



# Correlation study of cytokine levels in alveolar lavage fluid with exhaled nitric oxide and lung function in children with bronchial asthma

Ying Lou, Qiuping Ke, Huailiang Cui, Ying Shang, Chengsheng Yang

The Second Ward of Pediatrics, Xinxiang Central Hospital, The Fourth Clinical College of Xinxiang Medical University, Xinxiang, China

**Contributions:** (I) Conception and design: Y Lou; (II) Administrative support: Q Ke; (III) Provision of study materials or patients: Y Lou, C Yang; (IV) Collection and assembly of data: All authors; (V) Data analysis and interpretation: Y Lou, C Yang; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

**Correspondence to:** Ying Lou. The Second Ward of Pediatrics, Xinxiang Central Hospital, The Fourth Clinical College of Xinxiang Medical University, Xinxiang 453000, China. Email: louying59711@126.com.

**Background:** The associations between cytokines in the bronchoalveolar lavage fluid (BALF), lung cytokine expression, fractional exhaled nitric oxide (FeNO) and pulmonary function test results in pediatric asthmatics have not been extensively characterized. This study sought to explore correlations between cytokines BALF, FeNO, and pulmonary function test results.

**Methods:** From October 2018 to October 2020, a prospective study was conducted on 42 children with asthma and 17 children with pulmonary foreign bodies that required bronchoscopy. Pulmonary function tests and FeNO tests were performed on all patients. Patients were divided into a high FeNO group or low FeNO group based on their FeNO results. Interleukin (IL)-4, IL-5, IL-6, IL-8, IL-13, and IL-17 in the BALF were measured by enzyme-linked immunosorbent assays. Pearson correlations were used to assess the correlations between the cytokines in BALF, the pulmonary function test results, and the FeNO results. Pearson correlation was used to calculate the correlation coefficient “r” among alveolar lavage fluid cytokines, lung function, and FeNO. Receiver operating characteristic (ROC) curves were used to determine the area under the curve (AUC), sensitivity, and specificity of BALF cytokines for the high and low FeNO groups.

**Results:** IL-4, IL-5, IL-6, IL-8, IL-13, and IL-17 in BALF were significantly correlated with FeNO, but were not significantly correlated with the pulmonary function test results. Cytokine IL-4, IL-5, IL-6, IL-8, IL-13, and IL-17 in BALF were significantly different in the high FeNO, low FeNO, and control groups (all  $P < 0.05$ ). The AUCs for differentiating between low and high FeNO based on BALF cytokines ranged from 0.72 to 0.95. The sensitivity and specificity for discriminating between low and high FeNO based on IL-5 and IL-13 reached 95.7% and 100%, respectively.

**Conclusions:** The cytokine levels of the BALF of children with asthma were significantly elevated, correlated with FeNO, and can be used evaluate airway inflammation in children with asthma.

**Keywords:** Bronchial asthma; cytokine; nitric oxide; pulmonary function

Submitted Jun 17, 2021. Accepted for publication Aug 10, 2021.

doi: 10.21037/tp-21-322

**View this article at:** <https://dx.doi.org/10.21037/tp-21-322>

## Introduction

Bronchial asthma is a common chronic respiratory disease in children, characterized by chronic airway inflammation, airway hyperresponsiveness, and reversible

airflow obstruction. Studies have shown that effector cells secrete various cytokines that mediate and maintain tissue inflammation and injury, thus promoting the proliferation of goblet cells, smooth muscle cells, and the extracellular

matrix (1). Animal and clinical studies have shown significant differences in cytokine abundance in serum and bronchoalveolar lavage fluid (BALF) between bronchial asthma patients and healthy control patients. Significant differences have also been found in the cytokine expression of patients with different severities of bronchial asthma, and those in the remission stage or acute exacerbation stage (2,3). At present, pulmonary function examination is widely carried out in clinical settings. Studies have shown that the level of fractional exhaled nitric oxide (FeNO) better reflects the degree of airway inflammation than cytokine expression (4). This study sought to evaluate correlations between the levels of cytokines in BALF, lung function indexes, and FeNO in children with bronchial asthma, and explore the relationship between the degree of the local inflammatory response and lung function in bronchial asthma to provide novel insights into the pathogenesis of bronchial asthma. We present the following article in accordance with the STARD reporting checklist (available at <https://dx.doi.org/10.21037/tp-21-322>).

## Methods

### Research subjects

From October 2018 to October 2020, 42 children with bronchial asthma who underwent bronchoscopy at our department were selected as the research subjects of this study. To be eligible to participate in this study, patients had to meet the following inclusion criteria: (I) have been diagnosed with bronchial asthma according to the diagnostic criteria in the *Guidelines for the Diagnosis and Prevention of Bronchial Asthma in Children* (2016); (II) be aged 6–12 years and able to cooperate with the bronchoscopy, pulmonary function, and FeNO examinations; (III) have no history of infection and antibiotics or glucocorticoid use in the past month; (IV) have no other serious complications, such as severe hepatic and renal insufficiency; and (V) have a guardian who was willing to provide informed consent. Patients were excluded from the study if they were suffering from wheezing due to common causes, such as tracheal stenosis or softening. For the control group, 17 children were selected who attended the hospital for the treatment of a bronchial foreign body and underwent a bronchoscopy at our department during the same period. All the procedures in this study involving human participants were performed in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by the Ethics Committee of

Xinxiang Central Hospital, The Fourth Clinical College of Xinxiang Medical University (No.:2017-006), and informed consent was obtained from all the patients' guardians.

### Determination of cytokines in BALF

Bronchoalveolar lavage was performed according to the guidelines for pediatric bronchoscopy. During the operation, the bronchoscope was placed at the opening of the right middle bronchus, 0.5 mL/kg of normal saline was injected, and negative pressure suction was then used to collect the lavage fluid. The lavage solution was centrifuged at 1,000 rpm for 20 minutes, and the supernatant was collected in a test tube. The specimens were stored in a  $-20^{\circ}\text{C}$  refrigerator to avoid repeated freezing and thawing.

The supernatant samples were dissolved at room temperature, and interleukin (IL)-4, IL-5, IL-6, IL-7, IL-9, IL-13, and IL-17 were detected using an enzyme-linked immunosorbent assay kit produced by Jiangsu Biyuntian Company. Each specific step was executed in accordance with the manual.

### FeNO determination

FeNO was determined by a Niox Mino eNO analyzer produced by Aerocrine, Sweden. Before examination, the patients were instructed to avoid strenuous exercise, cold water, superheated water, carbonated drinks, and nitrogenous food (such as soybean milk and coffee) and were forbidden to eat and drink 1 hour before the examination. During the test, the expiratory flow rate was set to about 50 mL/s, and the expiratory time was set to 10 s. The test was repeated 3 times to ensure that the deviation of the 3 results was not more than 10%; the average of the 3 results was taken as the final result. Children with bronchial asthma were divided into a high FeNO group [FeNO  $\geq 25$  ppb (parts per billion)] or low FeNO group (FeNO  $< 25$  ppb) according to their FeNO value.

### Lung function test

Pulmonary function was measured by a pulmonary function instrument produced by Kanger Fusheng Co. Ltd, United States. 1 hour after FeNO determination, lung function was detected. Before the measurement, the environmental factors and flow rate were calibrated. Children were asked to follow the technician's directions to first breathe calmly,

**Table 1** Comparison of general data and lung function indexes between control group and children with bronchial asthma

Variables	Control group (n=17)	Bronchial asthma group (n=42)		Statistics	P value
		Low FeNO (n=19)	High FeNO (n=23)		
Age (years)	8.1±2.0	7.9±1.6	7.7±1.3	0.3	0.7423
Sex (M/F)	10/7	11/8	9/14	2.07	0.35
Height (cm)	134.0±11.4	129.4±11.5	137.8±14.1	2.327	0.1069
Weight (kg)	41.5±10.9	34.4±10.9	38.4±6.6	2.567	0.0858
FeNO (ppb)	11.41±3.81	14.58±2.78 <sup>a</sup>	43.17±3.89 <sup>a,b</sup>	508.7	0.0000
FEV1/FVC%	91.61±2.82	79.75±3.49 <sup>a</sup>	77.07±2.97 <sup>a,b</sup>	115.2	0.0000
MEF25 (L/s)	1.49±0.15	0.74±0.14 <sup>a</sup>	0.71±0.17 <sup>a,b</sup>	147.8	0.0000
MEF50 (L/s)	3.06±0.54	1.86±0.53 <sup>a</sup>	1.82±0.19 <sup>a,b</sup>	48.15	0.0000
MEF75 (L/s)	4.67±0.49	3.46±0.58 <sup>a</sup>	3.42±1.23 <sup>a,b</sup>	11.92	0.0000

<sup>a</sup>, compared with the control group, P<0.05; <sup>b</sup>, compared with the low FeNO group, P<0.05. FeNO, fractional exhaled nitric oxide.

to try to inhale fully, and to then exhale the gas faster, being careful to avoid mouth leakage. We selected forced expiratory volume in 1 second/forced vital capacity (FEV1/FVC%), and maximal expiratory flow at (MEF)25, MEF50, and MEF75 to analyze.

### Statistical analysis

Measurement data with normal distributions are expressed as means ± standard deviations. Count data are expressed as percentages. A univariate analysis of variance or chi-square test was used to compare differences between the general data, lung function indexes, and cytokine levels in BALF among the low FeNO, high FeNO, and control groups. Pearson correlation was used to calculate the correlation coefficient “r” between FeNO and cytokines in BALF and lung function in children with bronchial asthma. The receiver operating characteristic (ROC) curve was used to determine the cutoff, area under the curve (AUC), sensitivity, and specificity of low FeNO and high FeNO. A P<0.05 was considered statistically significant.

## Results

### General data and pulmonary function index

As *Table 1* shows, there was no statistical difference in age, gender, body weight, and other general data between the control, low FeNO, and high FeNO groups. FEV1/FVC%, MEF25, MEF50, and MEF75 in the low FeNO and high

FeNO groups were lower than those in the control group. The difference was statistically significant (all P<0.05). Conversely, there was no significant difference in the lung function indexes between the low and high FeNO groups (all P>0.05).

### Comparison of cytokines in BALF

The levels of IL-4, IL-5, IL-6, IL-8, IL-13, and IL-17 in the BALF of the low FeNO group and the high FeNO group were significantly higher than those of the control group (all P<0.05). In addition, the levels of IL-4, IL-5, IL-6, IL-8, IL-13, and IL-17 in the BALF of the high FeNO group were significantly higher than those of the low FeNO group (all P<0.05; see *Table 2*).

### The correlation between BALF, FeNO, and lung function in children with bronchial asthma

There was no significant correlation between FeNO and FEV1/FVC%, MEF25, MEF50, and MEF75 (all P>0.05). FeNO was significantly correlated with IL-4, IL-5, IL-6, IL-8, IL-13, and IL-17 in the BALF (r 0.306–0.745). There was no significant correlation between IL-4, IL-5, IL-6, IL-8, IL-13, IL-17, FEV1/FVC%, MEF25, MEF50, and MEF75 (all P>0.05; see *Table 3*).

### ROC curves of low FeNO and high FeNO in BALF

As *Table 4* and *Figure 1* show, low FeNO and High FeNO

**Table 2** Comparison of BALF between the control group and children with bronchial asthma

Cytokines	Control (n=17)	Low FeNO (n=19)	High FeNO (n=23)	Statistics	P
IL-4 (ng/L)	4.31±1.42	17.49±2.69 <sup>a</sup>	21.78±3.73 <sup>a,b</sup>	185.8	0.0000
IL-5 (ng/L)	6.39±2.10	24.25±7.49 <sup>a</sup>	32.76±5.07 <sup>a,b</sup>	117.2	0.0000
IL-6 (ng/L)	10.26±0.76	28.50±4.34 <sup>a</sup>	34.35±8.21 <sup>a,b</sup>	90.53	0.0000
IL-8 (ng/L)	4.04±0.67	6.96±0.97 <sup>a</sup>	8.86±0.74 <sup>a,b</sup>	176.0	0.0000
IL-13 (ng/L)	3.37±0.37	6.30±0.71 <sup>a</sup>	9.41±1.53 <sup>a,b</sup>	160.8	0.0000
IL-17 (ng/L)	42.84±6.25	70.50±11.45 <sup>a</sup>	82.59±12.70 <sup>a,b</sup>	67.47	0.0000

<sup>a</sup>, compared with the control group, P<0.05; <sup>b</sup>, compared with the low FeNO group, P<0.05. BALF, bronchoalveolar lavage fluid; FeNO, fractional exhaled nitric oxide.

**Table 3** Correlation among cytokines, FeNO, and lung function in the BALF of children with bronchial asthma (n=42)

Variables	FeNO	IL-4	IL-5	IL-6	IL-8	IL-13	IL-17
FeNO	–	0.473**	0.306*	0.337*	0.701***	0.745***	0.522***
FEV1/FVC%	–0.252	–0.172	–0.276	–0.230	–0.102	–0.098	–0.038
MEF25	–0.006	0.111	0.051	0.274	–0.016	0.036	0.240
MEF50	0.094	–0.139	0.235	–0.194	0.046	0.007	–0.110
MEF75	–0.058	0.146	–0.207	0.072	0.011	–0.267	0.029

\*, P<0.05; \*\*, P<0.01; \*\*\*, P<0.001. BALF, bronchoalveolar lavage fluid; FeNO, fractional exhaled nitric oxide.

**Table 4** The ability of cytokines in the BALF of children with bronchial asthma to distinguish between low FeNO and high FeNO

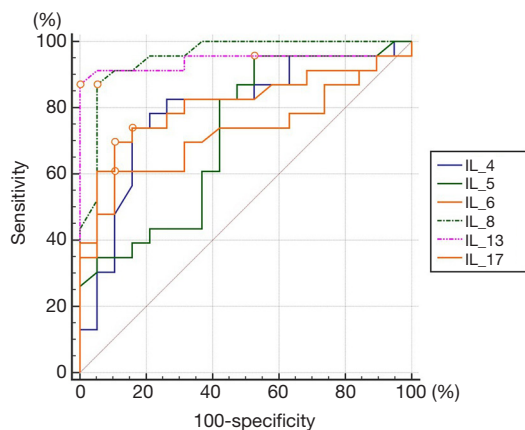
Cytokines	Cutoff	AUC (95% CI)	Sensitivity (%) (95% CI)	Specificity (%) (95% CI)
IL-4 (ng/L)	20.2	0.79 (0.64–0.90)	73.9 (51.6–89.8)	84.2 (60.4–96.6)
IL-5 (ng/L)	19.7	0.72 (0.56–0.85)	95.7 (78.1–99.9)	47.4 (24.4–71.1)
IL-6 (ng/L)	32.7	0.72 (0.57–0.85)	60.9 (38.5–80.3)	89.5 (66.9–98.7)
IL-8 (ng/L)	8.2	0.95 (0.84–0.99)	87.0 (66.4–97.2)	94.7 (74.0–99.9)
IL-13 (ng/L)	7.7	0.94 (0.82–0.99)	87.0 (66.4–97.2)	100 (82.4–100)
IL-17 (ng/L)	81.7	0.81 (0.66–0.92)	69.6 (47.1–86.8)	89.5 (66.9–98.7)

BALF, bronchoalveolar lavage fluid; FeNO, fractional exhaled nitric oxide; Cutoff, the best cutoff value; AUC, area under the curve; CI, confidence interval.

can be distinguished at the best cutoff point of IL-4, IL-5, IL-6, IL-8, IL-13, and IL-17 in the BALF of children with bronchial asthma (all P<0.05), and their AUCs were 0.79, 0.72, 0.72, 0.95, 0.94 and 0.81, respectively. The AUCs of IL-8 and IL-13 in the low FeNO group and the high FeNO group were significantly higher than the AUCs of IL-5 and IL-6 (all P<0.05). IL-5 had the highest sensitivity in distinguishing between low FeNO and high FeNO, while IL-13 had a specificity of 100%.

## Discussion

Bronchial asthma is a chronic airway inflammation involving a variety of inflammatory cells and proinflammatory cytokines. Previous studies have shown that the infiltration of inflammatory cells, such as neutrophils, eosinophils, and lymphocytes, and a cytokine network composed of type 2 inflammatory factors, such as IL-4, IL-5, and IL-13, are common pathophysiological mechanisms in patients with



**Figure 1** The ability of cytokines in the BALF of asthmatic children to differentiate between low FeNO and high FeNO. The circle on the ROC curve represents the best cutoff point. BALF, bronchoalveolar lavage fluid; FeNO, fractional exhaled nitric oxide; ROC, receiver operating characteristic.

bronchial asthma (1). Thus, it is helpful to evaluate and detect local cytokine changes in the lung to understand the pathogenesis of bronchial asthma, assess the severity of the disease, and judge the therapeutic effects of treatments.

This study showed that FEV1/FVC, MEF25, MEF50, and MEF75 in children with bronchial asthma were significantly lower than those in the control group, indicating that bronchial asthma can seriously harm the lung function and reduce the quality of life of patients. Our results are consistent with those of previous research (5). Studies have shown that in the inflammatory reaction, various lung tissue cells, such as inflammatory cells, alveolar epithelial cells, and vascular endothelial cells, convert L-arginine to L-citrulline and produce nitric oxide under the action of nitric oxide synthase (6). FeNO is closely related to the endogenous inflammatory response in lung tissue (especially eosinophilic inflammation) (7). Exhaled FeNO is moderately strongly correlated with the number of eosinophils in BALF, sputum, and blood (8). Thus, FeNO is often used as a non-invasive alternative indicator of lung inflammation in patients with bronchial asthma. In addition, there was no significant difference in lung function between the low FeNO and high FeNO groups, which suggested that lung function indexes do not reflect the level of pulmonary inflammation in children with bronchial asthma accurately and early.

The IL-4, IL-5, IL-6, IL-8, IL-13, and IL-17 levels in the BALF of asthmatic children were significantly

higher than those of the control group, and positively correlated with FeNO. Cytokines IL-4, IL-5, and IL-13 belong to type 2 cytokines, and promote airway eosinophil infiltration, mucus formation, airway hyperresponsiveness, and immunoglobulin E production in bronchial asthma (9). Consistent with the results of this study, Domvri *et al.* (10) found that the serum levels of IL-4, IL-5, IL-6, IL-13, and IL-17 in patients with bronchial asthma were significantly higher than those of patients in the control group, and IL-17 was significantly correlated with FeNO. Paro-Heitor *et al.* found that serum IL-5 levels were significantly associated with children's exhaled FeNO results, but FeNO was not significantly correlated with FEV1 and other lung function indicators (11,12). Cytokine IL-6 can be produced by inflammatory cells, vascular endothelial cells, and fibroblasts. Its primary biological effect in bronchial asthma is to promote tissue and cell damage (13). IL-6 in the serum, sputum, and BALF of patients with bronchial asthma was significantly increased (14). Reducing the expression of IL-6 can reduce the inflammation level of asthma lung tissue and improve lung function (15). IL-8 strongly attracts neutrophils to the inflammatory site, and IL-8 in the serum and BALF of asthma patients was significantly increased (16). IL-17 has proinflammatory factors secreted by Th17 cells, is significantly increased in asthma and other inflammatory diseases, and is significantly correlated with the severity of the disease (17).

There was no significant correlation between pulmonary function indexes and cytokines in the BALF of children with bronchial asthma, which suggested that pulmonary function indexes could not determine the degree of local inflammatory reaction in lung tissue. Huang *et al.* found that serum IL-4 and IL-13 were significantly negatively correlated with FEV1/FVC in 96 adult patients with bronchial asthma (18). Conversely, Grubek-Jaworska *et al.* found that sputum IL-13 was not associated with FEV1/FVC (19). The differences in these results may be due to the different characteristics of the patients, the different samples, and other factors that may affect the expression of cytokines.

The ROC curves showed that cytokines in the alveoli lavage fluid of asthmatic children had a certain value in the auxiliary identification of FeNO. For example, the AUC of IL-13 in identifying low FeNO and high FeNO was 0.94, with a specificity of up to 100%. Further research should be conducted to determine whether FeNO reflects the local inflammatory activity of lung tissue. In previous studies, the relationship between sputum or serum cytokine levels and

FeNo has been detected (20). However, the cytokines in the alveolar lavage fluid can more directly reflect the local inflammation level of the lung tissue, so the results obtained are more reliable.

In conclusion, the cytokines IL-4, IL-5, IL-6, IL-8, IL-13, and IL-17 in the alveoli lavage fluid of asthmatic children were significantly correlated with the nitric oxide in the exhaled air, which can be used to evaluate airway inflammation levels of children with asthma.

### Acknowledgments

*Funding:* None.

### Footnote

*Reporting Checklist:* The authors have completed the STARD reporting checklist. Available at <https://dx.doi.org/10.21037/tp-21-322>

*Data Sharing Statement:* Available at <https://dx.doi.org/10.21037/tp-21-322>

*Conflicts of Interest:* All authors have completed the ICMJE uniform disclosure form (available at <https://dx.doi.org/10.21037/tp-21-322>). The authors have no conflicts of interest to declare.

*Ethical Statement:* The authors are accountable for all aspects of the work, including ensuring that any questions related to the accuracy or integrity of any part of the work have been appropriately investigated and resolved. All the procedures in this study involving human participants were performed in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by the Ethics Committee of Xinxiang Central Hospital, The Fourth Clinical College of Xinxiang Medical University (No.:2017-006), and informed consent was obtained from all the patients' guardians.

*Open Access Statement:* This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the

formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

### References

1. Mims JW. Asthma: definitions and pathophysiology. *Int Forum Allergy Rhinol* 2015;5 Suppl 1:S2-6.
2. Vuolo F, Petronilho F, Sonai B, et al. Evaluation of Serum Cytokines Levels and the Role of Cannabidiol Treatment in Animal Model of Asthma. *Mediators Inflamm* 2015;2015:538670.
3. Zeng D, Li W, Zhou J, et al. Analysis of the immunoglobulin E molecular sensitization profile in children with allergic asthma and predictive factors for the efficacy of allergy immunotherapy. *Ann Transl Med*. 2020;8:1459.
4. Munakata M. Exhaled nitric oxide (FeNO) as a non-invasive marker of airway inflammation. *Allergol Int* 2012;61:365-72.
5. McGeachie MJ, Yates KP, Zhou X, et al. Patterns of Growth and Decline in Lung Function in Persistent Childhood Asthma. *N Engl J Med* 2016;374:1842-52.
6. Berry M, Hargadon B, Morgan A, et al. Alveolar nitric oxide in adults with asthma: evidence of distal lung inflammation in refractory asthma. *Eur Respir J* 2005;25:986-91.
7. Pijnenburg MW, De Jongste JC. Exhaled nitric oxide in childhood asthma: a review. *Clin Exp Allergy* 2008;38:246-59.
8. Fleming L, Tsartsali L, Wilson N, et al. Longitudinal relationship between sputum eosinophils and exhaled nitric oxide in children with asthma. *Am J Respir Crit Care Med* 2013;188:400-2.
9. Lambrecht BN, Hammad H, Fahy JV. The Cytokines of Asthma. *Immunity* 2019;50:975-91.
10. Domvri K, Porpodis K, Tzimagiorgis G, et al. Th2/Th17 cytokine profile in phenotyped Greek asthmatics and relationship to biomarkers of inflammation. *Respir Med* 2019;151:102-10.
11. Paro-Heitor ML, Bussamra MH, Saraiva-Romanholo BM, et al. Exhaled nitric oxide for monitoring childhood asthma inflammation compared to sputum analysis, serum interleukins and pulmonary function. *Pediatr Pulmonol* 2008;43:134-41.
12. Liu XT, Wang GL, Rong JY, et al. Correlation between fractional exhaled nitric oxide and airway reversibility in children with IgE-mediated asthma. *Zhongguo Dang Dai Er Ke Za Zhi* 2019;21:1172-6.
13. Rincon M, Irvin CG. Role of IL-6 in asthma and

- other inflammatory pulmonary diseases. *Int J Biol Sci* 2012;8:1281-90.
14. Tillie-Leblond I, Pugin J, Marquette CH, et al. Balance between proinflammatory cytokines and their inhibitors in bronchial lavage from patients with status asthmaticus. *Am J Respir Crit Care Med* 1999;159:487-94.
  15. Morjaria JB, Babu KS, Vijayanand P, et al. Sputum IL-6 concentrations in severe asthma and its relationship with FEV1. *Thorax* 2011;66:537.
  16. Papaporfyriou A, Papaioannou AI, Hillas G, et al. Inflammatory profile in optimally treated patients with adult versus early-onset asthma. *Postgrad Med* 2019;131:324-9.
  17. Agache I, Ciobanu C, Agache C, et al. Increased serum IL-17 is an independent risk factor for severe asthma. *Respir Med* 2010;104:1131-7.
  18. Huang AX, Lu LW, Liu WJ, et al. Plasma Inflammatory Cytokine IL-4, IL-8, IL-10, and TNF- $\alpha$  Levels Correlate with Pulmonary Function in Patients with Asthma-Chronic Obstructive Pulmonary Disease (COPD) Overlap Syndrome. *Med Sci Monit* 2016;22:2800-8.
  19. Grubek-Jaworska H, Paplińska M, Hermanowicz-Salamon J, et al. IL-6 and IL-13 in induced sputum of COPD and asthma patients: correlation with respiratory tests. *Respiration* 2012;84:101-7.
  20. Badar A, Salem AM, Bamasa AO, et al. Association Between FeNO, Total Blood IgE, Peripheral Blood Eosinophil and Inflammatory Cytokines in Partly Controlled Asthma. *J Asthma Allergy* 2020;13:533-43.
- (English Language Editor: L. Huleatt)

**Cite this article as:** Lou Y, Ke Q, Cui H, Shang Y, Yang C. Correlation study of cytokine levels in alveolar lavage fluid with exhaled nitric oxide and lung function in children with bronchial asthma. *Transl Pediatr* 2021;10(8):2069-2075. doi: 10.21037/tp-21-322