



# Highly homogeneous microbial communities dominated by *Mycoplasma pneumoniae* instead of increased resistance to macrolide antibiotics is the characteristic of lower respiratory tract microbiome of children with refractory *Mycoplasma pneumoniae* pneumonia

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**Background:** Although researchers have found that the microbiota changed during the lower respiratory tract (LRT) infection, little was known about the association between LRT microbiome and refractory *M. pneumoniae* pneumonia (RMPP).

**Methods:** From June 28th, 2019 to March 23rd, 2020, we enrolled fifty-two children diagnosed with RMPP or non-refractory *M. pneumoniae* pneumonia (NRMPP), and characterized the structure and function of microbiota in the bronchoalveolar lavage fluid (BALF) by metagenomic next generation sequencing (mNGS).

**Results:** Based on Bray-Curtis distance between samples, samples in RMPP group were highly homogeneous, and Shannon index in the RMPP group was much lower than NRMPP group while Simpson index, which presents the degree of dominance, was higher in RMPP group. The dominant taxon with relative abundance greater than 50% was merely *Mycoplasma* among RMPP and NRMPP patients, but the proportions of other taxonomic distribution were different. *M. pneumoniae* was the dominant species and occupied almost all niches in the vast majority of RMPP patients, whereas the other genera were dramatically lower. The NRMPP group was more enriched in antibiotic resistance genes (ARGs) than the RMPP group, and also exhibited a greater relative abundance of macrolide antibiotics resistance gene (*macB*) and fluoroquinolone antibiotic resistance genes (*patA-B*) in *M. pneumoniae* genome. In RMPP patients, higher relative abundance of *Streptococcus pneumoniae* had a strong correlation with increased hospitalization days while higher relative abundance of *Streptococcus pneumoniae* had a negative correlation with hospitalization days among NRMPP patients.

**Conclusions:** The microbiota of LRT in children with RMPP was much more homogeneous and simpler than that of the NRMPP group and with lower relative abundance of macrolide antibiotics resistance gene in *M. pneumoniae* genome. *M. pneumoniae* was absolutely dominant in the vast majority of RMPP patients. Prolonged hospitalization days was associated with relative abundance of *M. pneumoniae* in NRMPP patients while it was related with other pathogens' relative abundance (e.g., *Streptococcus pneumoniae*) in RMPP patients.

**Keywords:** Refractory pneumonia; *Mycoplasma pneumoniae* (*M. pneumoniae*); microbiome; lower respiratory tract (LRT); children

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## Introduction

*Mycoplasma pneumoniae* (*M. pneumoniae*) is an important pathogen of community-acquired pneumonia in children. For the past few years, because of the emergence of *M. pneumoniae* resistance to macrolides and a series of complex mechanisms, the incidence of refractory *M. pneumoniae* pneumonia (RMPP) has increased, often leading to poor prognosis and sequelae such as atelectasis, pulmonary necrosis, bronchiectasis, and bronchiolitis obliterans (1-3).

Researchers found that the microbiota changed significantly during the lower respiratory tract (LRT) infection and was closely related to the prognosis or course of pneumonia (4,5). However, the structure and function of LRT microbiota in patients with RMPP remained unexplored. We reasoned that characterization of LRT microbiota in children with RMPP may provide us with opportunities to uncover the ecosystem of the LRT, which can lead to the mechanism of aggravation of non-refractory *M. pneumoniae* pneumonia (NRMPP) to RMPP, and possible avenues for the prevention of RMPP.

Here, a prospective study was conducted in the Children's Hospital of Fudan University to characterize the microbiota in LRT of RMPP and NRMPP children as well as its function and to reveal the correlation between microbiota and laboratory tests.

We present the following article in accordance with the MDAR checklist (available at <http://dx.doi.org/10.21037/tp-20-404>).

## Methods

### *Patient enrollment and study design*

During the period between June 28, 2019 and March 23, 2020, we prospectively enrolled 52 children admitted to the hospitalization zone in the Division of Pulmonary Medicine, Children's Hospital of Fudan University, with clinically diagnosed NRMPP or RMPP, and collected bronchoalveolar lavage fluid (BALF) when bronchoscopy was necessary. The inclusion criteria were (I) diagnosed with pneumonia and meet the hospitalization standard of community-acquired pneumonia in children (has one of

the following): Under the condition of breathing air, RR >50 times/min; under the condition of breathing air, SaO<sub>2</sub> ≤0.92; dyspnea: chest wall inspiratory depression, flaring of alae nasi; intermittent apnea; persistent high fever for 3-5 days; chest X-ray and other imaging data confirmed bilateral or multiple lobes involvement or atelectasis, pleural effusion or short-term progression (6); (II) *M. pneumoniae* infection, which was confirmed by conducting a serological test (*M. pneumoniae*-IgM-positive and an antibody titer ≥1:160 or a ≥4-fold increase in the titer) and by detection of *M. pneumoniae* nucleic acid in the nasopharyngeal aspirate, sputum and BALF (7); (III) bronchoscopy, which was routinely performed when patients had more than one lobe of pulmonary consolidation or atelectasis by X-ray or CT scan. RMPP was defined as clinical symptoms and the pulmonary imaging of the pneumonia aggravated after regular treatment with macrolide antibiotics for more than seven days. The exclusion criteria were (I) the detection of any other pathogens in the patients' blood, nasopharyngeal aspirate, sputum or BALF via culture, viral antigen detection assays (adenovirus, respiratory syncytial virus, metapneumovirus, influenza, and parainfluenza), interferon-γ release assays, and T cell spot tests (T-SPOTs); (II) children who combined with other respiratory diseases, blood system disease, autoimmune disease or with organ dysfunction and in the acute stage of allergic disease were also excluded.

The ethical application and the consent procedure of this study were approved by the Ethics Committee of Children's Hospital of Fudan University (No. 2016-87). All procedures performed in this study involving human participants were in accordance with the Declaration of Helsinki (as revised in 2013), and informed consent was taken from all the parents.

### *Clinical characteristics*

Clinical information was collected from the medical records of the patients, and the age, sex, hospitalization days and antibiotic treatments before bronchoscopy were recorded. The maximum levels of the laboratory data were recorded before bronchoscopy was performed, which included white blood cell (WBC) counts, levels of C-reactive protein (CRP),

fibrinogen, lactate dehydrogenase (LDH) and D-dimer.

### **Sample collection**

Bronchoscopy was done by using standard methods and conscious intravenous sedation with midazolam. After topical anesthesia of the larynx, trachea, and carina was achieved with 2% lidocaine (Sanchine, China), the bronchoscope (Olympus, Japan) was wedged in the lesion's segment or lobe and BALF was performed with three aliquots of sterile saline (Baxter, China), 1 ml/kg each, followed by suction pressure of 100 mmHg. All BALF samples were then immediately frozen in a dry liquid nitrogen Dewar (ASONE, Japan) and stored at  $-80^{\circ}\text{C}$  until analysis.

### **Nucleic acid extraction**

BALF samples were collected as described above. DNA was extracted from BALF, using the TIANamp Magnetic DNA Kit (Tiangen, China) according to the manufacturer's protocols. The quantity and quality of DNA was assessed using the Qubit (Thermo Fisher Scientific, USA) and the NanoDrop (Thermo Fisher Scientific, USA), respectively. No template control (NTC) of each batch were also processed with clinical samples.

### **Library preparation and sequencing**

DNA libraries were prepared using the KAPA Hyper Prep kit (KAPA Biosystems, USA) according to the manufacturer's protocols, by fragmenting DNA into 150–300 bp. DNA libraries were qualified by Agilent 2100, and 75 bp single-ends sequenced on Illumina NextSeq 550Dx (Illumina, USA).

### **Bioinformatics analysis**

Raw sequencing data was split by *bcl2fastq2*, and high-quality sequencing data were generated using Trimmomatic by removing low quality reads, adapter contamination, and duplicated and short (length <35 bp) reads. Human host sequences were subtracted by mapping to human reference genome (hs37d5) using *bowtie2*. Reads that could not be mapped to the human genome were retained and aligned with microorganism genome database for microbial identification by Kraken, and for species abundance estimation by Bracken. The microorganism genome

database contained genomes or scaffolds of bacteria, fungi, viruses, and parasites (download from <ftp://ftp.ncbi.nlm.nih.gov/genomes/genbank/>). We used the following criteria for positive results of metagenomic next generation sequencing (mNGS): For *Mycobacterium*, *Nocardia* and *Legionella pneumophila*, the result was considered positive if a species detected by mNGS had a species-specific read number  $\geq 1$ . For bacteria (excluding *Mycobacterium*, *Nocardia* and *Legionella pneumophila*), fungi, virus and parasites, the result was considered positive if a species detected by mNGS had at least 3 non-overlapping reads. Pathogens detected in the negative NTC were excluded but only if the detected reads were  $\geq 10$ -fold than that in the NTC. Functional metagenome profiling was calculated using HUMAnN2 standard workflow with default settings. Tables of normalized gene family were converted into KEGG Orthology terms (KO) using *humann2\_regroup\_table* command of HUMAnN2. Normalized KO were then mapped to KEGG pathways and KEGG modules using *MinPath*, as provided with the HUMAnN2. Finally, the profiles of species and KEGG pathways were retained for further analysis if they showed non-zero abundance in at least 40% of samples.

DIAMOND was used in mapping CARD and Virulence Factor Database (VFDB) to identify resistance genes and virulence factors.

### **Draft genome of *M. pneumoniae***

Host-removed reads from all samples were co-assembled using *megahit*. Contigs of all samples were clustered to recover metagenome-assembled genomes (MAGs) using *MetaBAT2* under default parameters. The quality of MAGs was assessed using *CheckM* and *QUAST*. The taxonomic classification of MAGs was performed based on whole-genome alignment against the genome sequences published by IMG, NCBI, and PATRIC. Finally, we obtained the draft genome of *M. pneumoniae*, and gene prediction of *M. pneumoniae* was performed using *MetaGeneMark*, followed by identification of non-redundant genes using *CD-HIT*. The annotations of antibiotic resistance and virulence genes were acquired through CARD and VFDB using *BLASTP*.

### **Statistical analysis**

The statistical analyses of clinical records were performed using SPSS software (IBM, version 20.0). Data with a skewed distribution were presented as median values

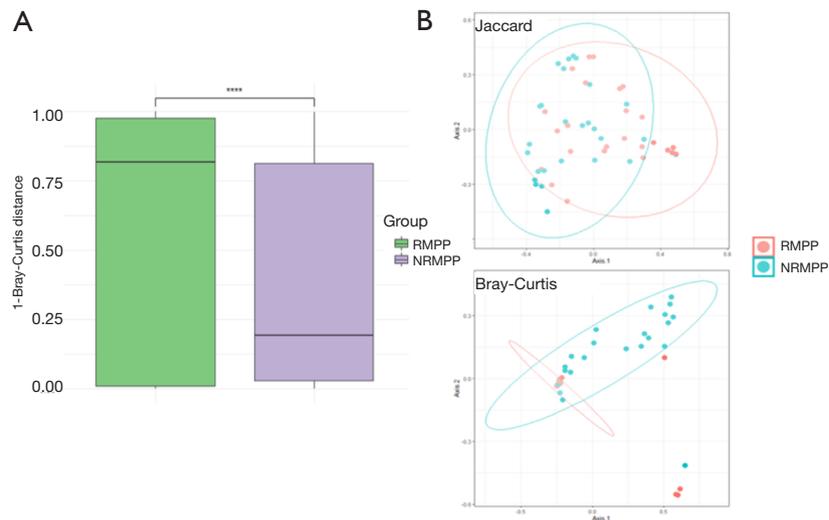
**Table 1** Demographical characteristics

Variable	NRMPP (n=27)	RMPP (n=25)	P value
Male	9 (33.3)	11 (44.0)	0.570
Age (months)	70 (51.0–84.0)	59 (45.0–71.5)	0.216
Hospitalization days (days)	7 (5.0–10.0)	9 (7.5–13.0)	0.008
Methylprednisolone treatment	25 (92.6)	24 (96.0)	1.000
Antibiotic treatments			
Macrolides			
Azithromycin	25 (92.6)	22 (88.0)	0.662
Clarithromycin	9 (33.3)	2 (8.0)	0.040
Cephalosporins			
Ceftriaxone	17 (63.0)	8 (32.0)	0.031
Cefoperazone/sulbactam	10 (37.0)	17 (68.0)	0.031
Fluoroquinolones			
Levofloxacin	4 (14.8)	6 (24.0)	0.492
Ciprofloxacin	1 (3.7)	4 (16.0)	0.183
Tetracyclines			
Minocycline	1 (3.7)	1 (4.0)	1.000
Laboratory data			
WBC counts ( $\times 10^9/L$ )	11.5 (9–14.2)	12.5 (10.35–16)	0.193
CRP (mg/L)	19.31 (10.32–57)	26 (17.5–41)	0.458
Fibrinogen (g/L)	3.52 (2.99–4.53)	3.86 (2.64–4.44)	0.847
LDH (U/L)	331 (284.0–347.0)	609 (512.5–811.5)	<0.001
D-dimer (mg/L)	0.85 (0.55–1.31)	1.48 (1.01–4.87)	0.003

Data are presented as median (interquartile ranges: Q1, Q3) or n (%).

(interquartile ranges: Q1, Q3). The different groups were compared using the Mann-Whitney U test. The statistical significance of the differences in the categorical variables was determined using the chi-square test. Statistical significance was determined at the two-tailed 0.05 level. Other statistical analyses were performed by R software (v4.0.1). Alpha diversity was estimated by the Shannon and Simpson index based on the taxonomic profile of each sample, beta diversity was assessed by the Bray-Curtis measure, and compared between NRMPP and RMPP patients by using Wilcoxon rank sum test, and was subsequently visualized by principal coordinate analysis (PCoA) plot. Permutational multivariate analysis of variance

(PERMANOVA) was performed by the R package “vegan” to analyze Bray-Curtis distance in different NRMPP and RMPP groups. Differential relative abundance of taxonomic groups at the genus level was tested by using Kruskal-Wallis rank sum test (R package “kruskal.test”). The genera with mean relative abundances greater than 0.4% and penetrance greater than 40% among all samples were compared. Spearman’s correlations between clinical characteristics and the relative abundances of genera were calculated by the R package “cor.test”, and FDR correction was adopted to adjust all P values. Statistically significant differences in the relative abundance of microbe and KEGG pathways among groups were assessed by the linear discriminant analysis of



**Figure 1** Comparison of microbial communities among the RMPP and NRMPP groups. (A) Beta diversity based on Bray-Curtis distance of two groups. \*\*\*\*,  $P < 0.001$ , PERMANOVA. (B) PCoA plot based on Bray-Curtis distance and Jaccard distance. Each plot represents one community from one patient. PERMANOVA, permutational multivariate analysis of variance; RMPP, refractory *M. pneumoniae* pneumonia; NRMPP, non-refractory *M. pneumoniae* pneumonia.

effect size (LEfSe) analysis.

## Results

### Demographical characteristics

In this study, the NRMPP group comprised 9 males and 18 females, and the RMPP group comprised 11 males and 14 females. The characteristics of these subjects are summarized in *Table 1*. No significant differences were observed between the baseline characteristics of the two groups ( $P > 0.05$ ). There was no difference in the treatment of methylprednisolone among groups ( $P > 0.05$ ). As for antibiotics treatment, no significant ( $P > 0.05$ ) difference in treatment of azithromycin, levofloxacin, ciprofloxacin, and minocycline was observed whereas a significant difference was observed in the treatment of ceftriaxone, cefoperazone/sulbactam and clarithromycin ( $P < 0.05$ ). Regarding laboratory data and hospitalizations days, higher LDH, D-dimer levels and prolonged hospitalization days were observed in the RMPP group than NRMPP group ( $P < 0.05$ ), whereas markers of systemic inflammation such as WBC counts, CRP and fibrinogen levels had no difference

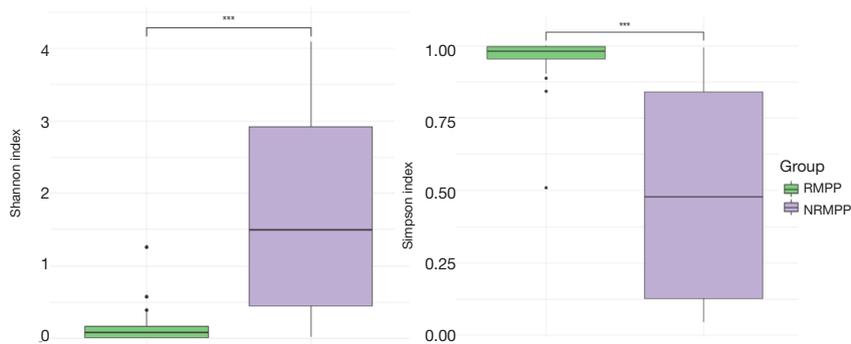
between groups ( $P > 0.05$ ).

### Comparison of microbial communities among the RMPP and NRMPP groups

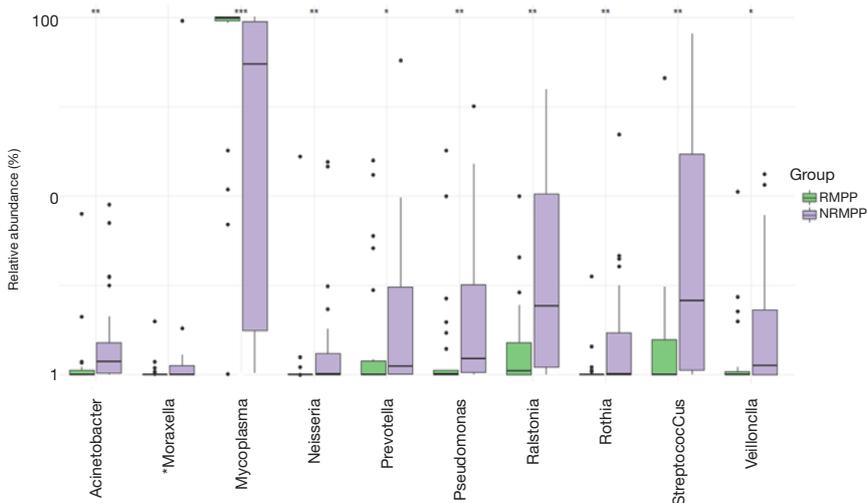
Principal coordinates analysis (PCoA) was used to compare microbial communities among the two groups. In addition, regarding beta diversity based on Bray-Curtis distance, there was a significant difference between the two groups ( $P < 0.001$ , PERMANOVA), which means that the intra-group difference of RMPP group was smaller than that of NRMPP group (*Figure 1A*). PCoA plot based on neither Bray-Curtis distance nor Jaccard distance could effectively distinguish RMPP and NRMPP group (*Figure 1B*).

### Alpha diversity analysis between the RMPP and NRMPP groups

Then alpha diversity was compared between the RMPP and NRMPP groups. Two of the alpha diversity calculated indexes, namely the Shannon and Simpson indices, representing the richness and evenness of the microbiota, were measured. A significant decrease in microbial diversity



**Figure 2** Comparison of two of the alpha diversity calculated indexes, Shannon, and Simpson indices between RMPP and NRMPP groups. \*\*\*, P<0.001, Wilcoxon rank sum test. RMPP, refractory *M. pneumoniae* pneumonia; NRMPP, non-refractory *M. pneumoniae* pneumonia.



**Figure 3** The most abundant genera (mean relative abundances greater than 0.4% and penetrance greater than 40% among all samples) among two groups. \*, P<0.05; \*\*, P<0.01; \*\*\*, P<0.001, Kruskal-Wallis rank sum test.

and increased dominance was observed in the samples collected from the RMPP group (Figure 2).

**The most abundant taxa between the RMPP and NRMPP groups**

We compared the most abundant phyla and genera between two groups (Figure 3; Table 2). At the phylum level, the top five phyla of the RMPP BALF microbiota were Tenericutes, Proteobacteria, Firmicutes, Bacteroidetes and Actinobacteria. As for the NRMPP group, the order of the top phyla was comparable to that of the RMPP group. Interestingly, the relative abundance of phylum Tenericutes significantly decreased in the NRMPP group whereas

the Proteobacteria, Firmicutes and Actinobacteria were more abundant than RMPP group. At the genus level, the RMPP BALF microbiota was dominated by *Mycoplasma*, *Streptococcus*, *Prevotella*, *Pseudomonas* and *Ralstonia*. In the NRMPP group, the relative abundance of genus *Mycoplasma* was dramatically lower than the RMPP group. The relative abundance of commensal genera such as *Streptococcus*, *Neisseria*, *Veillonella*, *Rothia* and *Prevotella* as well as that of the pathogenic genera, *Acinetobacter*, *Pseudomonas* and *Ralstonia*, was significantly higher in the NRMPP group (8). Additionally, we compared the *M. pneumoniae* load between the two groups. As shown in Table 2, *M. pneumoniae* load was significantly higher in almost all the RMPP patients than in the NRMPP group, indicating that *M. pneumoniae* occupied

**Table 2** The top five phyla and genera of two groups

Taxonomy (% relative abundance)	NRMPP (n=27)	RMPP (n=25)	P value
Phylum			
Tenericutes	54.23 (0.77–94.87)	98.52 (95.41–99.89)	<0.001
Proteobacteria	6.03 (1.26–31.97)	0.43 (0.05–3.01)	<0.001
Firmicutes	2.38 (0.34–26.67)	0.06 (0.01–0.96)	<0.001
Bacteroidetes	0.30 (0.12–2.64)	0.02 (0.00–0.22)	0.029
Actinobacteria	0.98 (0.38–4.20)	0.05 (0.00–0.23)	<0.001
Genus			
<i>Mycoplasma</i>	54.21 (0.77–94.87)	98.52 (95.41–99.89)	<0.001
<i>Streptococcus</i>	1.59 (0.06–1.67)	0.00 (0.00–0.58)	0.001
<i>Ralstonia</i>	1.42 (0.10–9.37)	0.05 (0.00–0.51)	0.004
<i>Prevotella</i>	0.12 (0.01–2.22)	0.00 (0.00–0.20)	0.041
<i>Pseudomonas</i>	0.24 (0.03–2.27)	0.00 (0.00–0.06)	0.009
<i>M. pneumoniae</i> load <sup>†</sup> (n)	184.9 (14.39–1,350.33)	1,609.34 (702.76–10,879.62)	0.007

Data are presented as median (interquartile ranges: Q1, Q3). <sup>†</sup>, *M. pneumoniae* load is presented as number of *M. pneumoniae* reads per million reads

almost all of the niches in the LRT. In addition, we noticed that four patients from RMPP group had significantly lower *M. pneumoniae* relative abundance, and high load of human adenovirus 7 or human bocavirus type I was detected from these samples.

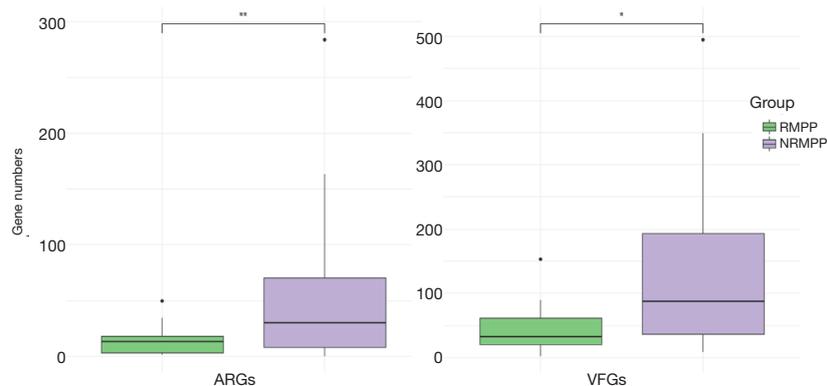
### Biomarkers of the RMPP and NRMPP groups

LEfSe was used to explore the taxa with the strongest effect on different microbiology conditions among groups (Figure S1). This analysis detected two species from two genera as biomarkers for the RMPP group, and eleven species from seven genera for the NRMPP group. The *Mycoplasma* and *Neisseria* genera had the most powerful influence on RMPP, with *Mycoplasma pneumoniae* and *Neisseria flavescens* as the most abundant species. The most dominant NRMPP-associated biomarker were class Bacilli, genera *Streptococcus*, *Ralstonia* and species *Ralstonia pickettii* and *Streptococcus salivarius*.

### Differences in the distribution of antibiotic resistance genes (ARGs) and virulence factor genes (VFGs) between the RMPP and NRMPP groups

Reads from all samples were used to re-assemble the draft

genome of *M. pneumoniae*, since data from a single group is not enough to obtain the complete genome (Figure S2). In total, 182,740 VFGs were uncovered in the *M. pneumoniae* genome including virulence-determined VFGs such as cytoadherence accessory protein, community-acquired respiratory distress syndrome (CARDS) toxin, ADP1\_MYCPN adhesin P1 and other virulence factors. The *M. pneumoniae* genome was found to consist of abundant ARGs against common antibiotics including fluoroquinolone, tetracycline, nitroimidazole, peptide, and macrolide. Next, we compared the annotated ARGs and VFGs in *M. pneumoniae* genome between NRMPP and RMPP groups (Figure 4). Interestingly, ARGs were successfully annotated in *M. pneumoniae* genome from 14 of 25 (56.0%) patients in RMPP group and 18 of 27 (66.7%) in NRMPP group. In addition, the NRMPP group held a greater number of annotated ARGs than the RMPP group as well as a higher relative abundance of macrolide ARGs (macB), fluoroquinolone resistance gene (patAB), nitroimidazole ARG (msbA) and peptide ARG (bcrA) (Figure 5A). It was suggested that the macrolide antibiotic resistance of *M. pneumoniae* in NRMPP may be higher than in RMPP. Moreover, the relative quantity of resistance genes of tetracycline antibiotics (tetT) in the RMPP group was higher than those of the NRMPP group. A 23rRNA



**Figure 4** Annotated numbers of ARGs and VFGs in *M. pneumoniae* genome between the RMPP and NRMPP groups. \*,  $P < 0.05$ ; \*\*,  $P < 0.01$ , Wilcoxon rank sum test. ARGs, antibiotic resistance genes; VFGs, virulence factor genes; RMPP, refractory *M. pneumoniae* pneumonia; NRMPP, non-refractory *M. pneumoniae* pneumonia.

gene single-nucleotide polymorphism mutation related to macrolide antibiotics resistance, A2063G, was detected from a patient from the RMPP group who received prolonged macrolide, cephalosporin, and fluoroquinolone antibiotics treatment (azithromycin, ceftazidime-sulbactam and levofloxacin). Finally, The RMPP group had less annotated VFGs than NRMPP group while the relative abundance of adhesion-related virulence genes was higher (Figure 5B).

#### Microbial function differences between the RMPP and NRMPP groups

By means of LEfSe, we assessed the biomarker pathways of the BALF microbiota in children with RMPP (Figure S3). Microbial functions relating to metabolism of terpenoids and polyketides, lipid metabolism, DNA replication or repair and carbohydrate metabolism were found to be RMPP-associated pathways.

#### Association between BALF microbiota and hospitalization days

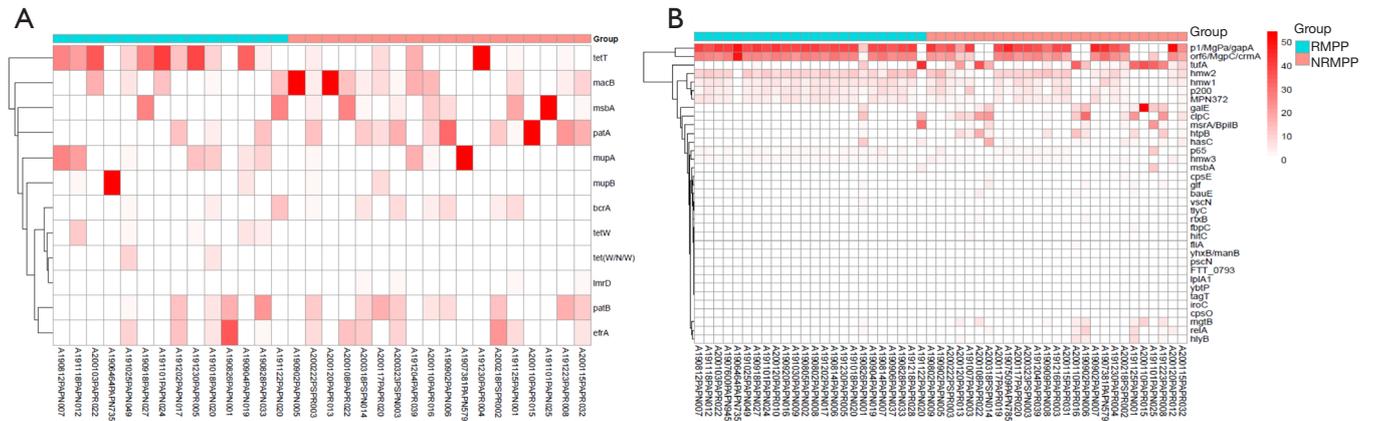
We used spearman correlation analysis to observe relationship between BALF microbiota and hospitalization days. In RMPP patients, higher relative abundance of *Streptococcus oralis* and *Streptococcus pneumoniae* as well as *Streptococcus* genera had a strong correlation with increased hospitalization days (Figure 6A). Interestingly, higher relative abundance of *Streptococcus pneumoniae* had a negative correlation with hospitalization days among NRMPP patients (Figure 6B). In addition, higher relative abundance

of *M. pneumoniae* and lower Shannon index were also associated with increased hospitalization days in NRMPP group (Figure 6C,D).

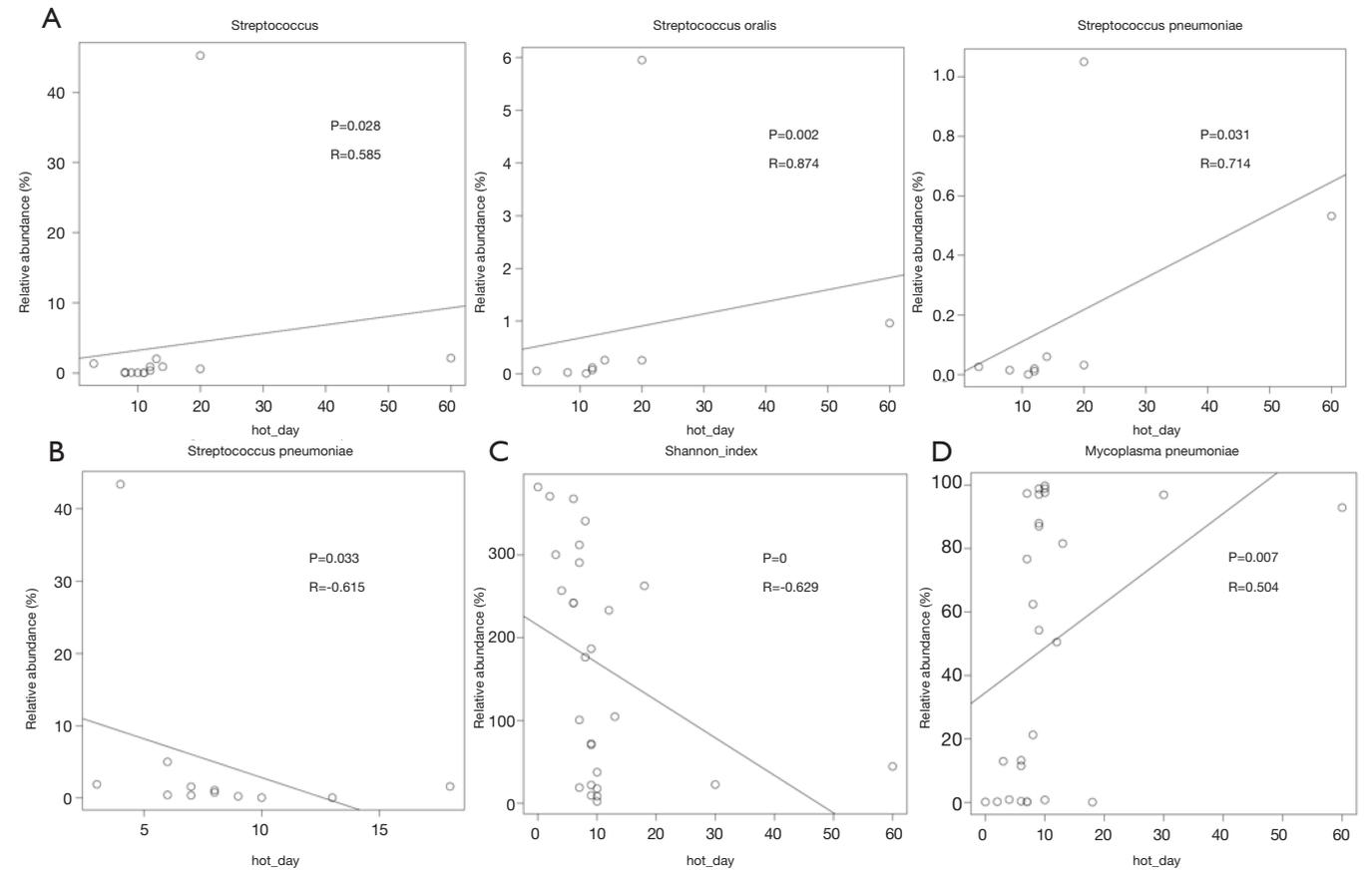
## Discussion

This is a pioneering study of LRT microbiome in children with RMPP. Although there have been many studies on the LRT microbiota of patients with pneumonia, the microbial function and host-microbial interaction remain to be explored, especially in children. It is clearly helpful to monitor changes in *M. pneumoniae* resistance and virulence, guide the use of antibiotics and, most importantly, advance our understanding of pathogenesis of RMPP.

The microbiota of LRT in children with RMPP is much simpler than that in children with NRMPP, and most of them are dominated by *M. pneumoniae*. Some researchers found that *M. pneumoniae* could eliminate other bacteria by directly competing for nutrients (9). Moreover, it clears other bacteria by activating the host inflammatory response (10). Together, these mechanisms could lead to decreased colonization of the commensal genera, *Prevotella*, *Streptococcus*, *Veillonella* and *Rotbia*, in the respiratory tract of children with RMPP (8,11,12). In addition, the proliferation of pathogenic genera such as *Pseudomonas* and *Acinetobacter* were also inhibited. The microbial structure of RMPP group is highly homogeneous according to its lower intra-group beta diversity. Thus, such high similarity of microbial communities may be the signature of RMPP. In addition, the relative abundance of adherence-associated VFG was higher in RMPP group, as well as the more active KEGG



**Figure 5** Relative abundance of ARGs (A) and VFGs (B) in *M. pneumoniae* genome among RMPP and NRMPP groups. ARGs were successfully annotated in *M. pneumoniae* genome from 14 of 25 patients in RMPP group and 18 of 27 patients in NRMPP group. Each small square was based on the mapped numbers of ARGs or VFGs in *M. pneumoniae* genome and was normalized to represents the percentage of a specific antibiotic resistance gene or virulence factor gene to all ARGs or VFGs. ARGs, antibiotic resistance genes; VFGs, virulence factor genes; RMPP, refractory *M. pneumoniae* pneumonia; NRMPP, non-refractory *M. pneumoniae* pneumonia.



**Figure 6** Association between BALF microbiota and hospitalization days in RMPP patients (A) and NRMPP patients (B,C,D). †, hospitalization days. BALF: bronchoalveolar lavage fluid. RMPP, refractory *M. pneumoniae* pneumonia; NRMPP, non-refractory *M. pneumoniae* pneumonia.

pathway of glycolipid metabolism, which may be a key player in the virulence of *M. pneumoniae* (13). It is possible that a large number of *M. pneumoniae* can cause excessive inflammatory response and immune damage. *M. pneumoniae* secretes adhesion molecules with P1 adhesion protein to adhere to the epithelial cells, gradually reduces the function of phagocytes through fusion with host target cells, and causes immune damage (14). Simultaneously, *M. pneumoniae* infection stimulates the activation of T cells, and promotes higher expressions of cytokines and inflammatory factors, aggravating the inflammatory response (15-17). Such conditions may be due to the fact that the extensive use of broad-spectrum antibiotics such as cefoperazone in the RMPP group reduces the diversity of LRT, providing a less-competitive environment for *M. pneumoniae* to spread. Once *M. pneumoniae* occupies the whole niche, a large number of adhered airway epithelial cells become necrotic and exfoliate, inflammatory cells infiltrate and proinflammatory cytokines are secreted, and the release of CARDS toxin causes serious tissue damage to patients and such excessive activation of the immune system may be the main cause of RMPP (18).

Our results revealed that the NRMPP group were enriched for more ARGs than the RMPP group, and also showed a higher relative abundance of the macrolide ARG (macB), fluoroquinolone resistance gene (patA-B) and other ARGs (bcrA, msbA). Thus, the higher relative abundance of the macrolide ARG may not relate to RMPP. It was also reported that antibiotic-resistant *M. pneumoniae* infection often causes only mild pneumonia, while antibiotic-sensitive *M. pneumoniae* infection often causes RMPP, and *M. pneumoniae* resistance was not associated with serious treatment failure or the occurrence of serious complications, suggesting that the infection of resistant *M. pneumoniae* may not be the main reason for RMPP (19). On the other hand, integrative and conjugative elements (ICEs) even the large chromosomal DNA fragments could be horizontally transferred among *M. pneumoniae* via conjugative transfers under the rigorous antibiotic selection pressure and complicated microbe-microbe interaction condition (20). Macrolides antibiotics, tetracycline and fluoroquinolones antibiotics are first-line and second-line drugs for the treatment of MPP, respectively. Such transfer of genetic components or ARGs (such as tetT, patA-B) may lead to widespread transfer of drug resistance genes among *M. pneumoniae* and the failure of treatment against MPP. In addition, many studies have suggested that point mutations in the 23S rRNA gene and the ribosomal proteins L4 and

L22 are predominantly responsible for macrolide resistance in *M. pneumoniae* (21,22). Most of the mutations that found occurred at sites 2063 and 2064 in the 50 subunit 23S ribosomal RNA region V while some occurred at sites 2617 (C2617G, C2617A) (23,24). This gene mutation has been shown to induce macrolide resistance (PMCID: PMC529214) and also associated to restriction fragment length polymorphism in the P1 cytoadhesion gene (PMCID: PMC228818). In our study, merely one patient who received prolonged and complex antibiotics treatment carried the A2063G mutation. Interestingly, macB, which was first identified as a macrolide antibiotic-specific transporter in *E. coli*, was widely identified in NRMPP patients (25). As a member of family of ATP-binding cassette-type transporters, it utilizes ATP hydrolysis as a driving force to pump out macrolide antibiotics. What should be recognized is that in China the macrolide-specific efflux system and modification of target sites are equally crucial mechanisms responsible for macrolide resistance in *M. pneumoniae*.

We uncovered that *Streptococcus oralis* and *Streptococcus pneumoniae* were associated with an increase in hospitalization days among RMPP patients. It is suggested that such species may be not conducive to the prognosis of RMPP. Actually, *Streptococcus oralis* was thought to be associated with ventilator-associated pneumonia and accelerated replication of influenza viruses in LRT while *Streptococcus pneumoniae* was a common pathogen of community-acquired pneumonia (26,27). Conversely, *Streptococcus pneumoniae* was negatively associated with prolonged hospitalization days in NRMPP group, while prolonged hospitalization days was associated with lower Shannon diversity and higher *M. pneumoniae* abundance. Thus, we proposed a hypothesis that the relative abundance and dominance of *M. pneumoniae* in LRT are factors determining the hospitalization days in the case of NRMPP due to its low density. For RMPP, the *M. pneumoniae* was close to covering the entire LRT, and the relative abundance of other pathogens became the determinant of the hospitalization days.

Although *M. pneumoniae* played an important role in RMPP pathogenesis, *Neisseria flavescens* was detected as a RMPP-associated biomarker via LEfSe. *Neisseria flavescens* is generally a commensal species but it turned out to be an important role in RMPP. In fact, *Neisseria flavescens* was previously reported to act as a pathogen in the LRT, and hence, it is possible that the pathogenic potential of such commensal species is activated in the inflammatory

environment (28-30).

There are several limitations of this study. All children were treated with at least one antibiotic before bronchoscopy, and the treatments of ceftriaxone and cefoperazone/sulbactam were not balanced between groups, which may lead to temporary changes in the microecology of their LRT, as antibiotics themselves can reduce microbiota diversity. In addition, the number of patients in this study was not particularly large. An important limitation of mNGS is that it is only at the DNA level, and thus, it cannot distinguish between live and dead microorganisms and cannot determine the expression of related functional genes. Combined with metatranscriptomic sequencing, however, it can provide a real-life profile and function of microorganisms.

## Conclusions

In summary, we explored the changes of microbiota structure and related functions in the LRT of patients with RMPP and NRMPP, which should advance our understanding of the pathogenesis of RMPP and provide foundations for further research.

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*Reporting Checklist:* The authors have completed the MDAR checklist. Available at <http://dx.doi.org/10.21037/tp-20-404>

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*Conflicts of Interest:* All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/tp-20-404>). The authors have no conflicts of interest to declare.

*Ethical Statement:* The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The ethical

application and the consent procedure of this study were approved by the Ethics Committee of Children’s Hospital of Fudan University (No. 2016-87). All procedures performed in this study involving human participants were in accordance with the Declaration of Helsinki (as revised in 2013), and informed consent was taken from all the parents.

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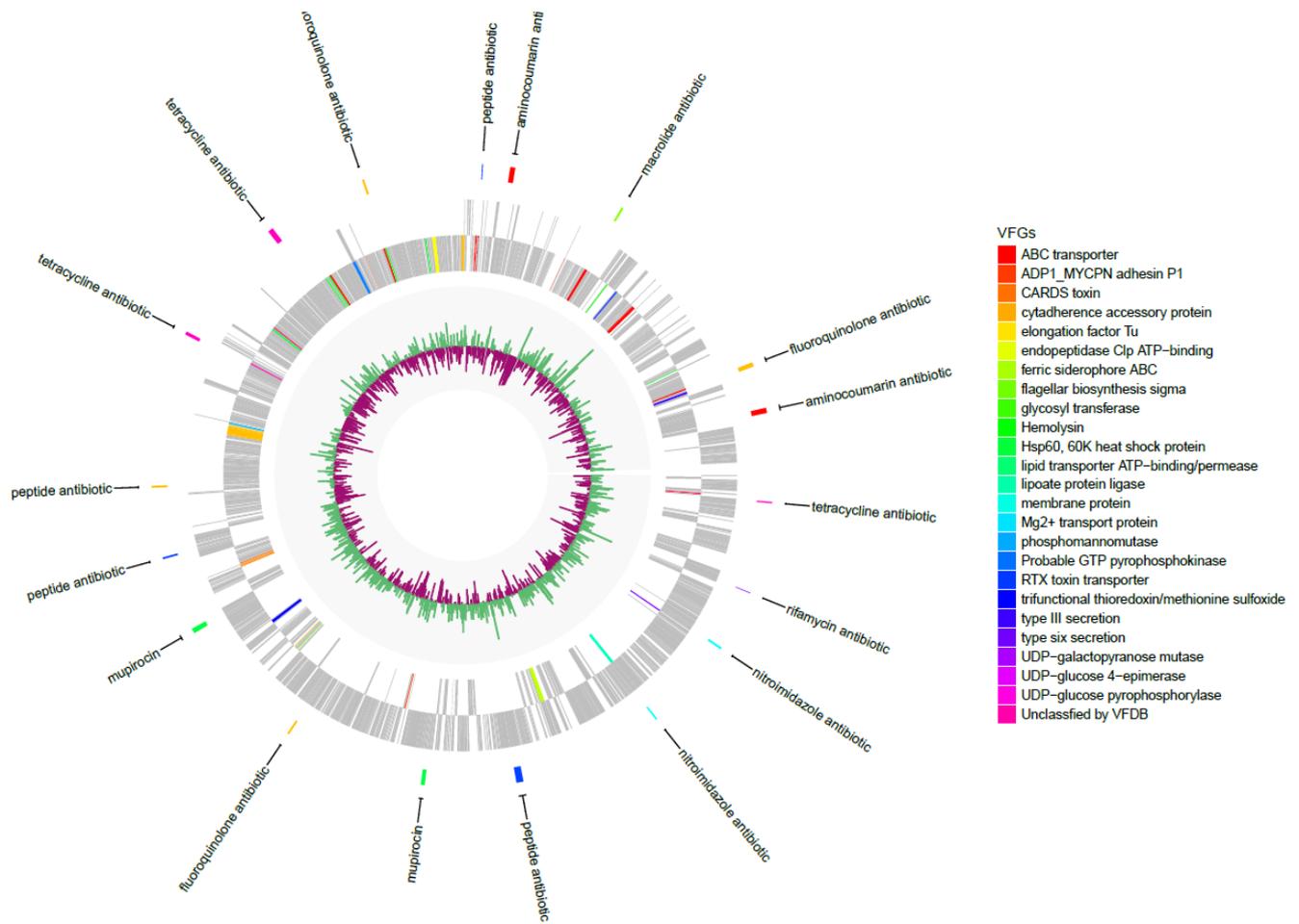
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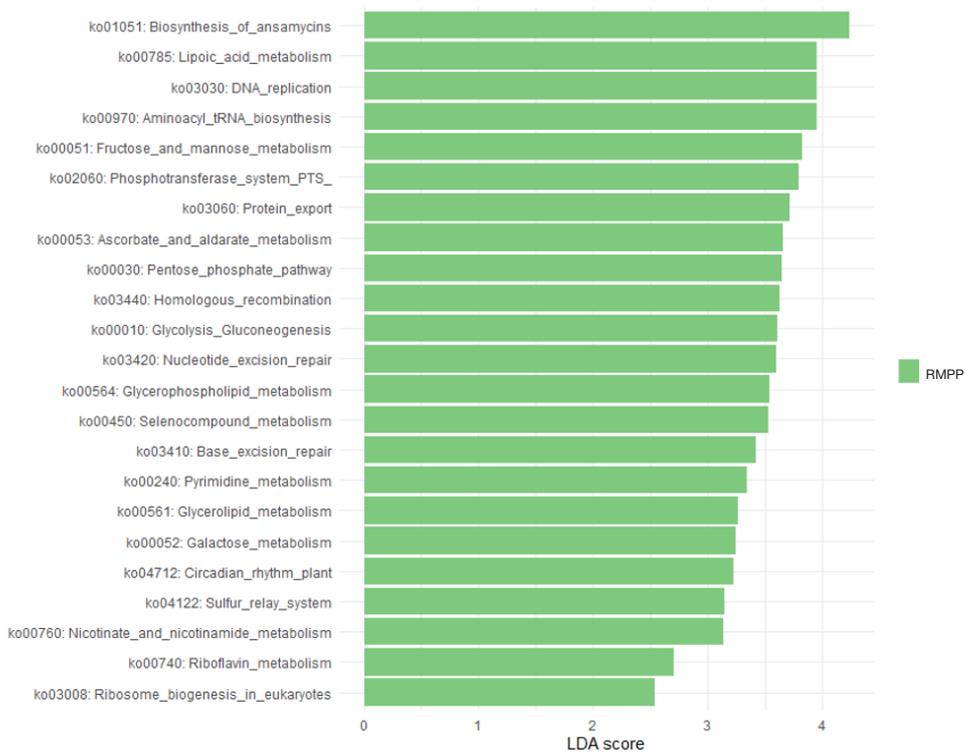
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**Figure S2** ARGs and VFGs shown on the draft genome of *M. pneumoniae*. Tracks from outside to inside represent ARGs, genes on the plus strand, genes on the negative strand, and guanine-cytosine skew, respectively. VFG colors indicate VFG type. ARGs, antibiotic resistance genes; VFGs, virulence factor genes.



**Figure S3** LEfSe analysis of KEGG pathway among RMPP group. The bar graph shows KEGG pathways at level 3 that the LDA score over 2.0. LEfSe, Linear discriminant analysis Effect Size; RMPP, refractory *M. pneumoniae* pneumonia.