



Senescence lncRNAs govern cell surface components: *lncRNA-OIS1* transcriptionally elevates DPP4

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With advancing age, senescent cells accumulate in tissues and organs, accelerating aging and age-related disease (e.g., diabetes, neurodegeneration, and cancers) (1). Cellular senescence is triggered by sublethal stresses including telomere shortening (replicative senescence), damage to DNA or other molecules (premature senescence), and oncogenic activation (oncogene-induced senescence, OIS) (2-5). Regardless of the trigger, all forms of senescence share common features like cell cycle arrest *via* p53 (TP53)/p21 (CDKN1A) and p16 (CDKN2A)/RB pathways, increased senescence-associated β -galactosidase (SA- β -Gal) activity, and the onset of a senescence-associated secretory phenotype (SASP) (6,7). OIS is tightly linked to tumor suppression, as it is elicited by unscheduled expression of oncogenic proteins such as HRAS, E2F1, RAF, BRAF, and MOS (8-11). The oncogenic protein HRAS^{G12V} (bearing a mutation of G to V at amino acid position 12 in HRAS) triggers senescence and is commonly used as a model to trigger OIS (12).

Long noncoding RNAs (lncRNAs) are transcripts longer than 200 nucleotides that generally lack protein-coding potential. RNA sequencing (RNA-seq) analysis has identified a large number of lncRNAs expressed in various cells and developmental

conditions (13-15). Although lncRNAs do not encode proteins, they are potent regulators of gene expression both at the transcriptional and post-transcriptional levels. They control transcription by modifying chromatin structure and recruiting transcriptional activators or repressors (16,17). For instance, the lncRNA *HOTAIR* (*HOX* transcript antisense RNA) is transcribed from the *HOXC* locus and mediates gene silencing of the *HOXD* locus by binding and recruiting the polycomb repressive complex 2 (PRC2) (18). Unlike *HOTAIR*, lncRNA *Evx1as* enhances *EVX1* transcription by binding to *Evx1as/EVX1* enhancer site and looping the chromatin between the promoter and enhancer. This conformation facilitates the assembly of machineries required for efficient *EVX1* transcription (19). Post-transcriptionally, lncRNAs regulate gene expression in many different ways. They can form scaffolds enabling protein assembly of complexes and act as decoys to regulate the availability of microRNAs and RNA-binding proteins (RBPs) to mRNAs (20,21). They can also influence the formation of ribonucleoprotein complexes encompassing mRNAs and RBPs (22,23); for example, *7SL* binds the 3' untranslated region (3'UTR) of *TP53* mRNA and suppresses expression of the tumor

suppressor TP53 by competing with the RBP HuR for interaction with *TP53* mRNA (24). LncRNAs can also form partial hybrids with mRNAs and thereby control mRNA turnover and translation. Together, these findings indicate that lncRNAs regulate gene expression by different mechanisms and are thus capable of regulating cellular processes like differentiation, proliferation, stress response, and senescence (25-27). Accordingly, they also influence pathologies including cardiovascular disease, cancer, diabetes, AIDS, and neurodegeneration (28).

There is growing interest in the function of lncRNAs in cellular senescence. An earlier survey of senescence-associated lncRNAs (SAL-RNAs) was conducted in replicatively senescent WI-38 fibroblasts. This study identified senescence-regulatory lncRNAs; for example, reduction of SAL-RNA1 enhanced the appearance of senescence traits (29). Other SAL-RNAs, including *TERC*, *HOTAIR*, *MALAT1*, *PINT*, *MEG3*, *ANRIL*, *Gadd7*, *7SL*, *UCA1* and *PANDA*, have been functionally linked to senescence (24,26,30). However, little is known about the lncRNAs that might be implicated in regulating OIS. Using RNA-seq analysis, Li *et al.*, have recently reported altered expression of lncRNAs upon HRAS^{G12V} induced-senescence in BJ fibroblasts. Among them, *lncRNA-OIS1* is upregulated during OIS and its silencing using shRNA selectively enhanced cell proliferation and reduced SA- β -gal activity (31).

To understand the mechanism through which *lncRNA-OIS1* regulates OIS, the authors analyzed gene expression profiles and found that cell cycle-related genes were highly enriched in *lncRNA-OIS1*-depleted cells. *In situ* hybridization (ISH) analysis indicated that *lncRNA-OIS1* is localized both in the nucleus and the cytosol, indicating that it may regulate gene expression in both compartments. As highlighted above, lncRNAs can regulate transcription in *cis* by influencing transcription in the vicinity of the locus from which they are transcribed or in *trans* by influencing transcription at a distant locus. Global run-on sequencing (GRO-seq) analysis revealed enhanced transcription of *lncRNA-OIS1* and the nearby gene *DPP4* (dipeptidyl peptidase 4, also known as CD26) upon OIS, while silencing *lncRNA-OIS1*

lowered *DPP4* mRNA production. DPP4 is an integral transmembrane glycoprotein that is widely expressed in several tissues (32). DPP4 is linked to cardiovascular diseases, metabolic diseases and cancer (33-35). It is involved in Type II diabetes mellitus (T2D), as it functions in the degradation of incretins such as glucagon-like peptide-1 (GLP-1); accordingly, a DPP4 inhibitor was developed to treat T2D and maintain insulin by preventing the degradation of incretins (36). While the specific mechanisms whereby DPP4 influences senescence are unknown, DPP4 was found to be highly abundant on the cell surface of senescent WI-38 cells and was used to target senescent cells using the antibody-dependent cell-mediated cytotoxicity (ADCC) methodology (37). Despite the robust increase of *DPP4* mRNA levels in senescent cells (33), the molecular regulators of this rise were unknown until *lncRNA-OIS1* was reported by Li *et al.*, (31). The notion that DPP4 was a key effector of the *lncRNA-OIS1*-elicited senescence was supported by the fact that silencing DPP4 restored senescence even if *lncRNA-OIS1* was silenced (31). These findings indicate that *lncRNA-OIS1* rises during OIS and transcriptionally induces DPP4, which becomes a major effector of the ensuing senescent program.

While the full set of *DPP4* transcriptional regulators is unknown, the identification of *lncRNA-OIS1* is a major step forward, paving the way for the discovery of associated transcription factors. Since DPP4 expression was found elevated in other senescent models, including replicative and premature senescence, we hypothesize that *lncRNA-OIS1* may drive DPP4 induction under different senescent triggers (Figure 1). It will be interesting to investigate if *lncRNA-OIS1* also regulates the transcription of other RNAs upregulated in senescence, both coding and noncoding. The function of *lncRNA-OIS1* in senescence *in vivo* also warrants analysis. With rising interest in devising approaches to eliminate senescent cells due to their harmful impact in older age, targeting *lncRNA-OIS1* could have therapeutic benefits, possibly reducing damaging senescence-associated processes like inflammation. Finally, since DPP4 is further linked to diseases like T2D, there could be additional therapeutic value in targeting *lncRNA-OIS1* in diabetes.

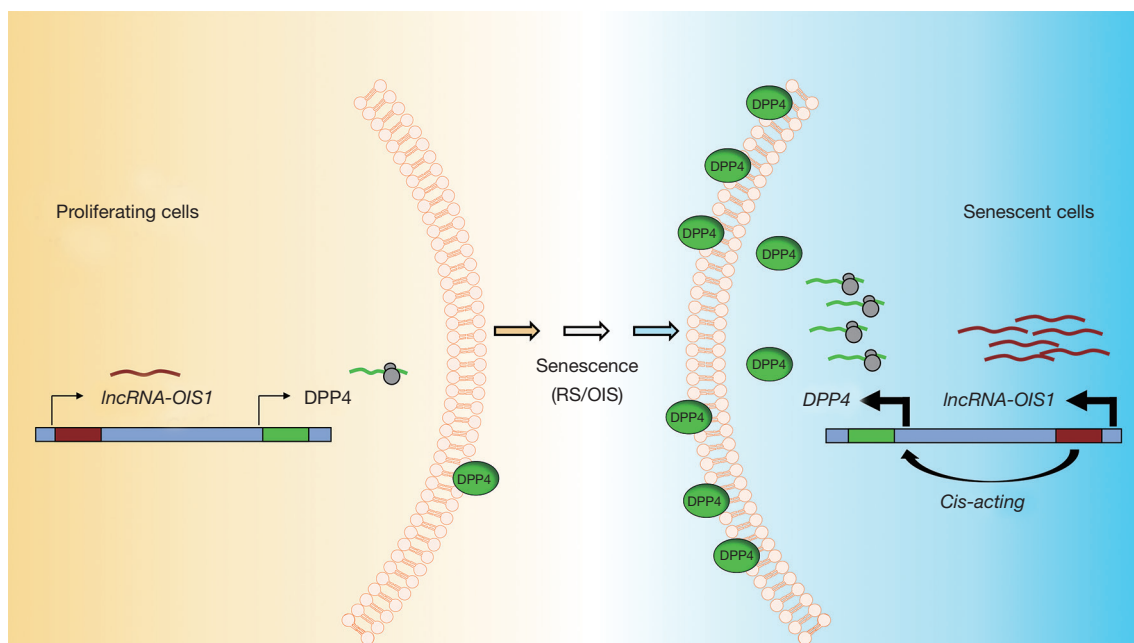


Figure 1 *lncRNA-OIS1* transcriptionally elevates DPP4 during senescence. In proliferating cells, *lncRNA-OIS1* and DPP4 are expressed at very low levels. Upon oncogene-induced senescence (OIS), *lncRNA-OIS1* is expressed to promote the transcription of *DPP4* mRNA. DPP4 in turn accumulates on the cell surface and accelerates the appearance of senescence traits.

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References

1. He SH, Sharpless NE. Senescence in Health and Disease. *Cell* 2017;169:1000-11.
2. Lopez-Otin C, Blasco MA, Partridge L, et al. The hallmarks of aging. *Cell* 2013;153:1194-217.
3. Herbig U, Ferreira M, Condel L, et al. Cellular senescence in aging primates. *Science* 2006;311:1257.
4. Campisi J. Aging, cellular senescence, and cancer. *Annu Rev Physiol* 2013;75:685-705.
5. Childs BG, Durik M, Baker DJ, et al. Cellular senescence in aging and age-related disease: from mechanisms to therapy. *Nat Med* 2015;21:1424-35.
6. Abdelmohsen K, Gorospe M. Noncoding RNA control of cellular senescence. *Wiley Interdiscip Rev RNA*

- 2015;6:615-29.
7. Munk R, Panda AC, Grammatikakis I, et al. Senescence-Associated MicroRNAs. *Int Rev Cell Mol Biol* 2017;334:177-205.
 8. Serrano M, Lin AW, McCurrach ME, et al. Oncogenic ras provokes premature cell senescence associated with accumulation of p53 and p16INK4a. *Cell* 1997;88:593-602.
 9. Zhu J, Woods D, McMahon M, et al. Senescence of human fibroblasts induced by oncogenic Raf. *Genes Dev* 1998;12:2997-3007.
 10. Dimri GP, Itahana K, Acosta M, et al. Regulation of a senescence checkpoint response by the E2F1 transcription factor and p14(ARF) tumor suppressor. *Mol Cell Biol* 2000;20:273-85.
 11. Lin AW, Barradas M, Stone JC, et al. Premature senescence involving p53 and p16 is activated in response to constitutive MEK/MAPK mitogenic signaling. *Genes Dev* 1998;12:3008-19.
 12. Kuilman T, Michaloglou C, Mooi WJ, et al. The essence of senescence. *Genes & Development* 2010;24:2463-79.
 13. Carninci P, Kasukawa T, Katayama S, et al. The transcriptional landscape of the mammalian genome. *Science* 2005;309:1559-63.
 14. Derrien T, Johnson R, Bussotti G, et al. The GENCODE v7 catalog of human long noncoding RNAs: analysis of their gene structure, evolution, and expression. *Genome Res* 2012;22:1775-89.
 15. Hangauer MJ, Vaughn IW, McManus MT. Pervasive transcription of the human genome produces thousands of previously unidentified long intergenic noncoding RNAs. *PLoS Genet* 2013;9:e1003569.
 16. Lee JT. Epigenetic regulation by long noncoding RNAs. *Science* 2012;338:1435-9.
 17. Bergmann JH, Spector DL. Long non-coding RNAs: modulators of nuclear structure and function. *Curr Opin Cell Biol* 2014;26:10-8.
 18. Tsai MC, Manor O, Wan Y, et al. Long Noncoding RNA as Modular Scaffold of Histone Modification Complexes. *Science* 2010;329:689-93.
 19. Luo S, Lu JY, Liu L, et al. Divergent lncRNAs Regulate Gene Expression and Lineage Differentiation in Pluripotent Cells. *Cell Stem Cell* 2016;18:637-52.
 20. Cesana M, Cacchiarelli D, Legnini I, et al. A Long Noncoding RNA Controls Muscle Differentiation by Functioning as a Competing Endogenous RNA (vol 147, pg 358, 2011). *Cell* 2011;147:358-69.
 21. Tay Y, Rinn J, Pandolfi PP. The multilayered complexity of ceRNA crosstalk and competition. *Nature* 2014;505:344-52.
 22. Yoon JH, Abdelmohsen K, Gorospe M. Functional interactions among microRNAs and long noncoding RNAs. *Seminars in Cell & Developmental Biology* 2014;34:9-14.
 23. Marchese FP, Raimondi I, Huarte M. The multidimensional mechanisms of long noncoding RNA function. *Genome Biol* 2017;18:206.
 24. Abdelmohsen K, Panda AC, Kang MJ, et al. 7SL RNA represses p53 translation by competing with HuR. *Nucleic Acids Res* 2014;42:10099-111.
 25. Li J, Tian H, Yang J, et al. Long Noncoding RNAs Regulate Cell Growth, Proliferation, and Apoptosis. *DNA Cell Biol* 2016;35:459-70.
 26. Grammatikakis I, Panda AC, Abdelmohsen K, et al. Long noncoding RNAs(lncRNAs) and the molecular hallmarks of aging. *Aging (Albany NY)* 2014;6:992-1009.
 27. Audas TE, Lee S. Stressing out over long noncoding RNA. *Biochim Biophys Acta* 2016;1859:184-91.
 28. Chen X, Yan CC, Zhang X, et al. Long non-coding RNAs and complex diseases: from experimental results to computational models. *Briefings in Bioinformatics* 2017;18:558-76.
 29. Abdelmohsen K, Panda A, Kang MJ, et al. Senescence-associated lncRNAs: senescence-associated long noncoding RNAs. *Aging Cell* 2013;12:890-900.
 30. Ozes AR, Miller DE, Ozes ON, et al. NF-kappaB-HOTAIR axis links DNA damage response, chemoresistance and cellular senescence in ovarian cancer. *Oncogene* 2016;35:5350-61.
 31. Li L, van Breugel PC, Loayza-Puch F, et al. LncRNA-OIS1 regulates DPP4 activation to modulate senescence induced by RAS. *Nucleic Acids Res* 2018;46:4213-27.
 32. Morimoto C, Schlossman SF. The structure and function of CD26 in the T-cell immune response. *Immunol Rev* 1998;161:55-70.
 33. Zhong J, Rao X, Rajagopalan S. An emerging role of dipeptidyl peptidase 4 (DPP4) beyond glucose control: potential implications in cardiovascular disease. *Atherosclerosis* 2013;226:305-14.
 34. Nargis T, Chakrabarti P. Significance of circulatory DPP4 activity in metabolic diseases. *IUBMB Life* 2018;70:112-9.
 35. Beckenkamp A, Davies S, Willig JB, et al. DPP4/CD26: a tumor suppressor or a marker of malignancy? *Tumour Biol* 2016;37:7059-73.

36. Duez H, Cariou B, Staels B. DPP-4 inhibitors in the treatment of type 2 diabetes. *Biochem Pharmacol* 2012;83:823-32.
37. Kim KM, Noh JH, Bodogai M, et al. Identification of senescent cell surface targetable protein DPP4. *Genes Dev* 2017;31:1529-34.

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