



Beyond brown adipogenesis the inheritance of imprinted H19

Yanting Chen¹, Qiyuan Yang², Min Du¹

¹Nutrigenomics and Growth Biology laboratory, Department of Animal Sciences and School of Molecular Biosciences, Washington State University, Pullman, WA, USA; ²Department of Molecular, Cell, and Cancer Biology, University of Massachusetts Medical School, Worcester, MA, USA

Correspondence to: Min Du, PhD. Department of Animal Sciences, Washington State University, Pullman, WA 99164, USA. Email: min.du@wsu.edu.

Comment on: Schmidt E, Dhaouadi I, Gaziano I, et al. LincRNA H19 protects from dietary obesity by constraining expression of monoallelic genes in brown fat. *Nat Commun* 2018;9:3622.

Received: 10 November 2018; Accepted: 22 November 2018. Published: 23 November 2018.

doi: 10.21037/ncri.2018.11.05

View this article at: <http://dx.doi.org/10.21037/ncri.2018.11.05>

High-calorie food and sedentary living style give rise to the contemporary pandemics of overweight and obesity. Currently, nearly one-third of population in the world are either overweight or obese, and this number is expected to continuously increase in next decades. Obesity not only compromises glucose tolerance and insulin sensitivity, but also links to serious diseases, including type 2 diabetes, non-alcoholic fatty liver, cardiovascular diseases, and several types of cancers (1). Obesity stems from low energy expenditure by which the excess of energy is stored in white adipose tissue (WAT). However, excessive accumulation of WAT, especially in visceral fat, releases free fatty acids, inflammatory cytokines, and other mediators, which can interfere with the functions of other organs, such as brain, liver, and muscle. On the contrary, the expansion and activation of brown adipose tissue (BAT) improve energy homeostasis through non-shivering thermogenesis (NST). Recently, the discovery of substantial amount of BAT and beige adipocytes in human WAT provides a promising therapeutic target for combatting obesity and its induced complications (2).

To effectively manipulating BAT development and thermogenic activity, it is necessary to understand the molecular mechanism regulating brown adipogenesis. In the last decade, a group of principal factors have been identified, which regulate brown adipogenesis. Uncoupling protein-1 (UCP-1) is a predominant protein for NST in BAT and beige fat, though the endogenous creatine was recently identified to incur NST independent on UCP-1 (3). The presence of UCP-1 produces a futile proton cycle which dissipates proton gradient force as heat instead of synthesizing ATP. Due to the unique role of UCP-1,

expression and activity of UCP-1 are considered as a hallmark for the acquisition of BAT identity. Available studies suggest that the regulation of *ucp-1* is mainly at the transcriptional level, although sulfenylation on Cys253 residue of UCP-1 is also required for its activity, which is up-regulated by reactive oxygen species (ROS) (4). In *ucp-1* gene, several nuclear regulatory elements have been identified. For example, its promoter region contains cAMP responsive element binding (CREB) and CCAAT/enhanced binding protein (C/EBP) sites, which can rapidly respond to cAMP, and transcription factors, C/EBP α and C/EBP β ; the enhancer region of *ucp-1* gene contains the binding elements responsive to nuclear receptors, harboring retinoid X receptor (RXR), peroxisome proliferator activated receptor (PPAR) γ , PPAR α , retinoid acid receptor (RAR) or thyroid hormone receptor. In addition, several essential signaling mediators and transcriptional cofactors are also proven to regulate UCP-1 expression, such as protein kinase A (PKA), PRDM16 and PGC-1 α . PRDM16 recruits PPAR γ to activate *ucp-1* activity, indispensable for brown adipogenic commitment (5). Moreover, PGC-1 α not only coordinates the dimerization of RXR/PPAR to bind and activate *ucp-1* transcription, but also initiates mitochondrial biogenesis (6). As a result, *ucp-1* transcription is inducible and can be enhanced by dietary bioactive compounds, nutrients, and drugs, including vitamin A, metformin, thiazolidinedione and fibrates, which is considered as alternative therapeutics for overweight, obesity and type 2 diabetes.

Although an extensive list of protein-coding genes have been identified, emerging evidences underscore the importance of non-coding RNAs (ncRNAs) in tuning brown adipogenesis. Long ncRNA (lncRNA) is

conceptually defined as over 200 nucleotides in length and no protein-coding potential. These lncRNAs were considered as “junk RNA” or “sequencing noise”, but recently recognized as key players in fundamental cellular processes, such as chromatin remodeling, transcription, post-transcriptional regulations, and protein trafficking; as a result, the expression and function of lncRNA are tightly regulated, which are tissue and developmental stage dependent. Disturbance of lncRNA processing or binding targets is linked to numerous diseases, such as obesity, type 2 diabetes and cancer (7). Several lncRNAs in BAT have been identified to be essential for BAT differentiation and NST, including brown fat lncRNA 1 (Blnc1), brown adipose tissue enriched lncRNA 1 (lncBATE1), lncBATE10, and PR domain protein 16 lncRNA (lncdPRDM16).

H19 is a highly conserved lncRNA in mammalian animals, which is located on chromosome 11p15.5 in humans. It is also the first identified imprinted lncRNA and transcribes from the maternal allele. Interestingly, another imprinting gene, insulin-like growth factor 2 (*Igf2*), is also located in the locus of *H19* gene, but is a paternal imprinted gene (PIG) (8). The *H19/IGF2* cluster is one of the well defined examples of gene imprinting.

H19 involves in embryogenesis, tumorigenesis and myogenesis through diverse functional patterns (9). During embryogenesis, methyl-CpG-binding domain protein 1 (MBD1) mediates the repressive effects of H19 on PIG, including *Igf2* (10). H19 guides MBD1 to DNA domain of PIG, subsequently recruiting histone methyltransferases (HMTs) to induce histone modifications, including H3K9me3 and H3K27me3. Given that miR-675 is transcribed from an exon of H19, H19 could exert its biological effects via miR-675. In placenta, H19 maintains the regular trophoblast cell proliferation through miR-675 mediated down-regulation of Nodal signaling (11). Moreover, H19 regulates myogenesis and tumorigenesis through miR-675 and K-homology splicing regulatory protein (KSRP). During myogenesis, H19 down-regulates SMAD signaling of the bone morphogenetic protein pathway by means of miR-675, which promotes skeletal muscle differentiation (12). Additionally, H19 dismisses RNA binding protein, KSRP, and stabilizes myogenin mRNA (13). Furthermore, H19 functions as an oncogene and inhibits the activity of tumor suppressor p53 under the help of miR-675 (14).

Up to now, only a handful of studies investigate the role of H19 in adipogenesis and lipid metabolism. A previous study suggested that H19 impedes white adipogenesis of bone marrow mesenchymal stem cells, which is mediated

by miR675 (15). However, in macrophages treated by oxygenized low density lipoprotein, H19 enhances lipid accumulation and secretion of inflammatory cytokines (16). The discriminative responses of H19 to adipogenesis and lipid synthesis suggest that H19 interacts with different partners to exert context-dependent functions.

The recent study from Elena *et al.* [2018] reported that H19 activates BAT development and NST potentially through down-regulating the expression of PIG without influence on the expression of maternal imprinted gene (MIG) in BAT. Although the repressive response of H19 on differentiation of white adipocytes has been reported in the past, the regulation of H19 on BAT had not yet been explored (15). In this study, a positive link was discovered between H19 and BAT function, which was stimulated by cold stress but repressed by obesity. Gain and loss of function studies further confirmed that BAT development and thermogenic function were activated by H19, including enhanced fatty acid oxidization, mitochondrial biogenesis and NST. H19 interference in BAT discouraged BAT activity and drove mice prone to obesity, insulin resistance and glucose tolerance. Supportively, overexpression of H19 in BAT improved the functionality of BAT, rendering mice resistant to high fat diet-induced obesity. Therefore, H19 is a potential target to promote the thermogenic function of BAT, preventing obesity and metabolic disorders. Interestingly, in subcutaneous and visceral WAT, H19 expression was not altered by cold stress and obesity, indicating beige adipogenesis is potentially independent on H19. The difference between beige adipogenesis and classic BAT development could be due to different developmental origins and niche environment; brown adipocytes are derived from the muscle-like progenitor cells expressing *Myf5* and *Pax7*, while beige and WAT adipocytes originate from non-myogenic potential cells (17,18). Consistent with the effects of H19 on BAT, H19 is abundantly expressed in skeletal muscle and also identified to be required for myogenesis (12,19).

Elena *et al.* [2018] also reported that interfering H19 expression increased PIG expression without affecting MIG expression. In addition, PIG expression was tended to be decreased by cold stress and increased by obesity. Taking these evidences together, authors provided an insightful explanation that PIG expression might inhibit BAT, but its expression was elaborately repressed by H19. Regulation of H19 on imprinted gene network (IGN) has been reported previously, but the specific repression of H19 on PIG is novel although it requires to be further studied (10,20).

This study delineates a link between the expression of monoallelically imprinted genes and brown adipogenesis. Our lab intensively explores the trans-generational effects of maternal obesity on energy homeostasis in mice (21-23). Strikingly, offspring from obese subjects are susceptible to obesity, even when young. Corresponding to obesity, the BAT development and functions are also impaired majorly due to impaired adipogenic commitment. As *H19* is MIG, an inappropriate epigenetic imprinting on *H19*, such as DNA hypermethylation, may explain the inheritance of maternal obesity to obesity and associated syndromes in offspring. Previous studies have demonstrated that dysregulation of *H19/IGF2* locus can also cause severe diseases, including Beckwith-Wiedemann's Syndrome and Silver-Russell's Syndrome (24). Authors also identified that MBD1 was responsible for mediating the repressive effect of H19 on PIG. MBD1 recruited HMTs to enhance H3K9me3 and H3K27me3 on PIG, causing gene expression repression. It has been reported that *igf2* is inhibited by H19 during embryogenesis via a similar mechanism (25). Although this study provided solid evidences to demonstrate that repression of PIG was related to MBD1, the mechanism by which H19 selectively targets PIG rather than MIG keeps elusive.

Altogether, Elena *et al.* [2018] provided insightful evidence on imprinted lncRNA H19 in regulating BAT development and function. The energy homeostasis improved by H19 via BAT activation suggests that H19 is a potential therapeutic target to prevent or reduce obesity. Meanwhile, since *H19* is a MIG, the dysregulated epigenetic modification on H19 in maternal subjects could pass to the next generation, rendering them prone to obesity and other diseases.

Acknowledgments

Funding: This study was supported by grants from the National Institutes of Health (R01-HD067449 and R21-AG049976) to M Du.

Footnote

Provenance and Peer Review: This article was commissioned and reviewed by the Section Editor Dr. Jing Shi (Department of Cardiology, the First Affiliated Hospital of Nanjing Medical University, Nanjing, China).

Conflicts of Interest: All authors have completed the ICMJE

uniform disclosure form (available at <http://dx.doi.org/10.21037/ncri.2018.11.05>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

References

1. Farooqi S. Obesity genes-it's all about the parents! *Cell Metab* 2009;9:487-8.
2. Seale P, Lazar MA. Brown fat in humans: turning up the heat on obesity. *Diabetes* 2009;58:1482-4.
3. Kazak L, Chouchani ET, Jedrychowski MP, et al. A creatine-driven substrate cycle enhances energy expenditure and thermogenesis in beige fat. *Cell* 2015;163:643-55.
4. Chouchani ET, Kazak L, Jedrychowski MP, et al. Mitochondrial ROS regulate thermogenic energy expenditure and sulfenylation of UCP1. *Nature* 2016;532:112-6.
5. Kajimura S, Seale P, Kubota K, et al. Initiation of myoblast to brown fat switch by a PRDM16-C/EBP- β transcriptional complex. *Nature* 2009;460:1154.
6. Wu Z, Puigserver P, Andersson U, et al. Mechanisms controlling mitochondrial biogenesis and respiration through the thermogenic coactivator PGC-1. *Cell* 1999;98:115-24.
7. Sun L, Goff LA, Trapnell C, et al. Long noncoding RNAs regulate adipogenesis. *Proc Natl Acad Sci U S A* 2013;110:3387-92.
8. DeChiara TM, Efstratiadis A, Robertson EJ. A growth-deficiency phenotype in heterozygous mice carrying an insulin-like growth factor II gene disrupted by targeting. *Nature* 1990;345:78-80.
9. Raveh E, Matouk IJ, Gilon M, et al. The H19 Long

- non-coding RNA in cancer initiation, progression and metastasis - a proposed unifying theory. *Mol Cancer* 2015;14:184.
10. Gabory A, Ripoche MA, Le Digarcher A, et al. H19 acts as a trans regulator of the imprinted gene network controlling growth in mice. *Development* 2009;136:3413-21.
 11. Gao WL, Liu M, Yang Y, et al. The imprinted H19 gene regulates human placental trophoblast cell proliferation via encoding miR-675 that targets Nodal Modulator 1 (NOMO1). *RNA Biol* 2012;9:1002-10.
 12. Dey BK, Pfeifer K, Dutta A. The H19 long noncoding RNA gives rise to microRNAs miR-675-3p and miR-675-5p to promote skeletal muscle differentiation and regeneration. *Genes Dev* 2014;28:491-501.
 13. Giovarelli M, Bucci G, Ramos A, et al. H19 long noncoding RNA controls the mRNA decay promoting function of KSRP. *Proc Natl Acad Sci U S A* 2014;111:E5023-8.
 14. Liu C, Chen Z, Fang J, et al. H19-derived miR-675 contributes to bladder cancer cell proliferation by regulating p53 activation. *Tumour Biol* 2016;37:263-70.
 15. Huang Y, Zheng Y, Jin C, et al. Long Non-coding RNA H19 Inhibits Adipocyte Differentiation of Bone Marrow Mesenchymal Stem Cells through Epigenetic Modulation of Histone Deacetylases. *Sci Rep* 2016;6:28897.
 16. Han Y, Ma J, Wang J, et al. Silencing of H19 inhibits the adipogenesis and inflammation response in ox-LDL-treated Raw264.7 cells by up-regulating miR-130b. *Mol Immunol* 2018;93:107-14.
 17. Lepper C, Fan CM. Inducible lineage tracing of Pax7-descendant cells reveals embryonic origin of adult satellite cells. *Genesis* 2010;48:424-36.
 18. Seale P, Bjork B, Yang W, et al. PRDM16 controls a brown fat/skeletal muscle switch. *Nature* 2008;454:961-7.
 19. Milligan L, Antoine E, Bisbal C, et al. H19 gene expression is up-regulated exclusively by stabilization of the RNA during muscle cell differentiation. *Oncogene* 2000;19:5810-6.
 20. Thorvaldsen JL, Duran KL, Bartolomei MS. Deletion of the H19 differentially methylated domain results in loss of imprinted expression of H19 and Igf2. *Genes Dev* 1998;12:3693-702.
 21. Yang Q, Liang X, Sun X, et al. AMPK/ α -Ketoglutarate Axis Dynamically Mediates DNA Demethylation in the Prdm16 Promoter and Brown Adipogenesis. *Cell Metab* 2016;24:542-54.
 22. Liang X, Yang Q, Zhang L, et al. Maternal high-fat diet during lactation impairs thermogenic function of brown adipose tissue in offspring mice. *Sci Rep* 2016;6:34345.
 23. Liang X, Yang Q, Fu X, et al. Maternal obesity epigenetically alters visceral fat progenitor cell properties in male offspring mice. *J Physiol* 2016;594:4453-66.
 24. Kaffer CR, Grinberg A, Pfeifer K. Regulatory mechanisms at the mouse Igf2/H19 locus. *Mol Cell Biol* 2001;21:8189-96.
 25. Monnier P, Martinet C, Pontis J, et al. H19 lncRNA controls gene expression of the Imprinted Gene Network by recruiting MBD1. *Proc Natl Acad Sci U S A* 2013;110:20693-8.

doi: 10.21037/ncri.2018.11.05

Cite this article as: Chen Y, Yang Q, Du M. Beyond brown adipogenesis the inheritance of imprinted H19. *Non-coding RNA Investig* 2018;2:64.