



Can cellular and humoral immunity predict response to BNT162b2 bivalent booster?

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Predicting response to coronavirus disease 2019 (COVID-19) vaccination is an important basis for future interventions to combat severe acute respiratory syndrome coronavirus 2 (SARS-CoV-2) during the foreseeable endemic phase, since ensuring sufficient humoral and cellular immunity is critical to minimizing the impact of the virus on the more susceptible segments of the population (1). To this end, we conducted a retrospective analysis of an ongoing serosurveillance protocol (2), to determine whether cellular or natural immunity can be used as a reliable marker of response to administration of the new bivalent COVID-19 vaccines.

We retrospectively analyzed 51 healthcare employees of the Hospital Pederzoli (Peschiera del Garda, Verona, Italy) previously vaccinated and boosted with the Pfizer/BioNTech mRNA monovalent BNT162b2 vaccine (Comirnaty, Pfizer Inc., NY, USA), before and 15 days after receiving a single BNT162b2 bivalent booster (Comirnaty, Pfizer Inc.) ≥ 1 year after the last monovalent dose of vaccine. Basal and post-bivalent booster immunity was assayed using the specific SARS-CoV-2 Cobas interferon gamma release assay (IGRA), and by measuring total anti-SARS-CoV-2 antibodies with Roche Elecsys (Roche Diagnostics, Basel, Switzerland) anti-SARS-CoV-2 electrochemiluminescence immunoassay (ECLIA). Positive response after BNT162b2 bivalent vaccine administration was defined as positive variation from the pre-vaccination value. Test results were analyzed with Analyse-it (Analyse-it Software Ltd., Leeds, UK). All subjects provided a written informed consent

for participating to the study. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013) and approved by the Ethics Committee of Verona and Rovigo Provinces (59COVIDCESC; November 8, 2021).

Both IGRA and total anti-SARS-CoV-2 antibodies levels increased after bivalent vaccine booster from 1.00 ± 1.62 to 1.07 ± 0.90 ng/mL ($P=0.385$), and from $12,193 \pm 7,646$ to $21,530 \pm 5,060$ kU/L ($P<0.001$), respectively. A positive IGRA response (i.e., >0.013 ng/mL) was detected in 44/51 (86.3%) and 50/51 (98.0%) subjects before and after vaccination, respectively (chi-square, 3.391; $P=0.033$). Instead, the serum of all subjects was reactive for total anti-SARS-CoV-2 antibodies both before and after vaccination (i.e., >0.8 kU/L: 51/51; 100%). In receiver operating characteristic (ROC) curve analysis, SARS-CoV-2 IGRA was a poor predictor of response to bivalent BNT162b2 vaccine [area under the curve (AUC), 0.63; 95% CI: 0.52–0.74; $P=0.010$], whereas better performance was observed for total anti-SARS-CoV-2 antibodies (AUC, 0.83; 95% CI: 0.75–0.91; $P<0.001$) (Figure 1). A threshold 11,723 kU/L for total anti-SARS-CoV-2 antibodies was associated with 0.96 sensitivity and a specificity of 0.59 for predicting response to bivalent BNT162b2 vaccine.

The results of this retrospective observational study suggest that SARS-CoV-2 IGRA may be a poor predictor of response to bivalent BNT162b2 vaccine in healthy individuals, likely because basal T cell immunity levels are already elevated at baseline due to repeated vaccination and/or natural infection. In contrast, monitoring of total anti-

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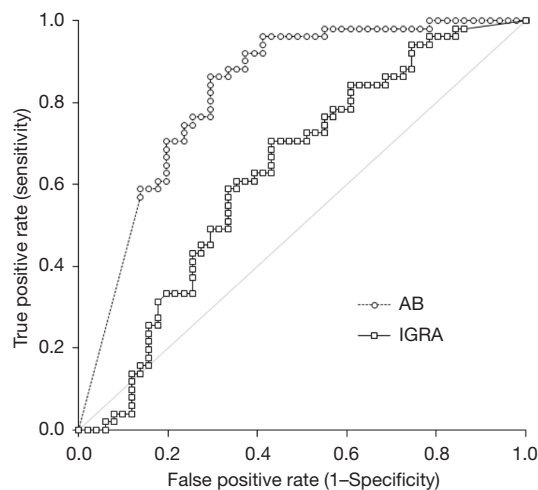


Figure 1 Receiver operating characteristic curve analysis of SARS-CoV-2 Cobas IGRA and Roche Elecsys anti-SARS-CoV-2 total AB for predicting response to bivalent BNT162b2 vaccination. AB, antibodies; IGRA, interferon gamma release assay; SARS-CoV-2, severe acute respiratory syndrome coronavirus 2.

SARS-CoV-2 antibodies remains a better option for this purpose.

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Footnote

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