were excluded because of active pulmonary tuberculosis or requiring contact isolation, and 113 patients declined to participate in the study. The remaining 84 patients received the NIRS device for estimating regional tissue oxygenation and H<sub>2</sub>O content. In seven patients, the wireless signals of the device were intermittently received by the back-end host system, and the data were incompletely recorded. Therefore, the integrated data of the remaining 77 patients were analysed. Of these 77 patients, 30 were women and 47 were men (Table 1). The median [IQR] age of the patients was 75 [65–83] years. In addition, 45 patients (58%) had septic shock and received vasoactive agent therapy. In the three consecutive days, the total mean (SD) of the patient's net intake and output before the different NIRS measurements was 514.86 (960.87) mL per day, and the total mean (SD) of the amount of fluid administered to patients before the different NIRS measurements was 1,376.16 (823.84) mL per day. Furthermore, 51 patients (66%) had pulmonary infection, and 61 patients (79%) were ICU survivors. The median (IQR) length of ICU stay was 7.00 (4.00–11.50) days. There were a total of 30 healthy controls recruited for the study (Figure 2). Among the healthy controls, 20 (67%) were men with a median [IQR] age of 24 [23–25] years, which was significantly lower than the age of patients with sepsis (*P*<0.001) (Table 1).

### *Comparisons of relative tissue concentrations of haemoglobin and water and tissue oxygenation between healthy controls and patients with severe sepsis*

In patients with severe sepsis, the first-day measurement of anterior tibial muscle [H<sub>2</sub>O] was higher [mean (SD, 95% CI), 10.57 (3.37, 9.81–11.34) *vs.* 7.40 (1.89, 6.70–8.11); Table 2] and the [HbO<sub>2</sub>], [HbT], and StO<sub>2</sub> were lower [0.20 (0.01, 0.20–0.20) *vs.* 0.22 (0.01, 0.22–0.23), 0.42 (0.02, 0.42–0.43) *vs.* 0.44 (0.02, 0.44–0.45), and 47.25% (1.97%, 46.80–47.70%) *vs.* 49.88% (1.26%, 49.41–50.35%), respectively;



**Figure 2** Flowchart of patient inclusion and exclusion in the study. TB, tuberculosis; NIRS, near-infrared spectroscopy; ICU, intensive care unit.

Table 2] than in healthy controls. In the next two consecutive days, the repeated measurements of  $[H_2O]$  were higher, whereas  $[HbO_2]$ ,  $[HbT]$ , and  $StO_2$  remained lower in patients than in healthy controls (Figure 3). The GEE analysis also showed that these four parameters were significantly different between the two groups (all  $P \leq 0.001$ , Figure 3).

However, in terms of  $[HbR]$ , the patients had the same value as the healthy controls [mean (SD, 95% CI), 0.22 (0.02, 0.22–0.23) vs. 0.22 (0.02, 0.22–0.23), in first-day measurements; Table 2], and the GEE analysis revealed that there was no difference among the 3-day repeated measurements between the groups ( $P=0.996$ , 0.248, and 0.121 on days 1, 2, and 3, respectively; see <https://cdn.amegroups.com/static/public/qims-22-127-1.docx>). Furthermore, concerning the changes in the parameters over time,  $[HbO_2]$  and  $StO_2$  gradually decreased in patients with severe sepsis (day 1 vs. day 3,  $P=0.041$  and  $P=0.036$ , respectively; Figure 3). However, there were no changes in the time trends of tissue  $H_2O$  content in either group or tissue oxygenation in healthy controls.

#### **Relationship between tissue oxygenation and water content in severe sepsis**

With respect to the relationship between tissue oxygenation and  $H_2O$  content, negative correlations were found between  $[H_2O]$  and  $[HbO_2]$  ( $r=-0.51$ ,  $-0.53$ , and  $-0.67$  on days 1, 2, and 3, respectively,  $P < 0.01$  for all; Table 3) and  $StO_2$  ( $r=-0.42$ ,  $-0.43$ , and  $-0.63$  on days 1, 2, and 3, respectively,  $P < 0.01$ , respectively; Table 3); however,  $[H_2O]$  and  $[HbR]$  were positively correlated ( $r=0.28$  on day 2 and 0.54 on day 3,  $P < 0.05$  and  $P < 0.01$ , respectively; Table 3). The correlations between  $[H_2O]$  and  $[HbO_2]$ ,  $[HbR]$ , and  $StO_2$  gradually became stronger during the 3-day repeated measurements (Figure 4A–4C). Furthermore, the correlation between  $[HbT]$  and  $StO_2$  was negative ( $r=-0.29$  on day 1,  $P < 0.05$ ;  $r=-0.46$  and  $-0.65$  on days 2 and 3, respectively,  $P < 0.01$  for both; Table 3 and Figure 4D). Meanwhile, the relationship between  $[HbO_2]$  and  $[HbR]$  changed over time: positive on day 1 ( $r=0.30$ ,  $P < 0.01$ ; Table 3) but negative on days 2 and 3 ( $r=-0.65$ ,  $P < 0.01$ , and  $-0.81$ ,  $P < 0.01$ , respectively; Table 3 and Figure 4E).

**Table 1** Demographic characteristics and clinical data of healthy controls and patients with severe sepsis

Variables	Controls (n=30)	Patients (n=77)	P value
Age (years), median [IQR]	24 [23–25]	75 [65–83]	<0.001
Sex, male, n (%)	20 (67%)	47 (61%)	0.589
Body height (cm), mean (SD)	170.53 (8.11)	160.19 (7.73)	<0.001
Body weight (kg), median (IQR)	68.15 (56.58–78.70)	58.00 (49.50–67.00)	0.001
Body mass index (kg/m <sup>2</sup> ), median (IQR)	22.43 (20.06–26.96)	22.49 (19.52–24.99)	0.528
Septic shock with using vasoactive agents, n (%)	–	45 (58%)	–
Glasgow coma scale, median [IQR]	–	8 [6–15]	–
Acute Physiology and Chronic Health Evaluation Score II, mean (SD)	–	18.91 (6.60)	–
Mean arterial pressure (mmHg), median (IQR)	–	87.50 (78.00–95.00)	–
Systolic arterial pressure (mmHg), mean (SD)	–	114.75 (19.58)	–
Diastolic arterial pressure (mmHg), median (IQR)	–	59.00 (53.50–70.50)	–
White blood cells (1,000/ $\mu$ L), mean (SD)	–	13.64 (8.56)	–
Haemoglobin (g/dL), mean (SD)	–	11.19 (2.65)	–
Creatinine (mg/dL), median (IQR)	–	1.59 (0.98–2.45)	–
Arterial lactate (mg/dL) (n=75)*, median (IQR)	–	19.40 (13.60–31.40)	–
Partial pressure of oxygen (mmHg), median (IQR)	–	105.80 (83.75–143.55)	–
Arterial oxygen saturation (%), median (IQR)	–	98.10 (96.95–99.35)	–
Intake and output, mL/day**			
Day 1 (n=70)*, median (IQR)		201.00 (–262.25 to 1,152.75)	
Day 2 (n=77), median (IQR)		531 (–132 to 1,380)	
Day 3 (n=75)*, mean (SD)		337.69 (1,222.80)	
Total mean, mean (SD)		514.86 (960.87)	
Intravascular fluid administration, mL/day**			
Day 1 (n=70)*, median (IQR)		973.00 (590.00–1,588.75)	
Day 2 (n=77), median (IQR)		1,414.00 (902.00–1,984.50)	
Day 3 (n=75)*, median [IQR]		988 [480–1,693]	
Total mean, mean (SD)		1,376.16 (823.84)	
Diagnosis, n (%)			
Pulmonary infection		51 (66%)	
Urinary tract infection		36 (47%)	
Hepatic or biliary tract infection		8 (10%)	
Spontaneous bacteria peritonitis		1 (1%)	
Pelvic infection		1 (1%)	
Cellulitis		4 (5%)	
Other		3 (4%)	
ICU survivor, n (%)	–	61 (79%)	–
ICU length of stay, days, median (IQR)	–	7.00 (4.00–11.50)	–

\*, variable with missing or unrecorded data; \*\*, data recorded on the day before the NIRS measurement. n, count; IQR, interquartile range; SD, standard deviation; ICU, intensive care unit; NIRS, near-infrared spectroscopy.

**Table 2** Comparisons of the relative tissue concentrations of haemoglobin and water and regional tissue oxygenation between healthy controls and patients with severe sepsis

Measurements	Days	Controls			Patients		
		Mean (SD)	95% CI	n	Mean (SD)	95% CI	n
[HbO <sub>2</sub> ] (a.u.)	1	0.22 (0.01)	0.22–0.23	30	0.20 (0.01)	0.20–0.20	77
	2	0.22 (0.01)	0.22–0.23	30	0.20 (0.02)	0.19–0.20	75
	3	0.22 (0.01)	0.22–0.23	30	0.19 (0.03)	0.19–0.20	64
[HbR] (a.u.)	1	0.22 (0.02)	0.22–0.23	30	0.22 (0.02)	0.22–0.23	77
	2	0.22 (0.02)	0.22–0.23	30	0.23 (0.03)	0.22–0.24	75
	3	0.22 (0.02)	0.22–0.23	30	0.23 (0.04)	0.22–0.24	64
[HbT] (a.u.)	1	0.44 (0.02)	0.44–0.45	30	0.42 (0.02)	0.42–0.43	77
	2	0.44 (0.02)	0.44–0.45	30	0.42 (0.02)	0.42–0.43	75
	3	0.44 (0.02)	0.43–0.45	30	0.42 (0.03)	0.42–0.43	64
StO <sub>2</sub> (%)	1	49.88 (1.26)	49.41–50.35	30	47.25 (1.97)	46.80–47.70	77
	2	49.97 (1.35)	49.47–50.47	30	46.40 (5.25)	45.19–47.61	75
	3	49.92 (1.28)	49.45–50.40	30	45.66 (6.80)	43.96–47.35	64
[H <sub>2</sub> O] (a.u.)	1	7.40 (1.89)	6.70–8.11	30	10.57 (3.37)	9.81–11.34	77
	2	7.60 (1.95)	6.87–8.33	30	10.52 (3.40)	9.74–11.30	75
	3	7.49 (1.92)	6.77–8.21	30	10.79 (3.40)	9.94–11.63	64

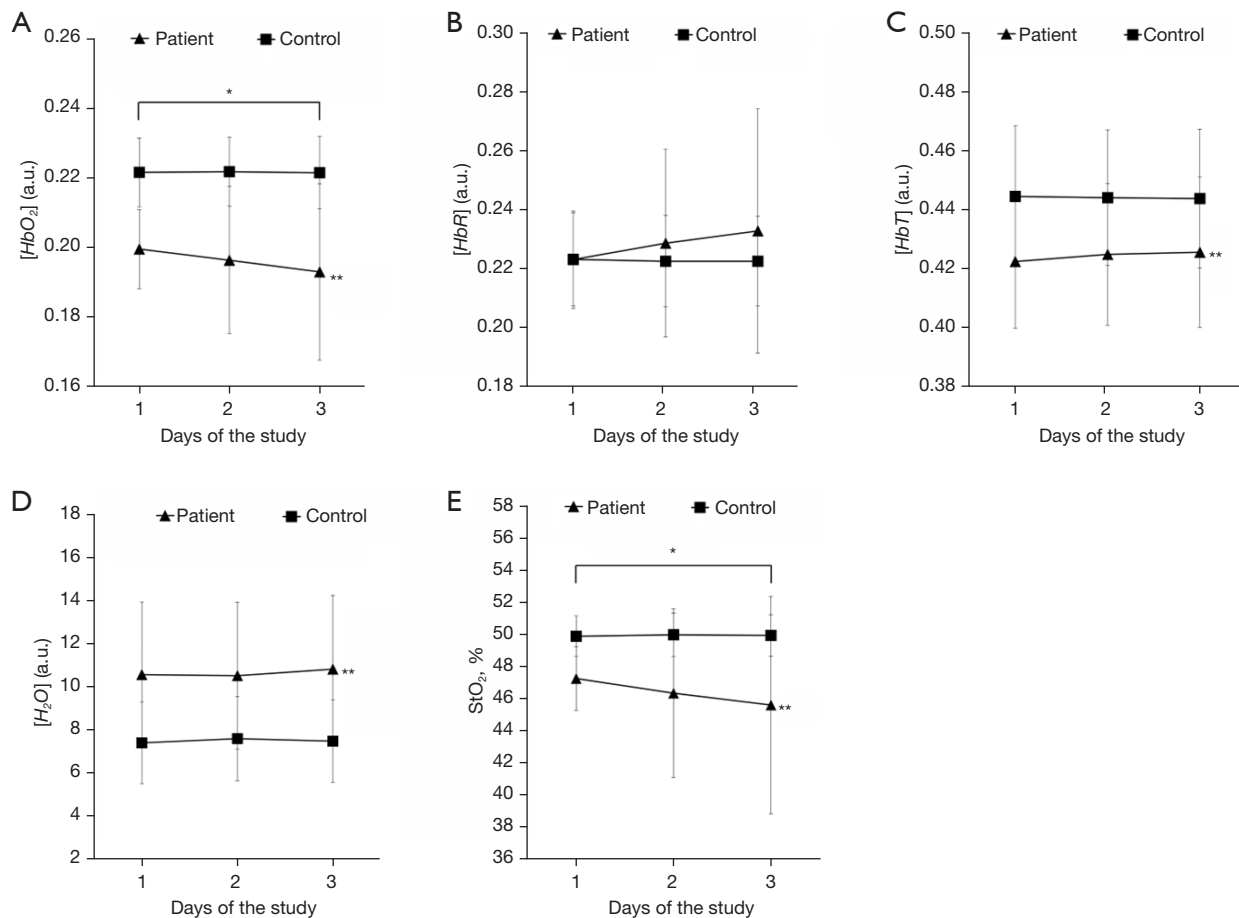
a.u., arbitrary unit; SD, standard deviation; CI, confidence interval; n, count; [HbO<sub>2</sub>], relative tissue concentration of oxy-haemoglobin; [HbR], relative tissue concentration of deoxy-haemoglobin; [HbT], relative tissue concentration of total haemoglobin; StO<sub>2</sub>, tissue haemoglobin oxygen saturation; [H<sub>2</sub>O], relative tissue concentration of H<sub>2</sub>O.

### *Association of regional tissue oxygenation and water content with systemic arterial pressure in severe sepsis*

With regard to the associations of systemic haemodynamics with regional tissue oxygenation and H<sub>2</sub>O content in patients with severe sepsis, we analysed the correlations between real-time arterial blood pressures and NIRS parameters in patients who also received pulse contour cardiac output (PiCCO) monitoring. A total of 21 patients were monitored using the PiCCO. One of them was monitored starting from day 2, and monitoring was stopped for six patients on day 3 after participating in the study. We found that only [H<sub>2</sub>O] was significantly positively correlated with systemic arterial pressure in the first-day measurement, and the correlation coefficients with systolic arterial pressure and mean arterial pressure (MAP) were 0.51 (P<0.05, Table 4 and Figure 5) and 0.45 (P<0.05, Table 4 and Figure 5), respectively. However, no significant correlation was found between tissue oxygenation and systemic arterial pressure.

### **Discussion**

Fluid administration in a fluid-responsive patient with sepsis can increase the left heart filling pressure and cardiac output, promote tissue perfusion and oxygenation, preserve organ function, and provide a survival benefit (32–34). However, the infused fluid is exchanged between the plasma and interstitium, and a large amount of fluid leaks from the capillaries and eventually accumulates in the interstitium (35). Increased vascular permeability in sepsis increases plasma volume loss, which becomes more pronounced when vasopressors are used to raise blood pressure (36,37). Therefore, the infusion may transiently affect haemodynamics but eventually cause tissue oedema and worsen tissue perfusion (38). Microcirculation improved by fluid administration was only observed in patients with severe sepsis diagnosed within 24 h (39). Liberal fluid administration can cause organ dysfunction and increase mortality risk (5,40,41). The current study used NIRS to detect tissue H<sub>2</sub>O and found that [H<sub>2</sub>O] was



**Figure 3** Time courses of relative tissue concentrations of haemoglobin and water and tissue haemoglobin oxygen saturation. (A)  $[HbO_2]$ , (B)  $[HbR]$ , (C)  $[HbT]$ , and (D)  $[H_2O]$ , and (E)  $StO_2$  measured from days 1 to 3 of the study in patients with severe sepsis and healthy controls are shown. Relative tissue concentrations of substances are expressed in arbitrary units (a.u.). Error bars represent the standard deviation of the mean. \*, GEE analysis showed that the parameters of patients with severe sepsis changed significantly over time (day 1 vs. day 3,  $P=0.041$  in  $[HbO_2]$  and  $P=0.036$  in  $StO_2$ ); \*\*, GEE analysis shows a significant difference between patients with severe sepsis and healthy controls ( $P\leq 0.001$ ).  $[HbO_2]$ , relative tissue concentration of oxy-haemoglobin;  $[HbR]$ , relative tissue concentration of deoxyhaemoglobin;  $[HbT]$ , relative tissue concentration of total haemoglobin;  $[H_2O]$ , relative tissue concentration of  $H_2O$ ;  $StO_2$ , tissue haemoglobin oxygen saturation; GEE, generalised estimating equation.

significantly higher in patients with severe sepsis than in healthy controls. Estimating  $[H_2O]$  using NIRS may offer a real-time and non-invasive quantitative assessment of tissue oedema. This parameter is potentially valuable for guiding fluid therapy in patients with severe sepsis. Further studies are needed to ascertain the relationship between fluid administration and  $[H_2O]$ .

$StO_2$  is a surrogate measure for assessing microcirculation (14,42). The value of  $StO_2$  varies depending on the measurement site (43,44). The normal gastrocnemius muscle  $StO_2$  is  $65\% \pm 19\%$  (45). Our study found that the

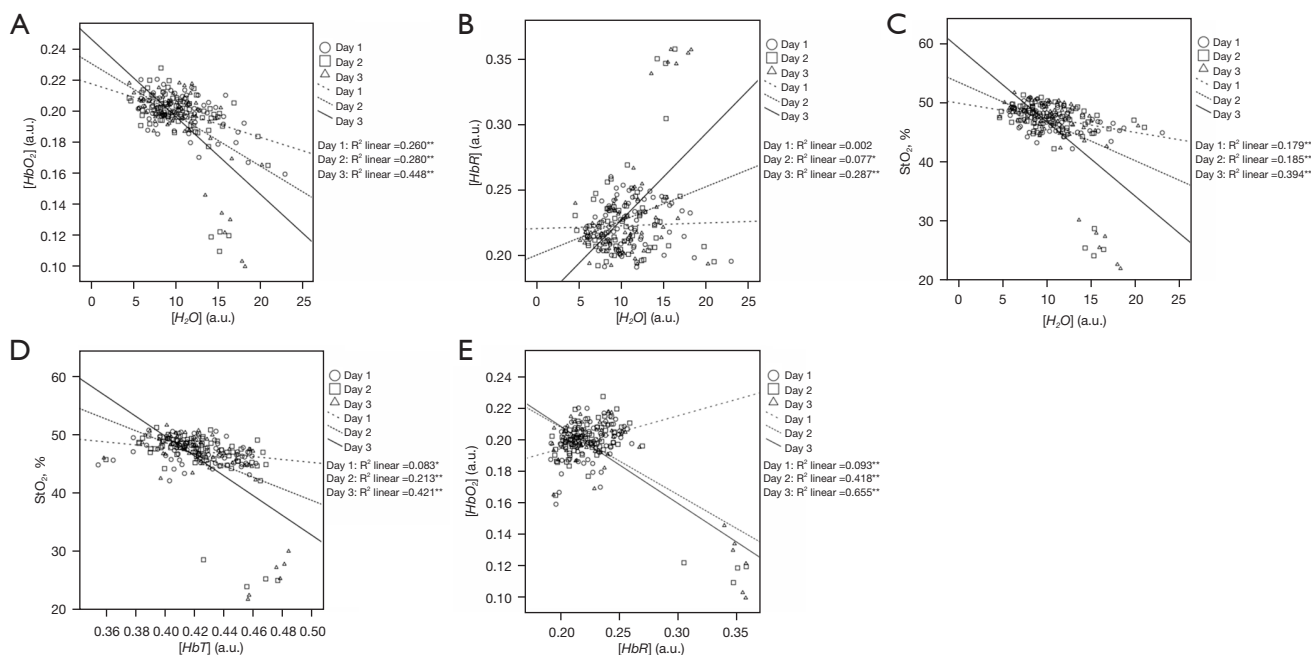
normal anterior tibial muscle  $StO_2$  was  $49.88\% \pm 1.26\%$ . Muscle  $StO_2$  changes during limb perfusion (18). Lower venous oxygen saturation and  $StO_2$  have been identified in sepsis-related hypoperfusion (19,46).  $StO_2$  is dependent on  $[HbO_2]$  and  $[HbR]$ .  $[HbO_2]$ ,  $[HbT]$ , and  $StO_2$  were significantly lower in patients with severe sepsis than in healthy controls. However, the  $[HbR]$  values were similar. In an early study, Davis and Barstow found that during exercise, changes in total tissue Hb and Mb measured by NIRS can reflect changes in microvascular hematocrit (17). In addition, NIRS-derived total Hb can indicate changes



**Table 3** Correlation matrix for relative tissue haemoglobin and water concentrations and tissue oxygenation in severe sepsis

Measurements	Days	[H <sub>2</sub> O]	[HbO <sub>2</sub> ]	[HbR]	[HbT]
[HbO <sub>2</sub> ]	1	-0.51**	-	-	-
	2	-0.53**	-	-	-
	3	-0.67**	-	-	-
[HbR]	1	0.04	0.30**	-	-
	2	0.28*	-0.65**	-	-
	3	0.54**	-0.81**	-	-
[HbT]	1	-0.22	0.72**	0.88**	-
	2	-0.10	0.02	0.75**	-
	3	0.21	-0.33**	0.82**	-
StO <sub>2</sub>	1	-0.42**	0.45**	-0.71**	-0.29*
	2	-0.43**	0.88**	-0.93**	-0.46**
	3	-0.63**	0.93**	-0.97**	-0.65**

Numbers in cells are Pearson’s correlation coefficients. \*\*P<0.01; \*P<0.05. The actual P values have been included in the <https://cdn.amegroups.com/static/public/qims-22-127-1.docx>. [HbO<sub>2</sub>], relative tissue concentration of oxy-haemoglobin; [HbR], relative tissue concentration of deoxy-haemoglobin; [HbT], relative tissue concentration of total haemoglobin; StO<sub>2</sub>, tissue haemoglobin oxygen saturation; [H<sub>2</sub>O], relative tissue concentration of H<sub>2</sub>O.

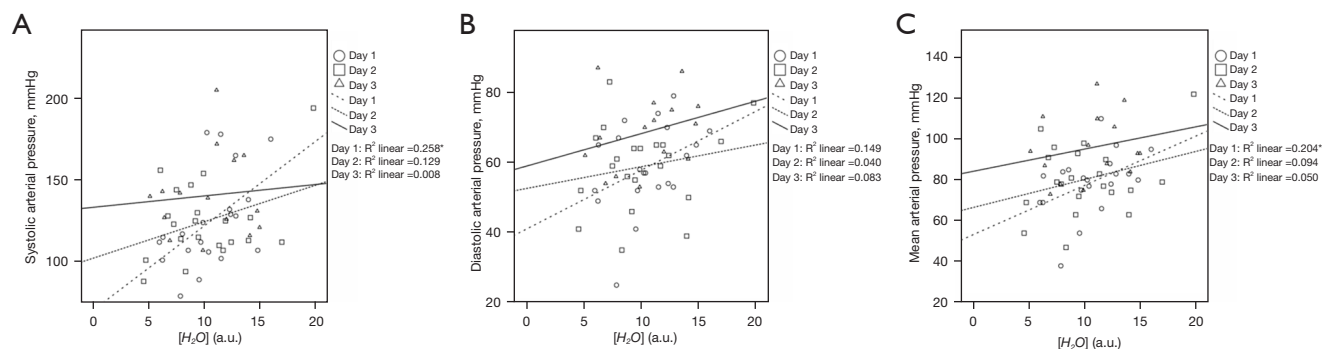


**Figure 4** A graphical representation of the relationships among the parameters of tissue oxygenation and water content. The correlations between the [H<sub>2</sub>O] and the (A) [HbO<sub>2</sub>], (B) [HbR], and (C) StO<sub>2</sub>; (D) correlation between StO<sub>2</sub> and the [HbT]; and (E) correlation between [HbO<sub>2</sub>] and [HbR] from day 1 to day 3 of the study are shown. Relative tissue concentrations of substances are expressed in arbitrary units (a.u.). \*\*P<0.01; \*P<0.05. The actual P values have been included in the <https://cdn.amegroups.com/static/public/qims-22-127-1.docx>. [H<sub>2</sub>O], relative tissue concentration of H<sub>2</sub>O; [HbO<sub>2</sub>], relative tissue concentration of oxy-haemoglobin; [HbR], relative tissue concentration of deoxyhaemoglobin; StO<sub>2</sub>, tissue haemoglobin oxygen saturation; [HbT], relative tissue concentration of total haemoglobin.

**Table 4** Correlations between arterial blood pressure and regional tissue oxygenation and water content in severe sepsis

Measurements	Days	[HbO <sub>2</sub> ]	[HbR]	[HbT]	StO <sub>2</sub>	[H <sub>2</sub> O]
Systolic arterial pressure	1	-0.15	0.23	0.11	-0.30	0.51*
	2	-0.39	-0.19	-0.29	-0.06	0.36
	3	-0.11	0.18	0.18	-0.15	0.09
Diastolic arterial pressure	1	0.09	0.25	0.24	-0.16	0.39
	2	-0.25	-0.13	-0.20	<-0.01	0.20
	3	-0.37	0.33	0.22	-0.35	0.29
Mean arterial pressure	1	-0.06	0.19	0.13	-0.20	0.45*
	2	-0.35	-0.15	-0.25	-0.06	0.31
	3	-0.26	0.30	0.25	-0.29	0.22

Numbers in cells are Pearson's correlation coefficients. \*P<0.05. The actual P values have been included in the <https://cdn.amegroups.com/static/public/qims-22-127-1.docx>. [HbO<sub>2</sub>], relative tissue concentration of oxy-haemoglobin; [HbR], relative tissue concentration of deoxy-haemoglobin; [HbT], relative tissue concentration of total haemoglobin; StO<sub>2</sub>, tissue haemoglobin oxygen saturation; [H<sub>2</sub>O], relative tissue concentration of H<sub>2</sub>O.



**Figure 5** A graphical representation of the relationships between tissue water content and systemic arterial pressures. The correlations between the [H<sub>2</sub>O] and (A) systolic arterial pressure, (B) diastolic arterial pressure, and (C) mean arterial pressure from day 1 to day 3 of the study are shown. Relative tissue concentrations of substances are expressed in arbitrary units (a.u.). \*P<0.05. The actual P values are included in the <https://cdn.amegroups.com/static/public/qims-22-127-1.docx>. [H<sub>2</sub>O], relative tissue concentration of H<sub>2</sub>O.

in muscular blood flow during exercise, measured using Doppler ultrasound (47). Therefore, [HbT] in the present study could be associated with tissue blood flow at rest. Besides, changes in HbR reflect microvascular oxygen extraction (48). Therefore, [HbR] may be related to tissue oxygen consumption (VO<sub>2</sub>) at rest. However, this does not mean that [HbR] can be a surrogate measure of VO<sub>2</sub>. Therefore, our findings suggest that patients with severe sepsis had reduced limb perfusion, resulting in lower [HbT]. Nevertheless, their peripheral skeletal muscle VO<sub>2</sub> might remain unchanged, showing consistent [HbR] values in healthy controls at rest. [HbO<sub>2</sub>] and StO<sub>2</sub> decreased with increasing oxygen extraction ratio (O<sub>2</sub>ER). To date, whether

muscle VO<sub>2</sub> increases in patients with sepsis remains controversial (49,50). VO<sub>2</sub> is physiologically dependent on perfusive oxygen delivery (QO<sub>2</sub>) below critical QO<sub>2</sub> (51). QO<sub>2</sub> depends on blood flow, the product of heart rate and cardiac output, and O<sub>2</sub> concentrations in arterial and venous blood (15). Therefore, patients with an obvious decrease in blood flow have a physiological dependence of VO<sub>2</sub> on QO<sub>2</sub>. However, whether pathological dependence of VO<sub>2</sub> on QO<sub>2</sub> occurs in septic patients has not yet been elucidated (52). Our findings are consistent with the findings of Meneguetti *et al.*, who found that VO<sub>2</sub> was not increased in patients with sepsis (53). Efforts to reduce metabolic demands from skeletal muscles using neuromuscular blockade do not alter

$VO_2$ ,  $QO_2$ , and  $O_2ER$  in septic patients (54). The above findings explain why maintaining supranormal  $QO_2$  may not benefit critically ill patients, as tissue oxygen demand may not be increased (55-57). Our study suggests that identifying the individual  $[HbT]$  and  $[HbR]$  using NIRS is crucial for non-invasively and indirectly understanding patients' tissue perfusion and  $VO_2$ . These data may help determine the peripheral tissue response to altered systemic haemodynamics using inotropic agents and fluid resuscitation in patients with severe sepsis.

As an estimation of tissue perfusion and oxygenation,  $StO_2$  is correlated with  $QO_2$ , responds to the ischaemic challenge, and may be associated with survival outcomes in patients with severe sepsis (19-21,58,59). Nevertheless, the clinical significance of  $StO_2$  remains controversial. Baseline  $StO_2$  cannot be used for the early detection of severe sepsis, and the routine implementation of resuscitation protocols incorporating  $StO_2 >80\%$  as a target does not provide a survival benefit (60,61). An important reason for these controversial findings is that  $StO_2$  is confounded by several factors, including tissue oedema (23). Compared with baseline  $StO_2$ , the measurement of the change in  $StO_2$  reperfusion slope during a vascular occlusion test (VOT) may eliminate the personal confounding factor of tissue oedema and is significantly related to microcirculation in patients with septic shock and the outcome of septic patients (46,62). However, VOT is not convenient for continuous real-time monitoring of microcirculation. In the present study, we found that  $StO_2$  was significantly negatively correlated with  $[H_2O]$ , which resulted from the negative correlation between  $[H_2O]$  and  $[HbO_2]$ . In addition, the relationship between  $[HbO_2]$ ,  $[HbR]$ , and  $StO_2$  and  $[H_2O]$  became more relevant during the study period, suggesting that the impact of tissue  $H_2O$  on tissue oxygenation may become increasingly apparent during ICU admission. Our study determined that regional tissue  $H_2O$  interacts with tissue  $HbO_2$  and  $StO_2$ . It remains unclear whether the oedematous interstitium physiologically leads to reduced regional tissue perfusion, thereby altering tissue oxygenation. More research is warranted to clarify the detailed mechanism by which tissue  $H_2O$  affects tissue perfusion and oxygenation, and whether fluid administration for normalising unstable haemodynamics in septic shock causes tissue oedema, leading to tissue perfusion deterioration.

In sepsis-induced hypoperfusion, additional fluid administration after the initial resuscitation should be guided by frequent reassessments of the patient's

haemodynamic status (3). Most measurements are obtained by monitoring systemic haemodynamics, and systemic arterial pressure is one of the most commonly measured parameters. The present study found that the regional tissue oxygenation parameters, including  $[HbO_2]$ ,  $[HbR]$ , and  $StO_2$ , were not related to the systemic arterial pressure in the early stage of ICU admission. Our findings seem to be consistent with the concept of loss of haemodynamic coherence between the macrocirculation and microcirculation in sepsis (6). Systemic arterial pressure poorly reflects regional tissue perfusion (63). In addition, microcirculation is profoundly disturbed in severe sepsis, and the organ tissue blood supply may not reflect tissue oxygenation (64). Meanwhile, as mentioned above, fluid volume expansion to increase systemic arterial pressure also increases intravascular hydrostatic pressure, which may exacerbate fluid accumulation and result in tissue oedema (65). The oedematous interstitium reduces capillary perfusion, which in turn worsens tissue oxygenation (66). This may explain why systemic arterial pressure was positively correlated with tissue  $H_2O$  but was not related to tissue oxygenation in our study. Targeting an MAP level  $>65$  mmHg is a general goal in the initial resuscitation of patients with septic shock (3). Jozwiak *et al.* recently found that the impact of a unique MAP target on peripheral oxygenation may differ widely among patients with septic shock (9). In this study, patients with severe sepsis were enrolled after ICU admission. Most of them had received initial fluid resuscitation for normalising systemic haemodynamics at the ER. Attempts to increase MAP levels early in ICU admission may not guarantee improved tissue oxygenation but may result in increased tissue  $H_2O$ . These findings suggest that care should be taken when administering fluid to normalise systemic blood pressure during early ICU admission, particularly if initial fluid resuscitation has already been performed. Initial fluid resuscitation may improve sepsis-induced hypoperfusion and improve patient survival (4). However, continued fluid administration following initial resuscitation may be harmful and should be guided by careful assessment of intravascular volume status and organ perfusion (67). Tissue  $H_2O$  can be measured using NIRS and is potentially helpful in appreciating the effectiveness of fluid therapy on regional tissue perfusion and oxygenation.

This study has some limitations. First, the estimated sample size required 70 people per group, but the healthy control group comprised only 30 people, reducing the statistical power. In addition, healthy controls had a lower mean age than those with severe sepsis. Therefore,

differences in NIRS parameters may be attributed to differences in age. However, Rosenberry *et al.* recently found that age is related to NIRS-derived post-occlusion StO<sub>2</sub> recovery kinetics, but not baseline forearm StO<sub>2</sub> (68). Therefore, severe sepsis remained a significant factor contributing to the difference in NIRS parameters between the two groups. Another limitation is the lack of a multivariate analysis investigating the causal relationship between [H<sub>2</sub>O] and tissue oxygenation parameters. Correlation analysis offers only crude assessments of the linear relationships between [H<sub>2</sub>O] and tissue oxygenation parameters. The same limitation exists in the findings on the relationship between systemic arterial pressure and [H<sub>2</sub>O]. However, the results provide evidence for the interaction of tissue H<sub>2</sub>O with tissue Hb and StO<sub>2</sub> used to assess tissue perfusion and oxygenation. Compared with baseline StO<sub>2</sub>, dynamic NIRS measurements are more relevant to sepsis-related microvascular dysfunction (19,69). However, we did not investigate the dynamic changes in [HbO<sub>2</sub>], [HbR], and [H<sub>2</sub>O] during VOT and their relationship to the StO<sub>2</sub> recovery slope. This crucial issue will be investigated and examined in future studies.

## Conclusions

A NIRS device transmitting tri-wavelength light can be used to concurrently assess regional tissue oxygenation and H<sub>2</sub>O content. The regional tissue H<sub>2</sub>O content was significantly increased in patients with severe sepsis. In the early phase of severe sepsis, elevated systemic arterial pressure may be related to increased regional tissue H<sub>2</sub>O, but not to tissue oxygenation. Therefore, the measurement of tissue H<sub>2</sub>O is crucial and should be considered when estimating microcirculation and tissue oxygenation in patients with severe sepsis. Further studies are needed to elucidate the physiological effect of tissue H<sub>2</sub>O on tissue oxygenation and whether it can be accurately measured.

## Acknowledgments

*Funding:* This work was supported by the Chang Gung Medical Foundation of Taiwan (Nos. CORPG6E0111, CORPG6E0112, and CORPG6E0113 to Chin-Kuo Lin).

## Footnote

*Reporting Checklist:* The authors have completed the STROBE reporting checklist. Available at <https://qims.amegroups.com/article/view/10.21037/qims-22-127/rc>

[amegroups.com/article/view/10.21037/qims-22-127/rc](https://qims.amegroups.com/article/view/10.21037/qims-22-127/rc)

*Conflicts of Interest:* All authors have completed the ICMJE uniform disclosure form (available at <https://qims.amegroups.com/article/view/10.21037/qims-22-127/coif>). CKL received grants from the Chang Gung Medical Foundation of Taiwan (Nos. CORPG6E0111, CORPG6E0112, and CORPG6E0113) for the study. The other authors have no conflicts of interest to declare.

*Ethical Statement:* The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by the Institutional Review Board of Chang Gung Medical Foundation (No. 103-5357B). Informed consent was obtained from the healthy volunteers and patients. Legal guardians provided informed consent to participate if the patient had cognitive impairment.

*Open Access Statement:* This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

## References

1. Singer M, Deutschman CS, Seymour CW, Shankar-Hari M, Annane D, Bauer M, Bellomo R, Bernard GR, Chiche JD, Cooper-Smith CM, Hotchkiss RS, Levy MM, Marshall JC, Martin GS, Opal SM, Rubenfeld GD, van der Poll T, Vincent JL, Angus DC. The Third International Consensus Definitions for Sepsis and Septic Shock (Sepsis-3). *JAMA* 2016;315:801-10.
2. Astiz ME, Rackow EC, Falk JL, Kaufman BS, Weil MH. Oxygen delivery and consumption in patients with hyperdynamic septic shock. *Crit Care Med* 1987;15:26-8.
3. Rhodes A, Evans LE, Alhazzani W, Levy MM, Antonelli M, Ferrer R, et al. Surviving Sepsis Campaign: International Guidelines for Management of Sepsis and Septic Shock: 2016. *Intensive Care Med* 2017;43:304-77.

4. Evans L, Rhodes A, Alhazzani W, Antonelli M, Coopersmith CM, French C, et al. Surviving Sepsis Campaign: International Guidelines for Management of Sepsis and Septic Shock 2021. *Crit Care Med* 2021;49:e1063-143.
5. Jaffee W, Hodgins S, McGee WT. Tissue Edema, Fluid Balance, and Patient Outcomes in Severe Sepsis: An Organ Systems Review. *J Intensive Care Med* 2018;33:502-9.
6. Ince C. Hemodynamic coherence and the rationale for monitoring the microcirculation. *Crit Care* 2015;19 Suppl 3:S8.
7. De Backer D, Creteur J, Preiser JC, Dubois MJ, Vincent JL. Microvascular blood flow is altered in patients with sepsis. *Am J Respir Crit Care Med* 2002;166:98-104.
8. Boerma EC, van der Voort PH, Spronk PE, Ince C. Relationship between sublingual and intestinal microcirculatory perfusion in patients with abdominal sepsis. *Crit Care Med* 2007;35:1055-60.
9. Jozwiak M, Chambaz M, Sentenac P, Monnet X, Teboul JL. Assessment of tissue oxygenation to personalize mean arterial pressure target in patients with septic shock. *Microvasc Res* 2020;132:104068.
10. Lam C, Tyml K, Martin C, Sibbald W. Microvascular perfusion is impaired in a rat model of normotensive sepsis. *J Clin Invest* 1994;94:2077-83.
11. Farquhar I, Martin CM, Lam C, Potter R, Ellis CG, Sibbald WJ. Decreased capillary density in vivo in bowel mucosa of rats with normotensive sepsis. *J Surg Res* 1996;61:190-6.
12. Bakker J, Ince C. Monitoring coherence between the macro and microcirculation in septic shock. *Curr Opin Crit Care* 2020;26:267-72.
13. Valeanu L, Bubenek-Turconi SI, Ginghina C, Balan C. Hemodynamic Monitoring in Sepsis-A Conceptual Framework of Macro- and Microcirculatory Alterations. *Diagnostics (Basel)* 2021;11:1559.
14. Charlton M, Sims M, Coats T, Thompson JP. The microcirculation and its measurement in sepsis. *J Intensive Care Soc* 2017;18:221-7.
15. Barstow TJ. Understanding near infrared spectroscopy and its application to skeletal muscle research. *J Appl Physiol (1985)* 2019;126:1360-76.
16. Jöbsis FF. Noninvasive, infrared monitoring of cerebral and myocardial oxygen sufficiency and circulatory parameters. *Science* 1977;198:1264-7.
17. Davis ML, Barstow TJ. Estimated contribution of hemoglobin and myoglobin to near infrared spectroscopy. *Respir Physiol Neurobiol* 2013;186:180-7.
18. Mancini DM, Bolinger L, Li H, Kendrick K, Chance B, Wilson JR. Validation of near-infrared spectroscopy in humans. *J Appl Physiol (1985)* 1994;77:2740-7.
19. Creteur J, Carollo T, Soldati G, Buchele G, De Backer D, Vincent JL. The prognostic value of muscle StO<sub>2</sub> in septic patients. *Intensive Care Med* 2007;33:1549-56.
20. Leone M, Bliidi S, Antonini F, Meyssignac B, Bordon S, Garcin F, Charvet A, Blasco V, Albanèse J, Martin C. Oxygen tissue saturation is lower in nonsurvivors than in survivors after early resuscitation of septic shock. *Anesthesiology* 2009;111:366-71.
21. Colin G, Nardi O, Polito A, Aboab J, Maxime V, Clair B, Friedman D, Orlikowski D, Sharshar T, Annane D. Masseter tissue oxygen saturation predicts normal central venous oxygen saturation during early goal-directed therapy and predicts mortality in patients with severe sepsis. *Crit Care Med* 2012;40:435-40.
22. Lee WL, Slutsky AS. Sepsis and endothelial permeability. *N Engl J Med* 2010;363:689-91.
23. Poeze M. Tissue-oxygenation assessment using near-infrared spectroscopy during severe sepsis: confounding effects of tissue edema on StO<sub>2</sub> values. *Intensive Care Med* 2006;32:788-9.
24. Kuo HC, Lo CC, Lin PX, Kao CC, Huang YH, Lin BS. Wireless optical monitoring system identifies limb induration characteristics in patients with Kawasaki disease. *J Allergy Clin Immunol* 2018;142:710-1.
25. Wang L, Jacques SL, Zheng L. MCML--Monte Carlo modeling of light transport in multi-layered tissues. *Comput Methods Programs Biomed* 1995;47:131-46.
26. Bashkatov AN, Genina EA, Kochubey VI, Tuchin VV. Optical properties of human skin, subcutaneous and mucous tissues in the wavelength range from 400 to 2000 nm. *Journal of Physics D: Applied Physics* 2005;38:2543.
27. Molavi B, Shadgan B, Macnab AJ, Dumont GA. Noninvasive Optical Monitoring of Bladder Filling to Capacity Using a Wireless Near Infrared Spectroscopy Device. *IEEE Trans Biomed Circuits Syst* 2014;8:325-33.
28. Cope M, Delpy DT, Reynolds EO, Wray S, Wyatt J, van der Zee P. Methods of quantitating cerebral near infrared spectroscopy data. *Adv Exp Med Biol* 1988;222:183-9.
29. Delpy DT, Cope M, van der Zee P, Arridge S, Wray S, Wyatt J. Estimation of optical pathlength through tissue from direct time of flight measurement. *Phys Med Biol* 1988;33:1433-42.
30. Dellinger RP, Levy MM, Rhodes A, Annane D, Gerlach H, Opal SM, et al. Surviving sepsis campaign: international guidelines for management of severe sepsis and septic

- shock: 2012. *Crit Care Med* 2013;41:580-637.
31. Sadaka F, Aggu-Sher R, Krause K, O'Brien J, Armbrrecht ES, Taylor RW. The effect of red blood cell transfusion on tissue oxygenation and microcirculation in severe septic patients. *Ann Intensive Care* 2011;1:46.
  32. Packman MI, Rackow EC. Optimum left heart filling pressure during fluid resuscitation of patients with hypovolemic and septic shock. *Crit Care Med* 1983;11:165-9.
  33. Kaufman BS, Rackow EC, Falk JL. The relationship between oxygen delivery and consumption during fluid resuscitation of hypovolemic and septic shock. *Chest* 1984;85:336-40.
  34. Molnár Z, Mikor A, Leiner T, Szakmány T. Fluid resuscitation with colloids of different molecular weight in septic shock. *Intensive Care Med* 2004;30:1356-60.
  35. Hahn RG, Lyons G. The half-life of infusion fluids: An educational review. *Eur J Anaesthesiol* 2016;33:475-82.
  36. Groeneveld AB, Teule GJ, Bronsveld W, van den Bos GC, Thijs LG. Increased systemic microvascular albumin flux in septic shock. *Intensive Care Med* 1987;13:140-2.
  37. Dubniks M, Persson J, Grände PO. Effect of blood pressure on plasma volume loss in the rat under increased permeability. *Intensive Care Med* 2007;33:2192-8.
  38. Armistead CW Jr, Vincent JL, Preiser JC, De Backer D, Thuc Le Minh. Hypertonic saline solution-hetastarch for fluid resuscitation in experimental septic shock. *Anesth Analg* 1989;69:714-20.
  39. Ospina-Tascon G, Neves AP, Occhipinti G, Donadello K, Büchele G, Simion D, Chiarego ML, Silva TO, Fonseca A, Vincent JL, De Backer D. Effects of fluids on microvascular perfusion in patients with severe sepsis. *Intensive Care Med* 2010;36:949-55.
  40. Maitland K, Kiguli S, Opoka RO, Engoru C, Olupot-Olupot P, Akech SO, Nyeko R, Mtove G, Reyburn H, Lang T, Brent B, Evans JA, Tibenderana JK, Crawley J, Russell EC, Levin M, Babiker AG, Gibb DM; FEAST Trial Group. Mortality after fluid bolus in African children with severe infection. *N Engl J Med* 2011;364:2483-95.
  41. Sirvent JM, Ferri C, Baró A, Murcia C, Lorencio C. Fluid balance in sepsis and septic shock as a determining factor of mortality. *Am J Emerg Med* 2015;33:186-9.
  42. Mesquida J, Gruartmoner G, Espinal C. Skeletal muscle oxygen saturation (StO<sub>2</sub>) measured by near-infrared spectroscopy in the critically ill patients. *Biomed Res Int* 2013;2013:502194.
  43. Beilman GJ, Groehler KE, Lazon V, Ortner JP. Near-infrared spectroscopy measurement of regional tissue oxyhemoglobin saturation during hemorrhagic shock. *Shock* 1999;12:196-200.
  44. Nagashima Y, Yada Y, Hattori M, Sakai A. Development of a new instrument to measure oxygen saturation and total hemoglobin volume in local skin by near-infrared spectroscopy and its clinical application. *Int J Biometeorol* 2000;44:11-9.
  45. Comerota AJ, Throm RC, Kelly P, Jaff M. Tissue (muscle) oxygen saturation (StO<sub>2</sub>): a new measure of symptomatic lower-extremity arterial disease. *J Vasc Surg* 2003;38:724-9.
  46. Neto AS, Pereira VG, Manetta JA, Espósito DC, Schultz MJ. Association between static and dynamic thenar near-infrared spectroscopy and mortality in patients with sepsis: a systematic review and meta-analysis. *J Trauma Acute Care Surg* 2014;76:226-33.
  47. Alvares TS, Oliveira GV, Soares R, Murias JM. Near-infrared spectroscopy-derived total haemoglobin as an indicator of changes in muscle blood flow during exercise-induced hyperaemia. *J Sports Sci* 2020;38:751-8.
  48. Hammer SM, Alexander AM, Didier KD, Smith JR, Caldwell JT, Sutterfield SL, Ade CJ, Barstow TJ. The noninvasive simultaneous measurement of tissue oxygenation and microvascular hemodynamics during incremental handgrip exercise. *J Appl Physiol* (1985) 2018;124:604-14.
  49. Girardis M, Rinaldi L, Busani S, Flore I, Mauro S, Pasetto A. Muscle perfusion and oxygen consumption by near-infrared spectroscopy in septic-shock and non-septic-shock patients. *Intensive Care Med* 2003;29:1173-6.
  50. De Blasi RA, Palmisani S, Alampi D, Mercieri M, Romano R, Collini S, Pinto G. Microvascular dysfunction and skeletal muscle oxygenation assessed by phase-modulation near-infrared spectroscopy in patients with septic shock. *Intensive Care Med* 2005;31:1661-8.
  51. Baigorri F, Russell JA. Oxygen delivery in critical illness. *Crit Care Clin* 1996;12:971-94.
  52. Russell JA, Phang PT. The oxygen delivery/consumption controversy. Approaches to management of the critically ill. *Am J Respir Crit Care Med* 1994;149:533-7.
  53. Meneguetti MG, de Araújo TR, Laus AM, Martins-Filho OA, Basile-Filho A, Auxiliadora-Martins M. Resting Energy Expenditure and Oxygen Consumption in Critically Ill Patients With vs Without Sepsis. *Am J Crit Care* 2019;28:136-41.
  54. Freebairn RC, Derrick J, Gomersall CD, Young RJ, Joynt GM. Oxygen delivery, oxygen consumption, and gastric intramucosal pH are not improved by a computer-

- controlled, closed-loop, vecuronium infusion in severe sepsis and septic shock. *Crit Care Med* 1997;25:72-7.
55. Gutierrez G, Palizas F, Doglio G, Wainsztein N, Gallesio A, Pacin J, Dubin A, Schiavi E, Jorge M, Pusajo J. Gastric intramucosal pH as a therapeutic index of tissue oxygenation in critically ill patients. *Lancet* 1992;339:195-9.
  56. Yu M, Levy MM, Smith P, Takiguchi SA, Miyasaki A, Myers SA. Effect of maximizing oxygen delivery on morbidity and mortality rates in critically ill patients: a prospective, randomized, controlled study. *Crit Care Med* 1993;21:830-8.
  57. Russell JA. Adding fuel to the fire--the supranormal oxygen delivery trials controversy. *Crit Care Med* 1998;26:981-3.
  58. Mesquida J, Gruartmoner G, Martínez ML, Masip J, Sabatier C, Espinal C, Artigas A, Baigorri F. Thenar oxygen saturation and invasive oxygen delivery measurements in critically ill patients in early septic shock. *Shock* 2011;35:456-9.
  59. Vorwerk C, Coats TJ. The prognostic value of tissue oxygen saturation in emergency department patients with severe sepsis or septic shock. *Emerg Med J* 2012;29:699-703.
  60. Goulet H, André S, Sahakian GD, Freund Y, Khelifi G, Claessens YE, Riou B, Ray P. Accuracy of oxygen tissue saturation values in assessing severity in patients with sepsis admitted to emergency departments. *Eur J Emerg Med* 2014;21:266-71.
  61. Nardi O, Zavala E, Martin C, Nanas S, Scheeren T, Polito A, Borrat X, Annane D. Targeting skeletal muscle tissue oxygenation (StO<sub>2</sub>) in adults with severe sepsis and septic shock: a randomised controlled trial (OTO-StS Study). *BMJ Open* 2018;8:e017581.
  62. Payen D, Luengo C, Heyer L, Resche-Rigon M, Kerever S, Damoiseil C, Losser MR. Is thenar tissue hemoglobin oxygen saturation in septic shock related to macrohemodynamic variables and outcome? *Crit Care* 2009;13 Suppl 5:S6.
  63. Pinsky MR, Payen D. Functional hemodynamic monitoring. *Crit Care* 2005;9:566-72.
  64. Østergaard L, Granfeldt A, Secher N, Tietze A, Iversen NK, Jensen MS, Andersen KK, Nagenthiraja K, Gutiérrez-Lizardi P, Mouridsen K, Jespersen SN, Tønnesen EK. Microcirculatory dysfunction and tissue oxygenation in critical illness. *Acta Anaesthesiol Scand* 2015;59:1246-59.
  65. Obonyo NG, Fanning JP, Ng AS, Pimenta LP, Shekar K, Platts DG, Maitland K, Fraser JF. Effects of volume resuscitation on the microcirculation in animal models of lipopolysaccharide sepsis: a systematic review. *Intensive Care Med Exp* 2016;4:38.
  66. Jerome SN, Akimitsu T, Korthuis RJ. Leukocyte adhesion, edema, and development of postischemic capillary no-reflow. *Am J Physiol* 1994;267:H1329-36.
  67. Malbrain MLNG, Langer T, Annane D, Gattinoni L, Elbers P, Hahn RG, De Laet I, Minini A, Wong A, Ince C, Muckart D, Mythen M, Caironi P, Van Regenmortel N. Intravenous fluid therapy in the perioperative and critical care setting: Executive summary of the International Fluid Academy (IFA). *Ann Intensive Care* 2020;10:64.
  68. Rosenberry R, Munson M, Chung S, Samuel TJ, Patik J, Tucker WJ, Haykowsky MJ, Nelson MD. Age-related microvascular dysfunction: novel insight from near-infrared spectroscopy. *Exp Physiol* 2018;103:190-200.
  69. Skarda DE, Mulier KE, Myers DE, Taylor JH, Beilman GJ. Dynamic near-infrared spectroscopy measurements in patients with severe sepsis. *Shock* 2007;27:348-53.

**Cite this article as:** Lin CK, Leu SW, Tsai YH, Zhou SK, Lin CM, Huang SY, Chang CC, Ho MC, Lee WC, Chen MC, Hung MS, Lin YC, Li JR, Lin BS. Increased tissue water in patients with severe sepsis affects tissue oxygenation measured by near-infrared spectroscopy: a prospective, observational case-control study. *Quant Imaging Med Surg* 2022;12(10):4953-4967. doi: 10.21037/qims-22-127