



Relationship between glucagon-like peptide-1 receptor gene polymorphism and bone mineral density in postmenopausal women in Shanghai

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Background: Glucagon-like peptide-1 receptor (GLP-1R) agonists are able to inhibit bone resorption to a certain extent and improve bone formation. GLP-1R single nucleotide polymorphism (SNP) is related to its activity, but the relationship between GLP-1R SNP and osteoporosis in postmenopausal women was still unclear. This study was to investigate the association between GLP-1R SNP and bone mineral density (BMD) in postmenopausal women in Shanghai.

Methods: Eight SNPs of GLP-1R were detected (rs3765467, rs1042044, rs2268657, rs6923761, rs2268641, rs2295006, rs4714210 and rs10305420) in 884 postmenopausal women in Shanghai. The correlation between GLP-1R SNP and BMD was further assessed.

Results: The A/A genotype of rs2295006 was negatively related to lumbar vertebrae 1–4 BMD ($P < 0.05$). Allele A was negatively related to hip BMD ($P < 0.05$). There was a negative correlation between haplotype CGAGCCA and lumbar BMD, and a positive correlation between haplotype CGGGCTA and lumbar BMD. The remaining seven GLP-1R SNPs had no relationship with BMD.

Conclusions: The rs2295006 of GLP-1R is related to the BMD of postmenopausal women in Shanghai, China.

Keywords: Osteoporosis; glucagon-like peptide-1 receptor (GLP-1R); single-nucleotide polymorphisms; polymorphism; haplotypes; bone mineral density (BMD)

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Introduction

Osteoporosis is a systemic bone disease characterized by low bone mass, destruction of bone microstructure, increased bone fragility, and being prone to fractures (1). Bone mineral density (BMD) has been the most common parameter in the diagnosis of osteoporosis, but it is also affected by some genetic factors. There is evidence showing that genetic factors account for 60–80% of BMD variability (2).

Multiple studies have shown that osteoporosis syndrome is a complex polygenic disease which is the result of a combination of genetic and environmental factors (3).

Postmenopausal osteoporosis is a common disease associated with aging and significantly affects the quality of life of postmenopausal women. Diabetic osteoporosis (DOP) is also a clinically concerned disease because of its high prevalence and disability rate (4–6). It has been shown

that the bone microenvironment and bone metabolism have changed significantly in the diabetic patients (7,8). These include a large reduction in bone mineral content, a poor balance of bone turnover, a significant reduction in bone density, and other clinical symptoms of osteoporosis (9-11).

Diabetes can affect bone metabolism in a variety of ways (12-14). There is evidence showing that some commonly used glucose-lowering drugs, such as glucagon-like peptide-1 receptor (GLP-1R) agonists, can inhibit the bone resorption and improve the bone formation (15-18). Glucagon-like peptide-1 (GLP-1) is an endogenous peptide hormone and can bind to GLP-1R, exerting glucose-dependent glucose-lowering effects (19).

In the GLP-1R knockout (GLP-1R^{-/-}) mouse model (16), Yamada *et al.* found the cortical BMD decreased significantly, and the bone fragility increased markedly, which increased the risk for osteoporosis. Moreover, GLP-1R^{-/-} mice displayed significantly increased bone resorption. The study of Stojanovic *et al.* indicated a close relationship between osteoporosis and hyperlipidemia (20). Nuche-Berenguer *et al.* found GLP-1 could reverse the reduction in bone mass secondary to hyperlipidemia (21).

GLP-1R is encoded by the *GLP-1R* gene mapped on chromosome 6 (6p21) of human genome (22). The gene polymorphism varies among races and different individuals may have different genotypes. A total of 33 single nucleotide polymorphisms (SNPs) have been identified in the GLP-1R gene of CHB (Han from Beijing) population in the second phase HapMap data (<http://www.hapmap.org>).

SNP mainly refers to a DNA sequence polymorphism caused by a single nucleotide variation at the genomic level. To date, no study has been undertaken to investigate the relationship between GLP-1R SNP and osteoporosis in the postmenopausal women. After reviewing literature, eight SNPs of GLP-1R gene (rs3765467, rs1042044, rs2268657, rs6923761, rs2268641, rs2295006, rs4714210 and rs10305420) were detected in the postmenopausal women in Shanghai using the established database and the relationship between GLP-1R SNP and BMD was further assessed. Our findings may provide evidence on the effects of GLP-1 on the osteoporosis and DOP.

The study was approved by the Ethics Committee of the Sixth People's Hospital, Shanghai Jiaotong University [2014-KY-001(K)]. Han Chinese women who were treated in the Department of Osteoporosis and Osteopathy of the Sixth People's Hospital were included. The inclusion criteria were as follows: (I) women had natural menopause for more

than 1 year; (II) women did not receive anti-osteoporotic treatment (except calcium and vitamin D supplement); (III) there was no disease affecting bone metabolism.

In this study, the iMLDR[®] multiple SNP typing (23) (Shanghai Tianhao Biotechnology Co., Ltd) was used to detect eight SNPs in 884 subjects.

Postmenopausal women received dual-energy X-ray absorptiometry (GE-LUNAR Prodigy USA) for the measurement of lumbar vertebrae 1-4 (L1-4), left femoral neck (femoral neck) and total hip (total hip) BMD (g/cm³). For the quality control, the instrument was standardized once daily. The coefficient of variation (CV) of the lumbar vertebrae, femoral neck and total hip BMD measurements was 1.39%, 2.22% and 0.70%, respectively (24).

Methods

Detection of SNPs

The selection of tag SNP is based on the International Human Genome Haplotype Program (International HapMap Project. http://www.Hapmap.org/cgi-perl/gbrowse/hapmap3_B36), and the criteria were as follows: (I) the minimum mean allele frequency (MAF) was >0.05; (II) the coefficient of linkage disequilibrium (LD) r^2 was >0.8; (III) GWAS (Genome-wide association study, genome-wide association analysis) SNPs that had been reported were included in this study. Finally, eight SNPs of GLP-1R gene were detected in the present study, namely rs3765467, rs1042044, rs2268657, rs6923761, rs2268641, rs2295006, rs4714210 and rs10305420.

Amplification was achieved by multiplex PCR reaction. Each measurable allele locus ligated product was obtained after two ligation reactions. The raw data files were analyzed using GeneMapper software version 4.1 (Applied Biosystems, USA). A total of 884 postmenopausal women were analyzed.

Statistical analysis

Statistical analysis was performed using SPSS version 24.0 (IBM SPSS Statistics 24, USA). The continuous variables with normal distribution are expressed as mean \pm standard deviation ($\bar{x} \pm SD$), and the variables with abnormal distribution as median and interquartile range. The continuous variables were compared with *t*-test between two groups; the chi-square test was used to compare the categorical variables. Haploview 4.2 was used to calculate

Table 1 Baseline characteristics of subjects included in this study

Characteristics	<60 years (n=224)	≥60 years (n=660)	P
Age (years)	54.9±5.8	71.3±7.4	0.00
Height (cm)	156.2±5.2	152.0±5.4	0.00
Weight (kg)	57.6±8.4	55.2±8.5	0.027
BMI (kg/cm ²)	23.6±3.3	23.9±3.5	0.497
Blood calcium (mmol/L)	2.34 (2.27–2.40)	2.32 (2.26–2.39)	0.718
Blood phosphorus (mmol/L)	1.14 (1.03–1.23)	1.12 (1.01–1.23)	0.700
Albumin (g/L)	47.00 (46.00–49.00)	46.00 (44.00–48.00)	0.008
Alkaline phosphatase (g/L)	69.00 (56.00–80.00)	72.00 (60.00–90.00)	0.004
Creatinine (μmol/L)	54.00 (49.00–60.75)	59.00 (52.00–66.00)	0.00
25(OH)D ₃ (ng/mL)	20.92 (16.28–26.86)	21.36 (15.56–27.97)	0.87
Parathyroid hormone (pg/L)	40.65 (32.82–53.34)	42.37 (31.63–56.22)	0.128
β-Collagen specific sequence (ng/L)	403.50 (223.00–5630)	366.00 (216.75–551.00)	0.68
L1–4 BMD (g/cm ²)	0.894 (0.806–0.992)	0.859 (0.773–0.968)	0.008
Neck BMD (g/cm ²)	0.758 (0.708–0.848)	0.692 (0.623–0.761)	0.00
Total BMD (g/cm ²)	0.801 (0.727–0.895)	0.742 (0.662–0.817)	0.00

BMI, body mass index; BMD, bone mineral density; 25(OH)D₃, 25-hydroxy vitamin D₃; L1–4, lumbar vertebra 1–4.

the D' value and linkage disequilibrium coefficient (r^2) of the linkage disequilibrium (LD) between SNPs, and the haplotype region and corresponding haplotype were obtained. After adjustment for age, a linear regression model was employed to assess the relationship between GNP-1R SNPs, haplotypes and BMD of different sites in postmenopausal women. A value of $P < 0.05$ was considered statistically significant.

Results

Characteristics of subjects

A total of 907 postmenopausal women were included, but some subjects were excluded from this study because the samples were contaminated, had poor quality or were not successfully typed after one failure. Finally, 884 samples from postmenopausal women (mean age: 67.2±10.0 years) were subjected to the detection of SNPs. In addition, subjects were divided into the <60 years group and the ≥60 years group. The baseline characteristics of subjects included for the final analysis are shown in *Table 1*.

Alleles frequency and haplotype

In this study, eight SNPs were genotyped and analyzed. During the test, the detection of rs10305420 failed, and thus the remaining 7 SNPs were examined in which the minimum allele frequency (MAF) was greater than 0.01. The genotype distribution met the Hardy-Weinberg equilibrium, and the MAF of 7 SNPs was similar to the genetic variation of Beijing Han population in China (CHBS) (*Table 2*).

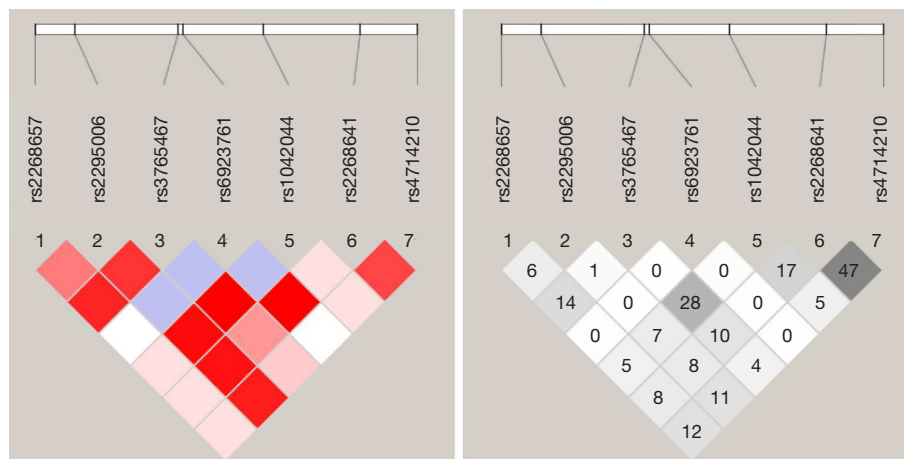
The linkage disequilibrium Lewontin's D' value and linkage disequilibrium coefficient r^2 of 7 SNPs were further calculated. There was a strong linkage disequilibrium between 7 SNPs in this study ($0.908 < D' < 1$). Thus, these 7 SNPs could be regarded as a linkage domain (*Figure 1*). Then, the haplotype and frequency in the linkage domain block were calculated, and results showed there were 15 haplotypes with the frequency greater than 1% in the linkage region (*Table 3*).

The relationship between 7 SNPs and BMD was further assessed in postmenopausal women. Results showed there was a correlation between rs2295006 and BMD at

Table 2 Seven single nucleotide polymorphisms of glucagon-like peptide-1 receptor gene

SNPs	Chr. position	SNP property	Alleles	HWE, P value	MAF in CHBS	MAF in this study
rs2268657	39020542	intron1	C/T	0.1902	0.34	0.326
rs2295006	46182304	nonsynon_exon2	G/A	0.7755	0.07	0.06
rs3765467	46182304	nonsynon_exon4	G/A	0.5521	0.23	0.255
rs6923761	39055485	nonsynon_exon5	G/A	1	0.01	0.01
rs1042044	39041502	nonsynon_exon7	C/A	1	0.47	0.46
rs2268641	39050266	intron12	C/T	0.5249	0.39	0.417
rs4714210	39055485	3'-UTR_exon13	G/A	0.1432	0.29	0.317

SNP, single nucleotide polymorphism; HWE, Hardy-Weinberg equilibrium; CHBS, Beijing Han population in China; MAF, minimum allele frequency.

**Figure 1** Chain transfer imbalance map of seven single nucleotide polymorphism of glucagon-like peptide-1 receptor gene.

specific site (Table 4). The A/A genotype of rs2295006 was negatively related to lumbar vertebrae 1–4 BMD ($P < 0.05$). There is a negative correlation between rs2295006 and BMD of lumbar vertebrae and total hip (Table 5).

The linear regression analysis was employed to evaluate the correlation between 15 haplotypes and BMD. Results showed haplotypes had no relationships with the age, height, weight and body mass index (BMI) ($P > 0.05$). A close correlation was noted between haplotypes CGAGCCA/CGGGCTA and lumbar vertebrae 1–4 BMD (Table 6).

The haplotype CGAGCCA was negatively related to the lumbar vertebrae 1–4 BMD ($P = 0.048$). There was a negative correlation between haplotype CGAGCCA and lumbar vertebrae 1–4 BMD (Table 7).

The haplotype CGGGCTA was positively related to the lumbar vertebrae 1–4 BMD ($P = 0.001$). There was a positive

correlation between haplotype CGGGCTA and lumbar vertebrae 1–4 BMD (Table 7).

Discussion

Genome wide association studies (GWAS) have confirmed that BMD is associated with multiple genetically susceptible regions (25,26). The association between vitamin D receptor gene polymorphism and osteoporosis was first reported by Morrison *et al.* in 1994 (27). Since then, more than 100 genetic polymorphisms have been identified to be associated with bone metabolism regulation, including sex hormones and their receptors, bone matrix component-related proteins and apolipoprotein E (ApoE). These findings suggest the important role of genetic factors in the pathogenesis of primary osteoporosis.

Table 3 15 haplotypes of 7 single nucleotide polymorphisms of glucagon-like peptide-1 receptor gene and their frequencies (>1%)

Index	1	2	3	4	5	6	7	8	9	10	11	12	13	14	15
rs2268657	C	C	C	T	T	C	T	C	C	C	T	C	T	C	T
rs2295006	G	G	G	G	G	G	G	G	G	G	G	G	A	A	G
rs3765467	A	G	G	G	G	G	G	G	G	A	G	G	G	G	G
rs6923761	G	G	G	G	G	G	G	G	G	G	G	G	G	G	G
rs1042044	C	A	C	A	C	A	A	A	C	C	C	C	A	A	A
rs2268641	C	C	C	C	C	T	T	T	T	T	T	T	T	T	T
rs4714210	A	A	A	A	A	A	G	A	G	G	G	A	G	G	A
Frequency	0.175	0.104	0.094	0.080	0.067	0.065	0.063	0.057	0.044	0.042	0.033	0.028	0.022	0.018	0.010

Table 4 Relationship between glucagon-like peptide-1 receptor single nucleotide polymorphisms and BMD in the postmenopausal women

Genotype	Age (years)	Height (cm)	Weight (kg)	BMI	L1–4, BMD (g/cm ²)	Femoral neck, BMD (g/cm ²)	Total hip, BMD (g/cm ²)
rs1042044							
P-dominant	0.416	0.519	0.605	0.445	0.359	0.197	0.153
P-recessive	0.674	0.115	0.072	0.275	0.768	0.638	0.416
P-addictive	0.778	0.597	0.163	0.257	0.446	0.269	0.164
rs2268641							
P-dominant	0.497	0.378	0.453	0.694	0.097	0.758	0.711
P-recessive	0.561	0.953	0.239	0.199	0.751	0.627	0.320
P-addictive	0.439	0.532	0.889	0.662	0.198	0.637	0.428
rs2268657							
P-dominant	0.275	0.087	0.894	0.478	0.804	0.889	0.886
P-recessive	0.861	0.779	0.628	0.594	0.423	0.510	0.516
P-addictive	0.359	0.145	0.911	0.430	0.577	0.683	0.851
rs2295006							
P-dominant	0.434	0.599	0.707	0.800	0.288	0.160	0.076
P-recessive	0.769	0.379	0.063	0.084	0.023*	0.152	0.101
P-addictive	0.497	0.482	0.409	0.495	0.145	0.107	0.045*
rs3765467							
P-dominant	0.838	0.261	0.661	0.934	0.229	0.421	0.256
P-recessive	0.595	0.663	0.145	0.210	0.584	0.930	0.461
P-addictive	0.958	0.293	0.340	0.640	0.236	0.543	0.228
rs4714210							
P-dominant	0.863	0.909	0.973	0.976	0.325	0.830	0.447
P-recessive	0.489	0.656	0.500	0.356	0.752	0.925	0.513
P-addictive	0.658	0.923	0.754	0.714	0.533	0.834	0.377
rs6923761							
P-dominant	0.227	0.986	0.231	0.195	0.283	0.456	0.430
P-recessive	0	0	0	0	0	0	0
P-addictive	0.227	0.986	0.231	0.195	0.283	0.456	0.430

*, P≤0.05. BMD, bone mineral density; BMI, body mass index; L1–4, lumbar vertebra 1–4.

Table 5 Correlation between glucagon-like peptide-1 receptor single nucleotide polymorphisms rs2295006 and BMD

BMD	Dominant		Recessive		Addictive	
	β	P	β	P	β	P
L1	-0.035	0.031*	0.057	0.453	-0.029	0.060
L2	-0.021	0.244	-0.067	0.428	-0.021	0.209
L3	-0.007	0.730	0.086	0.367	-0.003	0.885
L4	-0.013	0.538	-0.102	0.290	-0.015	0.429
L1-2	-0.018	0.527	-0.177	0.179	-0.023	0.387
L1-3	-0.037	0.240	-0.348	0.019*	-0.047	0.114
L1-4	-0.035	0.288	-0.354	0.023*	-0.045	0.145
L2-3	-0.055	0.057	-0.383	0.005**	-0.064	0.018*
L2-4	-0.054	0.077	-0.388	0.007**	-0.064	0.027*
L3-4	-0.061	0.025*	-0.432	0.001**	-0.071	0.005**
Neck	-0.018	0.160	-0.087	0.205	-0.019	0.107
Wards	-0.03	0.027*	-0.13	0.137	-0.032	0.013*
Troch	-0.016	0.185	-0.051	0.393	-0.016	0.154
Inter	-0.037	0.034*	-0.164	0.123	-0.039	0.016*
Total	-0.025	0.076	-0.108	0.175	-0.026	0.045*

*, $P \leq 0.05$; **, $P \leq 0.01$. BMD, bone mineral density; L, lumbar vertebra; β , regression coefficient.

It has been confirmed that the bone resorption experiences a circadian change which may be related to the secretion of incretin after eating (28). Animal studies have revealed that, in case of intake of the same energy and the same compositions of carbohydrate, protein, and fat, the BMD of rats with high frequency of food intake (more than once daily) was significantly higher than that of rats with food intake once daily (29). In addition, patients with long-term total parenteral nutrition (>3 months) often develop bone pain, hypercalciuria and elevated serum alkaline phosphatase (30). Blood calcium, blood phosphorus, 25-hydroxy vitamin D and parathyroid hormone remained normal. These findings were subsequently confirmed by many studies (31). Many investigators have therefore proposed the concept of “entero-osseous axis” (32), which means that bone metabolism may be regulated by incretin.

GLP-1 is a type of incretin, and human bone marrow mesenchymal stem cells can express GLP-1R (33). In the GLP-1R^{-/-} mouse model (16), the risk for osteoporosis increases significantly. In rats with type 2 diabetes mellitus (T2DM) and insulin-resistance (34), subcutaneous injection

Table 6 Relationship between 15 haplotypes of 7 single nucleotide polymorphisms of glucagon-like peptide-1 receptor gene and BMD in the postmenopausal women (linear regression analysis)

Haplotype	L1-4 BMD		Neck BMD		Total BMD	
	β	P	β	P	β	P
CGAGCCA	-0.037	0.048*	0.003	0.652	0.007	0.400
CGGGACA	-0.002	0.922	-0.001	0.919	-0.004	0.685
CGGGCCA	0.012	0.647	0.008	0.429	0.005	0.619
TGGGACA	-0.003	0.920	-0.007	0.514	-0.004	0.748
TGGGCCA	0.052	0.086	0.000	0.999	0.008	0.546
CGGGATA	0.007	0.807	-0.009	0.453	-0.009	0.499
TGGGATG	-0.017	0.590	0.010	0.399	0.008	0.532
CGGGATG	0.003	0.915	0.004	0.742	0.001	0.945
CGGGCTG	-0.004	0.910	-0.013	0.356	-0.018	0.271
CGAGCTG	0.013	0.732	-0.010	0.483	-0.006	0.701
TGGGCTG	0.007	0.867	-0.008	0.637	-0.019	0.292
CGGGCTA	0.154	0.001**	0.012	0.516	0.007	0.730
TAGGATG	-0.082	0.090	-0.024	0.197	-0.037	0.071
CAGGATG	0.043	0.449	-0.010	0.651	-0.011	0.641
TGGGATA	0.041	0.589	-0.005	0.867	0.015	0.643

*, $P \leq 0.05$; **, $P \leq 0.01$. BMD, bone mineral density; L1-4, lumbar vertebra 1-4; β , regression coefficient.

of GLP-1 enhanced bone synthesis in an insulin independent manner. It is speculated that this enhancement of bone synthesis may be related to the increase in OPG/RANKL ratio. In addition, dyslipidemia has been identified as a major risk factor for osteoporosis-related fractures (35,36). Most patients with osteoporosis will develop hyperlipidemia (20,37). In rats fed with high fat, results showed GLP-1 reversed the reduction of bone mass secondary to hyperlipidemia (21). The above findings suggest that the lack of GLP-1 may adversely affect the bone metabolism, and supplementation with GLP-1 can improve bone metabolism to a certain extent.

To date, only a few studies have investigated the effects of GLP-1R SNPs on the bone metabolism. In a Chinese study (38), the relationship between GLP-1R SNPs and BMD was investigated in 427 male core families. The six SNPs (rs2295006, rs3765468, rs6923761, rs1126476, rs1042044 and rs3765467) were detected in this study, and results showed no correlation between genotypes and BMD. However, rs1042044 and rs3765467 are quantitative

Table 7 Correlation between haplotype CGAGCCA/CGGGCTA and BMD

BMD	CGAGCCA		CGGGCTA	
	β	P	β	P
L1	-0.012	0.200	0.048	0.034*
L2	-0.010	0.337	0.072	0.004**
L3	-0.025	0.032*	0.055	0.049*
L4	-0.017	0.153	0.072	0.012*
L1-2	-0.026	0.097	0.124	0.001**
L1-3	-0.034	0.057	0.142	0.001**
L1-4	-0.037	0.048*	0.154	0.001**
L2-3	-0.040	0.017*	0.120	0.003**
L2-4	-0.041	0.020*	0.132	0.002**
L3-4	-0.017	0.261	0.102	0.007**
Neck	0.003	0.652	0.012	0.516
Wards	0.007	0.393	0.003	0.868
Troch	0.008	0.269	0.011	0.537
Inter	0.009	0.367	0.000	0.986
Total	0.007	0.400	0.007	0.730

*, $P \leq 0.05$; **, $P \leq 0.01$. BMD, bone mineral density; L, lumbar vertebra; β , regression coefficient.

trait loci for male lean tissue and adipose tissue mutations, respectively. The GLP-1R SNPs were significantly associated with the lean tissue and adipose tissue, suggesting that GLP-1R SNPs may indirectly affect BMD.

However, the association between GLP-1R SNPs and osteoporosis in postmenopausal women is still poorly understood. In the present study, eight SNPs of GLP-1R gene were detected according to previously reported, and the relationship between GLP-1R SNPs and BMD was further analyzed.

Our results showed, in 884 postmenopausal women, the A/A genotype of rs2295006 was negatively related to lumbar vertebrae 1–4 BMD ($P < 0.05$), and allele A was negatively related to total hip BMD ($P < 0.05$). That is, for rs2295006, only the homozygous genotype of A/A mutation had a negative correlation with lumbar vertebrae 1–4 BMD, and the negative correlation between locus and total hip BMD increases with the increase of allele A in a specific population. Therefore, in a specific individual, the rs2295006 of GLP-1R gene appears to be a homozygous

A/A mutant, which may result in decreased BMD in the lumbar vertebrae and total hip. In a specific population (such as Shanghai), if the frequency of the GLP-1R SNP rs2295006 allele A is higher, the BMD of lumbar vertebrae and total hip will be lower. Therefore, the allele A of GLP-1R rs2295006 predicts a decrease in BMD.

Of 15 haplotypes of 7 SNPs, a correlation between haplotype CGAGCCA/CGGGCTA and BMD was observed. There was a negative correlation between haplotype CGAGCCA and lumbar vertebrae BMD. The presence of haplotype CGAGCCA in the linkage domain may predicts a decrease in the lumbar vertebrae 1–4 BMD. There was a positive correlation between haplotype CGGGCTA and lumbar vertebrae BMD. That is, the presence of haplotype CGGGCTA in the linkage domain predicts an increased BMD of the lumbar vertebrae 1–4.

It has been reported that many SNPs may not directly induce the expression of disease related genes. However, they are close to certain disease genes and thus may become important markers of some diseases. These SNPs are also known as tag SNPs. The genetically determinant SNPs can affect the phenotype of some diseases and it is necessary to investigate the corresponding functions. In our study, results showed the A/A genotype of rs2295006 had a negative correlation with lumbar vertebrae 1–4 BMD. It can be regarded as a tag SNP and a determinant site associated with BMD may be close to it. This determinant site may be positively or negatively related to the BMD. This also suggests that rs2295006 can be used as a tag SNP to study multiple SNPs close to it.

In recent years, some studies have been conducted to investigate the biological effects of GLP-1R SNPs. Some studies have shown GLP-1R SNPs have a positive correlation with T2DM, islet cell function and obesity in different populations (39–42). Ma *et al.* (39) proposed that, in Han Chinese patients with T2DM, GLP-1R SNPs were associated with the risk of coronary heart disease. The GG genotype of rs4714210 predicted a lower risk of coronary heart disease as compared to the AA genotype (ORa =0.475, CIa =0.232–0.970, $p_a = 0.041$), and could also reduce the severity of atherosclerotic lesions (43). Therefore, the GLP-1R SNPs can be used as a predictor of risk for coronary heart disease in T2DM. In a Chinese study, the GLP-1R SNP rs2268657 was found to be associated with T2DM, which is mainly caused by the insufficient insulin secretion (40). In addition, rs3765467 significantly affected the response of islet B cells after infusion of GLP-1 in healthy individuals (41). Therefore, the GLP-1R SNPs may be used to guide

the therapeutic use of GLP-1R agonists. In addition, the rs2268641 of GLP-1R gene is associated with obesity in European Americans (42), and the rs6923761 of GLP-1R gene is closely related to the fasting serum GLP-1 level in newly diagnosed T2DM patients (44). Further investigation on obese patients with polycystic ovary syndrome (45) revealed that rs6923761 and rs10305420 of GLP-1R gene are associated with the therapeutic efficacy of GLP-1R agonists.

Recent studies have shown that GLP-1 is able to regulate the bone metabolism (46). In the present cross-sectional study, the correlation between GLP-1R SNPs and BMD was investigated in postmenopausal women. Our results showed A/A genotype and haplotype CGAGCCA of rs2295006 were negatively related to BMD, suggesting that this SNP may negatively regulate bone metabolism. There was a positive correlation between haplotype CGGGCTA and lumbar vertebrae BMD, and this mutation enhanced its positive correlation with BMD. There were several limitations in our study. Only 7 SNPs of GLP-1R were detected in our study and they could not represent all the GLP-1R SNPs in the postmenopausal women. Thus, the effects of other GLP-1R SNPs on the BMD in the postmenopausal women cannot be excluded. The sample size is relatively small, and subjects were included from a local region, which limit the expansion of our findings. Thus, more prospective studies with large sample size are needed to confirm our findings.

Conclusions

In summary, our study indicates a correlation between GLP-1R SNPs and BMD in the postmenopausal women. The GLP-1R SNPs can be used to guide the assessment of metabolic diseases and the treatment of osteoporosis in the postmenopausal women. However, the specific mechanism underlying the relationship between GLP-1R SNPs and BMD is needed to be further studied.

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Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/apm-19-396>).

The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was approved by the Ethics Committee of the Sixth People's Hospital, Shanghai Jiaotong University [2014-KY-001(K)].

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References

1. Association OaBMDBoCM. Guideline for the diagnosis and treatment of Primary osteoporosis (2011). Chinese Journal of Osteoporosis and Bone Mineral Research 2011;04:2-17.
2. Montazeri-Najafabady N, Dabbaghmanesh MH, Mohammadian Amiri R, et al. Influence of Estrogen Receptor Alpha Polymorphism on Bone Mineral Density in Iranian Children. Hum Hered 2019;84:82-9.
3. Ensrud KE, Crandall CJ. Osteoporosis. Ann Intern Med 2017;167:ITC17-32.
4. Okazaki R. Management of osteoporosis in diabetes mellitus. Nihon Rinsho 2009;67:1003-10.
5. Kanazawa I. Diabetes-related osteoporosis. Nihon Rinsho 2015;73:1718-22.
6. Hayes JS, Coleman CM. Diabetic Bone Fracture Repair: A Progenitor Cell-Based Paradigm. Curr Stem Cell Res Ther 2016;11:494-504.
7. Bai YY, Peng XG, Wang BH, et al. Effects of stem cell microenvironment on bone marrow-derived endothelial progenitor cells from diabetic mice. Exp Clin Cardiol 2014;20:1057-68.
8. Piscitelli P, Neglia C, Vigilanza A, et al. Diabetes and bone: biological and environmental factors. Curr Opin Endocrinol Diabetes Obes 2015;22:439-45.
9. Strotmeyer ES, Cauley JA, Schwartz AV, et al. Diabetes is associated independently of body composition with

- BMD and bone volume in older white and black men and women: The Health, Aging, and Body Composition Study. *J Bone Miner Res* 2004;19:1084-91.
10. Danielson KK, Elliott ME, LeCaire T, et al. Poor glycemic control is associated with low BMD detected in premenopausal women with type 1 diabetes. *Osteoporos Int* 2009;20:923-33.
 11. Yamamoto M, Yamaguchi T, Yamauchi M, et al. Diabetic patients have an increased risk of vertebral fractures independent of BMD or diabetic complications. *J Bone Miner Res* 2009;24:702-9.
 12. Kasahara T, Imai S, Kojima H, et al. Malfunction of bone marrow-derived osteoclasts and the delay of bone fracture healing in diabetic mice. *Bone* 2010;47:617-25.
 13. Gusova A, Pavlova M, Melnichenko G, et al. Bone mineral density in diabetic postmenopausal women with bone fracture. *Eur Soc Endocrinol* 2012;29:135.
 14. Guo Q. Healing of Bone Fracture in Type 1 Diabetic Rat Models: a Potential Gene Therapy Using Bone Morphogenetic Protein: University of Dundee; 2015.
 15. Trujillo JM, Nuffer W. GLP-1 receptor agonists for type 2 diabetes mellitus: recent developments and emerging agents. *Pharmacotherapy* 2014;34:1174-86.
 16. Yamada C, Yamada Y, Tsukiyama K, et al. The murine glucagon-like peptide-1 receptor is essential for control of bone resorption. *Endocrinology* 2008;149:574-9.
 17. Meng J, Ma X, Wang N, et al. Activation of GLP-1 Receptor Promotes Bone Marrow Stromal Cell Osteogenic Differentiation through beta-Catenin. *Stem Cell Reports* 2016;6:579-91.
 18. Ma X, Meng J, Jia M, et al. Exendin-4, a glucagon-like peptide-1 receptor agonist, prevents osteopenia by promoting bone formation and suppressing bone resorption in aged ovariectomized rats. *J Bone Miner Res* 2013;28:1641-52.
 19. Donnelly D. The structure and function of the glucagon-like peptide-1 receptor and its ligands. *Br J Pharmacol* 2012;166:27-41.
 20. Stojanovic OI, Lazovic M, Lazovic M, et al. Association between atherosclerosis and osteoporosis, the role of vitamin D. *Arch Med Sci* 2011;7:179-88.
 21. Nuche-Berenguer B, Lozano D, Gutierrez-Rojas I, et al. GLP-1 and exendin-4 can reverse hyperlipidic-related osteopenia. *J Endocrinol* 2011;209:203-10.
 22. Simó R, Hernández C. GLP-1R as a Target for the Treatment of Diabetic Retinopathy: Friend or Foe? *Diabetes* 2017;66:1453-60.
 23. Zhang XY, He JW, Fu WZ, et al. Associations of Serum Osteocalcin and Polymorphisms of the Osteocalcin Gene with Bone Mineral Density in Postmenopausal and Elderly Chinese Women. *J Nutrigenet Nutrigenomics* 2016;9:231-42.
 24. Gao G, Zhang ZL, Zhang H, et al. Hip axis length changes in 10,554 males and females and the association with femoral neck fracture. *J Clin Densitom* 2008;11:360-6.
 25. Rivadeneira F, Styrkarsdottir U, Estrada K, et al. Twenty bone-mineral-density loci identified by large-scale meta-analysis of genome-wide association studies. *Nat Genet* 2009;41:1199-206.
 26. Styrkarsdottir U, Halldorsson BV, Gretarsdottir S, et al. Multiple genetic loci for bone mineral density and fractures. *N Engl J Med* 2008;358:2355-65.
 27. Morrison NA, Qi JC, Tokita A, et al. Prediction of bone density from vitamin D receptor alleles. *Nature* 1994;367:284-7.
 28. Hygum K, Starup-Linde J, Harsløf T, et al. The diurnal variation of bone formation is attenuated in adult patients with type 2 diabetes. *Eur J Endocrinol* 2019;181:221-31.
 29. Li F, Muhlbauer RC. Food fractionation is a powerful tool to increase bone mass in growing rats and to decrease bone loss in aged rats: modulation of the effect by dietary phosphate. *J Bone Miner Res* 1999;14:1457-65.
 30. Klein GL, Targoff CM, Ament ME, et al. Bone disease associated with total parenteral nutrition. *Lancet* 1980;2:1041-4.
 31. Faienza MF, D'Amato E, Natale MP, et al. Metabolic Bone Disease of Prematurity: Diagnosis and Management. *Front Pediatr* 2019;7:143.
 32. Christensen MB, Gasbjerg LS, Heimbürger SM, et al. GIP's involvement in the pathophysiology of type 2 diabetes. *Peptides* 2020;125:170178.
 33. Sanz C, Vazquez P, Blazquez C, et al. Signaling and biological effects of glucagon-like peptide 1 on the differentiation of mesenchymal stem cells from human bone marrow. *Am J Physiol Endocrinol Metab* 2010;298:E634-43.
 34. Nuche-Berenguer B, Moreno P, Esbrit P, et al. Effect of GLP-1 treatment on bone turnover in normal, type 2 diabetic, and insulin-resistant states. *Calcif Tissue Int* 2009;84:453-61.
 35. Esposito K, Capuano A, Sportiello L, et al. Should we abandon statins in the prevention of bone fractures? *Endocrine* 2013;44:326-33.
 36. García-Gavilán JF, Bulló M, Canudas S, et al. Extra virgin olive oil consumption reduces the risk of osteoporotic

- fractures in the PREDIMED trial. *Clin Nutr* 2018;37:329-35.
37. Mandal CC. High Cholesterol Deteriorates Bone Health: New Insights into Molecular Mechanisms. *Front Endocrinol (Lausanne)* 2015;6:165.
 38. Xiang S-K. Associations of GLP-1 and its receptor gene polymorphisms with parameters related to bone metabolism: Suzhou University; 2016.
 39. Ma X, Lu R, Gu N, et al. Polymorphisms in the Glucagon-Like Peptide 1 Receptor (GLP-1R) Gene Are Associated with the Risk of Coronary Artery Disease in Chinese Han Patients with Type 2 Diabetes Mellitus: A Case-Control Study. *J Diabetes Res* 2018;2018:1054192.
 40. Zheng Y, Luo T, Zhao Y, et al. Correlation between glucagon-like peptide 1 receptor gene polymorphism and type 2 diabetes in Shanghai patients. *Chinese Journal of Endocrinology and Metabolism* 2005;21:511-3.
 41. Sathananthan A, Man CD, Micheletto F, et al. Common genetic variation in GLP1R and insulin secretion in response to exogenous GLP-1 in nondiabetic subjects: a pilot study. *Diabetes Care* 2010;33:2074-6.
 42. Li P, Tiwari HK, Lin WY, et al. Genetic association analysis of 30 genes related to obesity in a European American population. *Int J Obes (Lond)* 2014;38:724-9.
 43. Ravassa S, Zudaire A, Diez J. GLP-1 and cardioprotection: from bench to bedside. *Cardiovasc Res* 2012;94:316-23.
 44. de Luis DA, Aller R, Izaola O, et al. Role of rs6923761 gene variant in glucagon-like peptide 1 receptor in basal GLP-1 levels, cardiovascular risk factor and serum adipokine levels in naive type 2 diabetic patients. *J Endocrinol Invest* 2015;38:143-7.
 45. Jensterle M, Pirs B, Gorcar K, et al. Genetic variability in GLP-1 receptor is associated with inter-individual differences in weight lowering potential of liraglutide in obese women with PCOS: a pilot study. *Eur J Clin Pharmacol* 2015;71:817-24.
 46. Ceccarelli E, Guarino EG, Merlotti D, et al. Beyond glycemic control in diabetes mellitus: effects of incretin-based therapies on bone metabolism. *Front Endocrinol (Lausanne)* 2013;4:73.

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