



Dysbiosis of the gut microbiota and colorectal cancer: the key target of molecular pathological epidemiology

Keisuke Kosumi^{1,2}, Kosuke Mima^{1,2}, Hideo Baba², Shuji Ogino^{1,3,4,5}

¹Department of Oncologic Pathology, Dana-Farber Cancer Institute and Harvard Medical School, Boston, MA, USA; ²Department of Gastroenterological Surgery, Graduate School of Medical Science, Kumamoto University, Kumamoto, Japan; ³Program in MPE Molecular Pathological Epidemiology, Department of Pathology, Brigham and Women's Hospital and Harvard Medical School, Boston, MA, USA; ⁴Department of Epidemiology, Harvard T.H. Chan School of Public Health, Boston, MA, USA; ⁵Broad Institute of MIT and Harvard, Cambridge, MA, USA

Correspondence to: Keisuke Kosumi, MD, PhD. Department of Oncologic Pathology, Dana-Farber Cancer Institute and Harvard Medical School, 450 Brookline Ave., Room SM1040, Boston, MA 02215, USA. Email: kosumi-kmm@umin.ac.jp; Shuji Ogino, MD, PhD, MS. Program in MPE Molecular Pathological Epidemiology, Department of Pathology, Brigham and Women's Hospital, 450 Brookline Ave., Room SM1036, Boston, MA 02215, USA. Email: shuji_ogino@dfci.harvard.edu.

Comment on: Guo S, Li L, Xu B, *et al.* A Simple and Novel Fecal Biomarker for Colorectal Cancer: Ratio of *Fusobacterium Nucleatum* to Probiotics Populations, Based on Their Antagonistic Effect. *Clin Chem* 2018;64:1327-37.

Received: 10 September 2018; Accepted: 24 September 2018; Published: 25 September 2018.

doi: 10.21037/jlpm.2018.09.05

View this article at: <http://dx.doi.org/10.21037/jlpm.2018.09.05>

Microbiome is a dramatically advancing research field in the study of human cancers (1). Microbial distribution varies spatially and temporally from the mouth to the rectum during the individual's lifetime. These gut bacteria maintain intestinal homeostasis by regulating various biological functions such as mucosal barrier, immunity, and metabolic processes. Accumulating evidence indicates that the gut bacteria play a crucial role in human health (1), and the dysbiosis of the gut microbiota is associated with various disorders, including obesity, diabetes, inflammatory bowel disease, and many types of cancer especially colorectal cancer. According to emerging studies, several bacterial species [*Fusobacterium nucleatum* (*F. nucleatum*), *Bacteroides fragilis*, and *Escherichia coli*] appeared to have a critical role in the colorectal carcinogenesis (2,3). In the tumor microenvironment of colorectal carcinoma, there should be a complicated community of cancer cells, immune cells, and a great number of microorganisms. Experimental studies have shown that gut bacteria activate anti-tumor immunity and boost the efficacy of immunotherapy (4,5). Since the gut microbiota is likely influenced by food, lifestyle, antibiotics, and probiotics, a better understanding of the relationship between human cancer, immune system, and gut microbiome may promote clinical application including

immunotherapy.

F. nucleatum is a common gram-negative anaerobe, which is one of the most prevalent bacteria in the oral cavity. *F. nucleatum* has been associated with inflammatory diseases including periodontitis, gingivitis, appendicitis, and liver abscess. Accumulating evidence suggests that the amount of *F. nucleatum* is remarkably higher in colorectal cancer tissue than matched normal colorectal mucosa (6-8). The fraction of colorectal cancer with *F. nucleatum* increases linearly along the large intestinal subsites from the rectum to the cecum, based on a continuum model of pathogenic influences of gut bacteria on colorectal carcinogenesis (9). An experimental study suggests that metastasis of *F. nucleatum*-present colorectal cancer might be prohibited by antimicrobial interventions (3). Human population-based studies indicate that an enrichment of *F. nucleatum* is associated with high-level microsatellite instability (MSI), advanced disease stage, low-level T-cell infiltration in colorectal carcinoma tissue (7), and worse clinical outcome (8). Collectively, *F. nucleatum* DNA in colorectal carcinoma tissue may serve as a prognostic biomarker, and provide insights for the establishment of new prevention and treatment strategies targeting bacteria in colorectal cancer. On the other hand, *Faecalibacterium prausnitzii* (*Fp*)

is an anaerobic bacterium of the most important butyrate-producing bacteria in the gut. *Fp* is inversely associated with the activity of inflammatory bowel disease, and the amount of *Fp* is reduced in colorectal cancer patients (10). The presence of members of *Bifidobacterium* genus (*Bb*) in the gut lumen may suppress colorectal carcinogenesis through prevention of enteropathogenic infection and inhibition of secondary bile acid production (11-13). *Fp* and *Bb* have gained a lot of attention in the field of colorectal cancer prevention.

In a recent article published in *Clinical Chemistry*, Guo and colleagues reported a simple fecal biomarker for detecting colorectal cancer utilizing two Chinese cohorts of 903 individuals (cohort I and cohort II) (14). To evaluate fecal dysbiosis, they first analyzed the fecal microbiota of patients with colorectal cancer (n=20) and healthy controls (n=20) by Illumina MiSeq sequencing. As a result, they found the increase of *F. nucleatum*, as well as the decreases of *Fp* and *Bb*. For further analyses, they measured the relative abundance of *F. nucleatum*, *Fp*, and *Bb* by quantitative polymerase chain reaction (PCR) in the stools of cohort I. Cohort I included 215 patients with colorectal cancer, 178 patients with benign colon disease, 100 patients with non-gastrointestinal cancer, and 156 healthy controls. The relative abundance of *F. nucleatum* was higher in colorectal cancer patients than in benign colon disease patients (P=0.043), non-gastrointestinal cancer patients (P<0.0001), and healthy controls (P<0.0001). Regarding fecal *Fp* and *Bb*, both relative abundances were lower in colorectal cancer patients than in non-gastrointestinal cancer patients (P=0.0001; P=0.05) and healthy controls (both P<0.0001). To investigate the association of dysbiosis with colorectal cancer, they focused on the ratio of *F. nucleatum* to *Fp* (*Fn/Fp*) and *F. nucleatum* to *Bb* (*Fn/Bb*). *Fn/Fp* was significantly higher in colorectal cancer patients than in non-gastrointestinal cancer patients and healthy controls (both P<0.0001). *Fn/Bb* was significantly higher in colorectal cancer patients than in the other three groups (all P<0.0001). As the next step, they evaluated the diagnostic performance of the microbial ratios, and investigated the antagonistic effect of *Fn* against two different indicator bacteria (*Fp* and *Bb*). *Fn/Bb* had a superior sensitivity of 85% and specificity of 92% in detecting colorectal cancer patients from healthy controls [area under the curve (AUC) =0.91]. The other AUCs were 0.88 for *F. nucleatum*, 0.74 for *Fp*, 0.87 for *Bb*, and 0.91 for *Fn/Fp*. The combination of *Fn/Bb* and *Fn/Fp* improved the diagnostic value (AUC =0.94). The AUCs to distinguish colorectal

cancer patients from benign colon disease were 0.84 for *F. nucleatum*, 0.55 for *Fp*, 0.75 for *Bb*, 0.53 for *Fn/Fp*, 0.92 for *Fn/Bb*, and 0.92 for the combination of *Fn/Bb* and *Fn/Fp*. Moreover, they investigated these five AUCs to distinguish early stage colorectal cancer patients (stage I + II) from healthy controls. Each AUC was 0.64 for *F. nucleatum*, 0.75 for *Fp*, 0.69 for *Bb*, 0.82 for *Fn/Fp*, 0.88 for *Fn/Bb*, and 0.89 for the combination of *Fn/Bb* and *Fn/Fp*. The combination of *Fn/Bb* and *Fn/Fp* offered 60% specificity and 90% sensitivity in detecting only stage I of colorectal cancer (AUC =0.80). As a validation set, the cohort II including 152 patients with colorectal cancer and 102 healthy controls was utilized. The performance for identifying colorectal cancer was also confirmed in the validation cohort II. Finally, Guo and colleagues concluded that the ratio of *Fn* to the important bacterium, such as *Fp* or *Bb*, was identified as a useful biomarker for screening early colorectal cancer.

There have been great interests in developing non-invasive methods, such as a stool screening with safety, efficiency, convenience, and cost-effective. The effectiveness of guaiac fecal occult blood test and fecal immunochemical testing have been established to reduce both colorectal cancer incidence and mortality, thus allowing their use for many colorectal cancer screening programs. However, the issue is that the specificity of these techniques has been relatively low. Colon and rectal cancers represent heterogeneous sets of neoplasms with differing combinations of genetic and epigenetic alterations, the accrual of which is influenced by complicated interplay between tumor cells, host cells, and microorganisms. The analysis of mucosal tissues may be more appropriate to evaluate the mechanisms of microbiota which are physiopathologically involved in colorectal cancer development. However, considering that the invasiveness of classic endoscopic approaches or endoscopic biopsy, collection of mucosal samples is more challenging to conduct than that of fecal samples, particularly in healthy individuals. Consequently, new molecular colorectal cancer screening methods targeting microbiome in stools are required. A vast majority of studies on the diversity of the microbiota in human gut have been conducted on fecal samples to identify more competent diagnostic markers for colorectal cancer.

Shah and colleagues recently conducted the first microbiome-based meta-analysis for colorectal cancer to identify a common microbial marker in fecal samples, utilizing the results from nine studies including 79 colorectal adenomas patients, 195 colorectal cancer

patients and 235 controls (15). They emphasized a marked increase in *Parvimonas micra* ATCC 33270, *Streptococcus anginosus*, *Parabacteroides distasonis*, and other members of *Proteobacteria* in addition to the previously reported taxa such as *Fusobacterium* species. In their study, the AUC to distinguish colorectal cancer from controls was approximate 0.80. According to another recent study by Liang and colleagues, the quantification of *F. nucleatum* in stools by quantitative PCR could discriminate colorectal cancer from healthy controls with a high sensitivity of 78% and specificity of 80% (AUC =0.87) (16). Compare to fecal samples of healthy individuals, those of colorectal cancer patients have modifications in the microbiota composition, namely dysbiosis. Although *F. nucleatum* has continuously gained much attention for a possible role in initiating and progressing colorectal cancer, comprehensive analyses of the human microbiome ecosystem utilizing 16S ribosomal RNA (rRNA) next-generation sequencing (NGS) are strongly required. However, at this point, there is no consensus in terms of the dysbiosis of fecal samples for screening and diagnosis. Analyses utilizing quantitative PCR and 16S rRNA NGS are the most used approaches for investigating the composition of the gut bacterial community. However, it is challenging to define the gut dysbiosis because of some reasons. First, the gut microbial ecosystem is likely affected by a number of factors, such as race, age, sex, diet, lifestyle, medication, and supplemental use. Second, there is the diversity of methods including the regions of the 16S rRNA targeted (V1, V2, V3, V4, V5, and V6 regions) for the NGS approach. Due to the complexity of gut microbial ecosystem and methods, there is no consensus in terms of the dysbiosis observed in colorectal cancer compared to healthy individuals using quantitative PCR or 16S rRNA NGS in fecal samples. It is reasonable to assume that there is heterogeneity of microbiome in colorectal tumor microenvironment or stools of colorectal cancer patients. The microbial heterogeneity in colorectal cancer may make it challenging to define the microbial dysbiosis for screening and diagnosis of colorectal cancer. Further investigations are warranted to consider the microbial heterogeneity in colorectal cancer, not only for screening but also classification of colorectal cancer.

We argue that molecular pathological epidemiology (MPE) can establish a basis for individualized cancer prevention and treatments (17-20). MPE is a transdisciplinary integrative research field which has been derived from efforts to integrate the methodology of molecular pathology into population-based epidemiologic research. Conventional

epidemiology typically investigates the relationships between epidemiologic exposures and a certain single disease entity based on the assumption that the single disease entity should represent a homogeneous process including disease course, etiologies, and pathogenesis. Beyond this conventional approach, MPE research attempts to decrypt differential associations of an exposure with several distinct subtypes classified by molecular or pathological features of the disease. Microbial MPE addresses etiologic heterogeneity according to subgroups of colorectal cancer classified by tumor tissue microbial profiling (21). Given the evidence that *F. nucleatum*-positive colorectal cancers may have some distinct features (22), it is of great interest to investigate the association of environmental factors with colorectal cancer in strata of tumor *F. nucleatum* status. Utilizing two large prospective cohorts, we reported that prudent diets (rich in whole grains and dietary fiber) are correlated with a lower risk for colorectal cancers with *F. nucleatum* but not ones without *F. nucleatum* (23). Specific nutritional components may induce intestinal inflammation, and the dietary inflammatory effects can be estimated based on an empiric dietary inflammatory pattern (EDIP) score. We found that higher EDIP scores were associated with increased risk of colorectal cancers with *F. nucleatum*, especially proximal colon cancers (24). These findings suggest that diet changes the gut microbiome to promote or inhibit colorectal carcinogenesis, and dietary interventions could be useful for cancer prevention and precision medicine. Emerging evidence also suggests potential roles of several bacteria such as *Bacteroides fragilis*, *Escherichia coli*, *Parvimonas micra*, *Fp*, and *Bb* on colorectal carcinogenesis. There has been an increasing understanding that colorectal cancer represents a heterogeneous group of tumor cells which develop through the multilevel accumulation of various combinations of molecular alterations in the colorectal epithelium. In fact, a huge number of microorganisms configure the bacterial ecosystem in gut lumen as well as tumor microenvironment, and those form a complicated network system through interactions (25). Emerging evidence indicates the heterogeneity of the microbial community in stool samples (15). Therefore, we seek the comprehensive analyses of the human microbiome ecosystem in stools utilizing 16S rRNA NGS to classify colorectal cancers into subtypes based on the heterogeneity of bacteria. If such a test is established, the fusion of MPE approach and screening test targeting bacteria can make it possible to advance our understanding of disease pathogenesis and

provide appropriate precision medicine to individuals.

In summary, the population-based study by Guo *et al.* supports the existing evidence on utility of fecal analyses targeting bacteria. Considering the heterogeneity of genetic and epigenetic changes in colorectal cancer, colorectal cancer may have the heterogeneity of microbiome in stools as well as in tumor microenvironment. Comprehensive analyses of the human microbiome ecosystem utilizing 16S rRNA NGS are strongly required, and the analyses must help us to understand colorectal carcinogenesis and subtypes of colorectal cancers more accurately. MPE research is a transdisciplinary integrative research field, which intrinsically pursue the addresses the disease heterogeneity in human population. We propose that MPE research combined with fecal bacterial analyses can play critical roles in providing rationales and discovering insights into precision medicine for colorectal cancer.

Acknowledgments

Funding: This work was supported by U.S. National Institutes of Health (NIH) grants (R35 CA197735 to S Ogino); by Nodal Award (2016-02) from the Dana-Farber Harvard Cancer Center (to S Ogino). This work was supported in part by a Grant-in-Aid for Scientific Research from the Japan Society for the Promotion of Science [grant number 16K19941 (to K Kosumi) and 17H05094 (to K Mima)] and JSPS Fujita Memorial Fund for Medical Research (to K Kosumi). K Kosumi was supported by a grant from Overseas Research Fellowship from Japan Society for the Promotion of Science (JP2017-775). K Mima was supported by grants from Takeda Science Foundation, KANAE Foundation for the Promotion of Medical Science, YOKOYAMA Foundation for Clinical Pharmacology, and the Uehara Memorial Foundation. The content is solely the responsibility of the authors and does not necessarily represent the official views of NIH. The funders had no role in submit the manuscript to publication, or preparation of the manuscript.

Footnote

Provenance and Peer Review: This article was commissioned and reviewed by Section Editor Wei Li (Department of Clinical Microbiology & Infectious Diseases, Qilu Hospital of Shandong University, Jinan, China).

Conflicts of Interest: All authors have completed the ICMJE

uniform disclosure form (available at <http://dx.doi.org/10.21037/jlpm.2018.09.05>). This work was supported by U.S. National Institutes of Health (NIH) grants (R35 CA197735 to S Ogino); by Nodal Award (2016-02) from the Dana-Farber Harvard Cancer Center (to S Ogino). This work was supported in part by a Grant-in-Aid for Scientific Research from the Japan Society for the Promotion of Science [grant number 16K19941 (to K Kosumi) and 17H05094 (to K Mima)] and JSPS Fujita Memorial Fund for Medical Research (to K Kosumi). K Kosumi was supported by a grant from Overseas Research Fellowship from Japan Society for the Promotion of Science (JP2017-775). K Mima was supported by grants from Takeda Science Foundation, KANAE Foundation for the Promotion of Medical Science, YOKOYAMA Foundation for Clinical Pharmacology, and the Uehara Memorial Foundation. The content is solely the responsibility of the authors and does not necessarily represent the official views of NIH. The funders had no role in submit the manuscript to publication, or preparation of the manuscript. Dr. Baba has nothing to disclose. The authors have no other conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

References

1. Routy B, Gopalakrishnan V, Daillere R, et al. The gut microbiota influences anticancer immunosurveillance and general health. *Nat Rev Clin Oncol* 2018;15:382-96.
2. Brennan CA, Garrett WS. Gut Microbiota, Inflammation, and Colorectal Cancer. *Annu Rev Microbiol* 2016;70:395-411.
3. Bullman S, Pedamallu CS, Sicinska E, et al. Analysis of *Fusobacterium* persistence and antibiotic response in

- colorectal cancer. *Science* 2017;358:1443-8.
4. Gopalakrishnan V, Helmink BA, Spencer CN, et al. The Influence of the Gut Microbiome on Cancer, Immunity, and Cancer Immunotherapy. *Cancer Cell* 2018;33:570-80.
 5. Gopalakrishnan V, Spencer CN, Nezi L, et al. Gut microbiome modulates response to anti-PD-1 immunotherapy in melanoma patients. *Science* 2018;359:97-103.
 6. Kostic AD, Chun E, Robertson L, et al. *Fusobacterium nucleatum* potentiates intestinal tumorigenesis and modulates the tumor-immune microenvironment. *Cell Host Microbe* 2013;14:207-15.
 7. Mima K, Sukawa Y, Nishihara R, et al. *Fusobacterium nucleatum* and T Cells in Colorectal Carcinoma. *JAMA Oncol* 2015;1:653-61.
 8. Mima K, Nishihara R, Qian ZR, et al. *Fusobacterium nucleatum* in colorectal carcinoma tissue and patient prognosis. *Gut* 2016;65:1973-80.
 9. Mima K, Cao Y, Chan AT, et al. *Fusobacterium nucleatum* in Colorectal Carcinoma Tissue According to Tumor Location. *Clin Transl Gastroenterol* 2016;7:e200.
 10. Ferreira-Halder CV, Faria AVS, Andrade SS. Action and function of *Faecalibacterium prausnitzii* in health and disease. *Best Pract Res Clin Gastroenterol* 2017;31:643-8.
 11. Ubeda C, Djukovic A, Isaac S. Roles of the intestinal microbiota in pathogen protection. *Clin Transl Immunology* 2017;6:e128.
 12. Laforest-Lapointe I, Arrieta MC. Patterns of Early-Life Gut Microbial Colonization during Human Immune Development: An Ecological Perspective. *Front Immunol* 2017;8:788.
 13. Fukuda S, Toh H, Hase K, et al. Bifidobacteria can protect from enteropathogenic infection through production of acetate. *Nature* 2011;469:543-7.
 14. Guo S, Li L, Xu B, et al. A Simple and Novel Fecal Biomarker for Colorectal Cancer: Ratio of *Fusobacterium nucleatum* to Probiotics Populations, Based on Their Antagonistic Effect. *Clin Chem* 2018;64:1327-37.
 15. Shah MS, DeSantis TZ, Weinmaier T, et al. Leveraging sequence-based faecal microbial community survey data to identify a composite biomarker for colorectal cancer. *Gut* 2018;67:882-91.
 16. Liang Q, Chiu J, Chen Y, et al. Fecal Bacteria Act as Novel Biomarkers for Noninvasive Diagnosis of Colorectal Cancer. *Clin Cancer Res* 2017;23:2061-70.
 17. Ogino S, Nowak JA, Hamada T, et al. Integrative analysis of exogenous, endogenous, tumour and immune factors for precision medicine. *Gut* 2018;67:1168-80.
 18. Ogino S, Jhun I, Mata DA, et al. Integration of pharmacology, molecular pathology, and population data science to support precision gastrointestinal oncology. *NPJ Precis Oncol* 2017;1.
 19. Ogino S, Giannakis M. Immunoscore for (colorectal) cancer precision medicine. *Lancet* 2018;391:2084-6.
 20. Ogino S, Nowak JA, Hamada T, et al. Insights into Pathogenic Interactions Among Environment, Host, and Tumor at the Crossroads of Molecular Pathology and Epidemiology. *Annu Rev Pathol* 2018. [Epub ahead of print].
 21. Hamada T, Keum N, Nishihara R, et al. Molecular pathological epidemiology: new developing frontiers of big data science to study etiologies and pathogenesis. *J Gastroenterol* 2017;52:265-75.
 22. Mima K, Ogino S, Nakagawa S, et al. The role of intestinal bacteria in the development and progression of gastrointestinal tract neoplasms. *Surg Oncol* 2017;26:368-76.
 23. Mehta RS, Nishihara R, Cao Y, et al. Association of Dietary Patterns With Risk of Colorectal Cancer Subtypes Classified by *Fusobacterium nucleatum* in Tumor Tissue. *JAMA Oncol* 2017;3:921-7.
 24. Liu L, Tabung FK, Zhang X, et al. Diets That Promote Colon Inflammation Associate With Risk of Colorectal Carcinomas That Contain *Fusobacterium nucleatum*. *Clin Gastroenterol Hepatol* 2018. [Epub ahead of print].
 25. Kosumi K, Hamada T, Koh H, et al. The Amount of Bifidobacterium Genus in Colorectal Carcinoma Tissue in Relation to Tumor Characteristics and Clinical Outcome. *Am J Pathol* 2018. (In press).

doi: 10.21037/jlpm.2018.09.05

Cite this article as: Kosumi K, Mima K, Baba H, Ogino S. Dysbiosis of the gut microbiota and colorectal cancer: the key target of molecular pathological epidemiology. *J Lab Precis Med* 2018;3:76.