



Investigative algorithms for disorders affecting human plasma alkaline phosphatase: a narrative review

Michael Irving¹, Alexa R. Shipman², Kate E. Shipman^{3,4}

¹Biochemistry and Immunology Department, University Hospital Coventry and Warwickshire NHS Trust, Coventry, UK; ²Portsmouth Hospitals University NHS Trust, Portsmouth, UK; ³University Hospitals Sussex NHS Foundation Trust, Worthing, UK; ⁴Brighton and Sussex Medical School, Brighton, UK

Contributions: (I) Conception and design: All authors; (II) Administrative support: All authors; (III) Provision of study materials or patients: All authors; (IV) Collection and assembly of data: All authors; (V) Data analysis and interpretation: All authors; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

Correspondence to: Michael Irving, PhD. Biochemistry and Immunology Department, University Hospital Coventry and Warwickshire NHS Trust, Coventry CV2 2DX, UK. Email: Michael.irving@uhcw.nhs.uk.

Background and Objective: The following article is part of a special series to aid the physician in diagnosing the cause of various plasma abnormalities. Patients presenting with low or high alkaline phosphatase (ALP) activity can present a diagnostic challenge, particularly in the absence of symptoms. The objective is to provide information and algorithms to support the physician to order and interpret appropriate investigations when faced with this situation.

Methods: A narrative, focused literature review was performed of English language resources using PubMed, OMIM, ScienceDirect, and Google. References published from database inception to June 2023 were searched for from February 2023 to June 2023. Further articles were identified from reference lists.

Key Content and Findings: Bone and liver are the primary sources of ALP activity. When activity is low correlates with potassium, magnesium, and calcium concentrations which can help rule out specimen contamination and identify bone disease and malnutrition. When activity is high correlates with liver function tests, followed by a bone profile if normal. Analytical limitations include the range of isoforms present such as macro-ALP which can affect the results obtained.

Conclusions: Diagnostic algorithms are presented that should support healthcare professionals to efficiently and systematically approach people with these abnormalities.

Keywords: Alkaline phosphatase (ALP); low; high; diagnosis; algorithm

Received: 12 September 2023; Accepted: 04 December 2023; Published online: 10 January 2024.

doi: 10.21037/jlpm-23-63

View this article at: <https://dx.doi.org/10.21037/jlpm-23-63>

Introduction

The alkaline phosphatases (ALPs) are a group of isoenzymes that catalyse the hydrolysis of organic phosphate esters. Located primarily in the cytoplasm they are anchored to the plasma membrane by a glycosylphosphatidylinositol anchor in almost all tissues (1,2), with a half-life of 7 days (3). Optimal enzyme conditions include an alkaline pH, between 8–11 (4). The ALP metalloenzyme family is encoded by multiple genes in humans, is expressed in multiple tissues including liver and bone, and each enzyme requires three

metal ions, two Zn^{2+} and one Mg^{2+} , in its active site (5).

ALP is widely included in panels designed to assess liver and bone function with alanine aminotransaminase (ALT) and/or aspartate aminotransferase (AST), total protein, albumin, and bilirubin, or calcium, albumin with or without phosphate respectively (6). The International Federation of Clinical Chemistry (IFCC) stated adult range is 45–135 U/L (7). Diagnostic algorithms will be presented to provide an approach to investigate abnormalities of ALP activity. This article is not meant to replace current

Table 1 The search strategy summary

Items	Specification
Date of search	February 2023–June 2023
Databases and other sources searched	PubMed, OMIM, ScienceDirect, Google
Search terms used	“ALP”, “Alkaline Phosphatase”, “Hypophosphatasia”, “isoenzyme”, “pregnancy”
Timeframe	From database inception to June 2023
Inclusion criteria	All papers and reviews were included, restricted to English
Selection process	M.I. and K.E.S. conducted initial search, with refinement by all other authors to obtain consensus and agreement
Any additional considerations, if applicable	Seminal texts were also searched and the references of important articles and texts were obtained and checked for relevance

guidelines and reviews, nor replace thorough clinical assessment. Instead, these algorithms are aimed to enhance the understanding of the role of diagnostics in the clinical pathway. Quality and efficiency of patient care are promoted by the appropriate use of diagnostics. We present this article in accordance with the Narrative Review reporting checklist (available at <https://jlp.amegroups.org/article/view/10.21037/jlp-23-63/rc>).

Methods

The narrative literature review was undertaken with review of PubMed, OMIM, ScienceDirect, and Google and seminal texts (*Table 1*). The search was performed from February 2023 to June 2023 from database inception. Language was restricted to English. Diagnostic algorithms were created from synthesis of the information obtained from literature review.

Metabolic role of ALP

ALP is a zinc metalloenzyme; it is activated by magnesium, or other cations. Encoded by four distinct genes that encode the isoforms found in humans with a great many functions (*Table 2*) (14). The most abundant, *ALPL*, accounts for the tissue non-specific ALP (TNSALP) that is found in liver, bone, and kidney, encoded on chromosome 1. Post translational modification further results in distinct carbohydrate compositions between those produced in the liver (hepatocellular and biliary canalicular subtypes) and in bone osteoblasts (BALP) (5). *ALPP*, *ALPP2* and *ALPI* encode tissue specific ALP found in the placenta (syncytiotrophoblasts), germ cells, and intestines (enterocyte

luminal surface) respectively (5). Tumours have been associated with excretion of variations of the placental isoform e.g., Regan isoenzyme (15). Both liver and intestinal types are found in the brush border of the renal proximal tubule (*Table 2*). The three major substrates for ALP are inorganic pyrophosphate (PP_i), pyridoxal-5-phosphate (PLP), and phosphoethanolamine (PEA) (16).

Although ubiquitous there is a considerable difference in relative enzyme activity between tissues (*Table 3*) (8). Although activity may be higher in the kidney, bone is heavier therefore the highest ALP tissue activity in total is found in the placenta followed by the intestine, bone, kidney, and liver in descending order (2). In health, human serum contains predominantly bone and liver isoforms, with approximately equal activity from both. Elevations of ALP activity are seen in health, e.g., during pregnancy and placenta formation (placental isoform), growing (bone isoform, and see later section on special states) or can represent a wide range of disorders of tissues (particularly those with high ALP activity) including endocrine and medication causes (2).

Liver based ALP is synthesised in many tissues including hepatocytes and osteoblasts (*Figures 1,2*) (3,14). At least 90% (8) of the ALP is attached to exterior surfaces, 3% is found in the cytosol and the rest in the extracellular fluid and vessels. ALP is eliminated by being taken up by hepatocytes and catabolised in lysosomes (14) (*Figure 1*). In adults, there is a continuous process of bone remodelling, involving the resorption of bone by osteoclasts, followed by the synthesis and maintenance of bone by osteoblasts (*Figure 2*). This is a process that is regulated by complex interactions between large numbers of factors and hormones and is highly co-ordinated (18). ALP, no matter the source, is not cleared by the kidneys

Table 2 Table of human alkaline phosphatase isozymes (8-13)

Human genes	Names	Tissue distribution	Function
<i>ALPL</i>	Tissue nonspecific alkaline phosphatase	Developing nervous system	Hydrolyses pyrophosphate supplying inorganic phosphate for mineralization, reduces extracellular pyrophosphate and phosphorylcholine concentration, dephosphorylation and detoxification of lipopolysaccharides, sphingosine 1-phosphate receptor signalling, antiendotoxin mediator and anti-inflammatory, regulation of adenosine concentrations
	Liver-bone-kidney type alkaline phosphatase	Skeletal tissue, liver, kidney	Hydrolyses a variable spectrum of phosphate-containing compounds, contributes to DNA synthesis, attenuates inflammation, influences mitochondrial respiration, extracellular matrix mineralization
<i>ALPP</i>	Placental alkaline phosphatase	Syncytio-trophoblast, tumours	Indicative of tissue having stem cell functions, tumour marker, detoxification of bacterial endotoxin
<i>ALPP2</i>	Germ cell alkaline phosphatase	Testis, malignant trophoblast, testicular cancer	Indicative of tissue having stem cell functions, sperm glycolytic reactions and fructose formation, tumour marker to diagnose carcinoma- <i>in situ</i> of the testis, seminoma
<i>ALPI</i>	Intestinal alkaline phosphatase	Gut	Intestinal tight junction integrity and maintains barrier function, attenuates inflammation, regulation of intestinal surface pH, absorption of lipids, detoxification of free nucleotides and bacterial lipopolysaccharides, possible modulation of the gut microbiota, regulation of transmucosal passage of bacteria, dephosphorylation of extracellular adenosine triphosphate

Table 3 Relative tissue activity of alkaline phosphatase in human tissue (2,8,17)

Tissue	Activity per g of wet tissue with two substrates, liver activity set to 100 as benchmark		IU/g of tissue, mean \pm standard deviation
	B-glycerophosphate	Phenylphosphate	
Placenta	–	3,214	69 \pm 44
Ileum (mucosa)	1,714	2,524	38 \pm 14
Kidney	619	–	2.1 \pm 0.7
Bone	–	571	Age dependent
Colon (mucosa)	471	–	2.3 \pm 0.8
Adrenal	167	–	
Lung	129	–	2.1 \pm 0.5
Spleen	129	–	
Liver	100	100	2.6 \pm 1.4
Brain	76	–	
Stomach (mucosa)	62	–	
Heart	–	33	
Pancreas	–	10	
Skeletal muscle	–	5	
Serum (adult)	0.2	–	
Testes	–	–	0.5 \pm 0.1

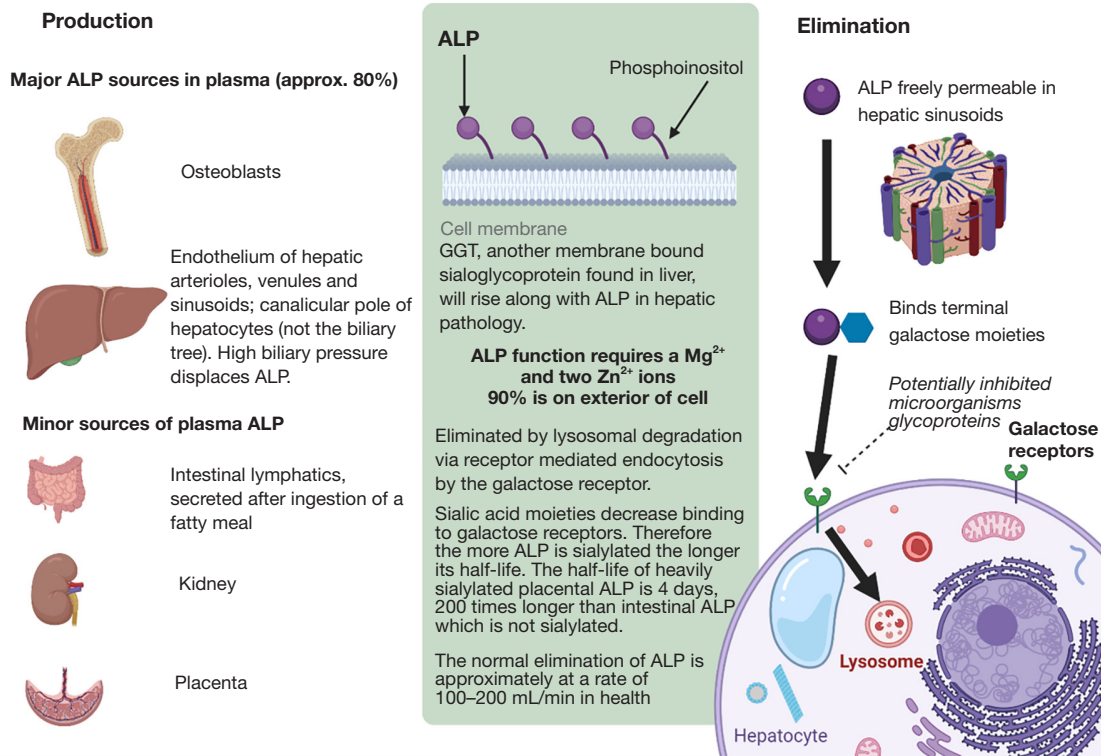


Figure 1 Summary of ALP origin, production, and elimination in the body. ALP, alkaline phosphatase; GGT, gamma glutamyl transferase; Mg, magnesium; Zn, Zinc.

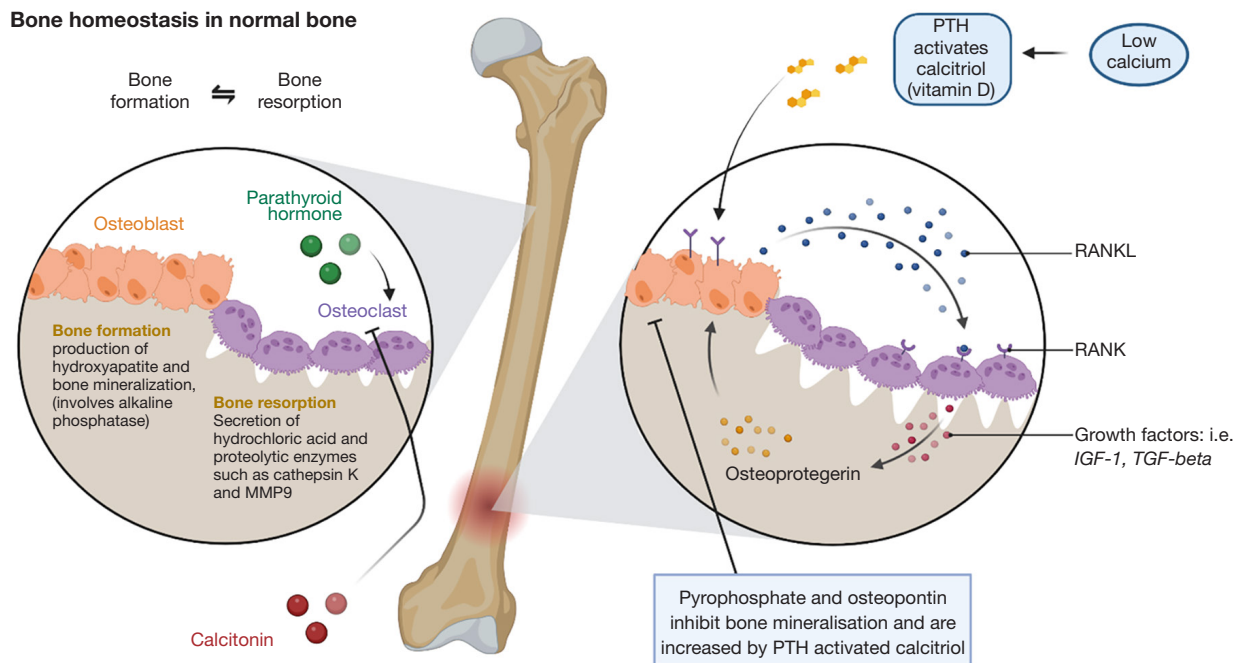


Figure 2 Bone homeostasis in normal bone. PTH, parathyroid hormone; RANKL, receptor activator of nuclear factor kappa-B ligand; RANK, receptor activator of nuclear factor kappa-B; IGF-1, insulin-like growth factor 1; TGF-beta, transforming growth factor beta.

making it a useful tool for assessment of bone turnover in the presence of chronic kidney disease (CKD) (19).

In CKD, abnormalities of mineral and bone metabolism occur with extra-skeletal calcification, called CKD-mineral bone disorder (CKD-MBD), which is distinct from the morphological diagnosis of renal osteodystrophy which is a consequence of it (20,21). Fibroblast growth factor 23 (FGF23) is raised prior to the more characteristic findings of hyperphosphataemia, high parathyroid hormone (PTH) or low activated vitamin D (1,25VitD) and is likely a key factor for renal, bone, mineral, and cardiovascular complications (22). Although FGF23 is phosphaturic, in CKD the renal response is diminished resulting in hyperphosphataemia which stimulates PTH secretion with a subsequent increase in bone turnover and measurable ALP activity. Furthermore, the impaired urinary tubular function and elevated serum FGF23 concentration lead to decreased vitamin D activation and enhanced PTH secretion under conditions of relative calcium deficiency, thereby giving rise to secondary hyperparathyroidism. BALP may be slightly superior in diagnosing bone disease in CKD-MBD but not enough to justify the additional cost for routine practice (21).

Measurement of ALP

ALP activity is dependent on zinc and activated by magnesium thereby order of blood draw is important to prevent cation chelation, and hence reduction in measured activity, by common tube additives, such as ethylenediaminetetraacetic acid (EDTA), oxalate, or citrate (23). Delayed separation of serum from the cells may very marginally increase ALP activity but analysis within 24 hours is unlikely to affect results in a clinically significant way irrespective if stored at room temperature or refrigerated (24,25).

The activity of ALP is measured by the change of absorbance at 405 nm by the yellow quinoid version of 4-nitrophenoxide formed at an alkaline pH following ALP catalysing the cleavage of phosphate from 4-nitrophenyl phosphate. The rate can be increased by including a phosphate acceptor e.g., adenosine monophosphate, which is also included in the IFCC reference method based on the above reaction (26).

Isoforms can be detected through a variety of means e.g., electrophoresis (27), differential deactivation e.g., by heat (1,28), differential response to inhibitors (1,29), affinity for lectins (30,31) and immunoassay (32,33). However, in clinical practice other routinely available tests are mostly reliable at identifying the source (as well as the marked

improvement in imaging technology and availability) and the time, and cost, it may take to get these specialist tests makes them almost obsolete (34).

There are several caveats to consider when interpreting ALP results:

- ❖ Race and sex can impact ALP activity: positive correlation with increases in mean body mass index (BMI) of populations, seen in those of Hispanic descent greater than those of African American, which were greater than those of Caucasian. Males have higher activities than females within populations, which is also positively correlated to BMI (35).
- ❖ People who smoke have activity 10% higher than those who do not (36).
- ❖ Activity fluctuates approximately 6% from week to week in a healthy individual (37).

ALP can become bound to immunoglobulin, called macro-ALP, which prevents it being cleared as quickly (38,39). This is analytically correct, i.e., there is an increase in ALP activity in the serum, but does not represent increased tissue turnover and therefore a potential cause of spurious results.

Low ALP activity

In a study of unselected male patients, the causes of a low ALP, which was rare, occurring in only 0.2% of almost 70,000 samples, were:

- ❖ Cardiac surgery and cardiopulmonary bypass (26.5%), mean pre-surgical ALP was 71 U/L which fell to 20 U/L, corresponding magnesium concentration fell from a mean of 0.98 to 0.54 mmol/L.
- ❖ Malnutrition (12.0%), mean ALP of 18 U/L, secondary to decreased activity of both bone and hepatic ALP.
- ❖ Severe magnesium deficiency (mean concentration 0.48 mmol/L) affected 4.8% with a mean ALP activity of 21 U/L.
- ❖ Hypothyroidism (2.4%), ALP activity returning to normal once euthyroid.
- ❖ Severe anaemia attributed to iron deficiency (1.2%) (40).

In an audit of a year's cases another group identified blood transfusion, cardiopulmonary bypass, and chemotherapy as causes, but a few cases had no identifiable cause (41). This led the team to conclude that the lower limit of ALP activity is too arbitrary to be useful to pick up important pathology. The causes of low ALP activity will be discussed below with *Figure 3* providing a diagnostic algorithm aimed at helping

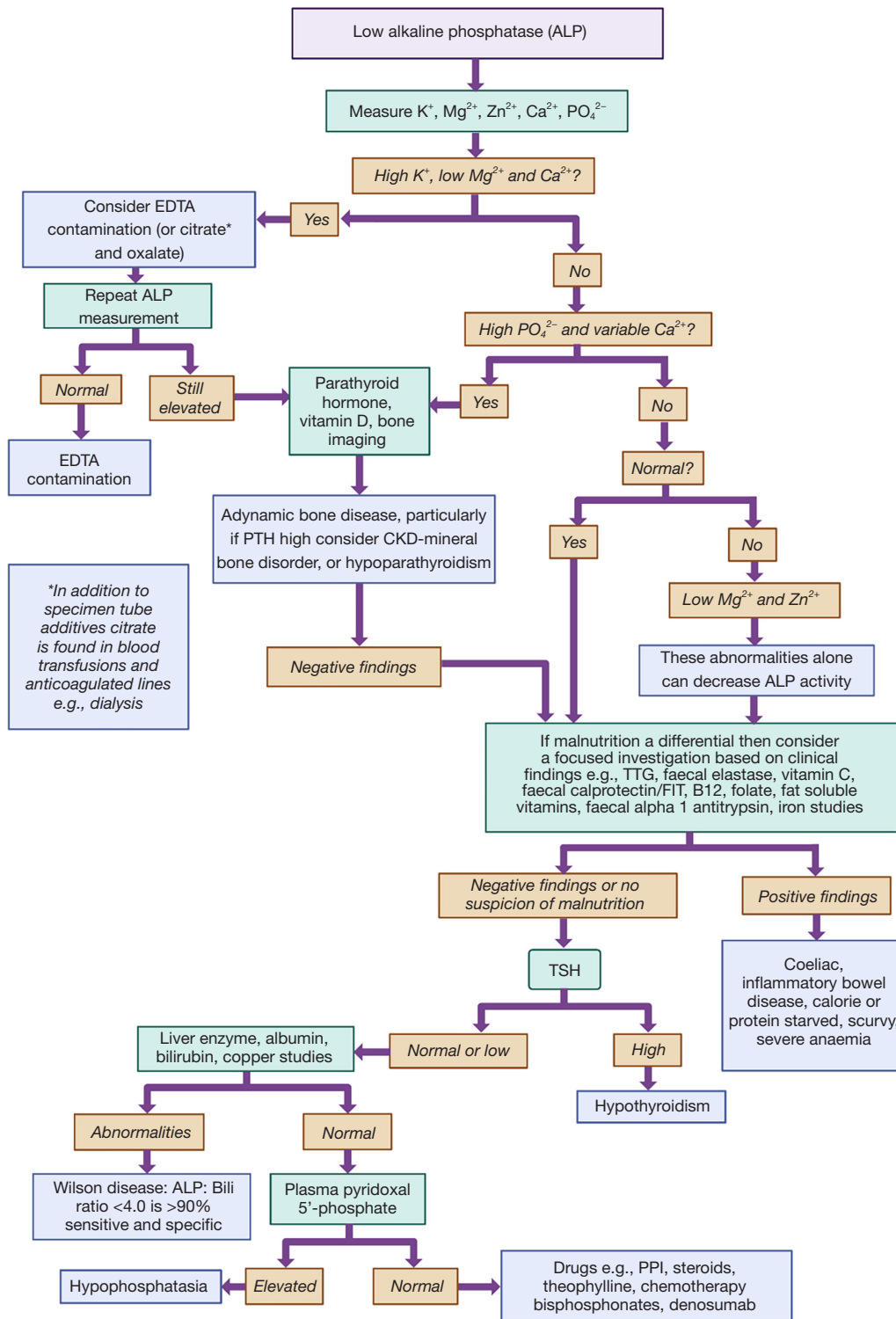


Figure 3 Algorithm and supporting information for the laboratory investigation of low ALP. ALP, alkaline phosphatase; K, potassium; Mg, magnesium; Ca, calcium; PO, phosphate; EDTA, ethylenediaminetetraacetic acid; PTH, parathyroid hormone; CKD, chronic kidney disease; TTG, tissue transglutaminase; FIT, faecal immunochemical test; B12, vitamin B12; TSH, thyroid stimulating hormone; PPI, protein pump inhibitor.

the clinician approach the investigation.

Pseudo hypophosphatasia

For an unexpected isolated low ALP activity EDTA, or other tube additives such as oxalate or citrate, contamination should be considered. Serum ALP activity decreased significantly at EDTA concentrations of >1.86 mmol/L (unexpected hyperkalaemia and hypocalcaemia are further clues) but this degree of contamination is not commonly reached in most cases of contamination (42,43). EDTA, citrate and oxalate bind cations and hence reduce ALP activity which requires both zinc and magnesium ions. Citrate is also found in blood transfusions and anticoagulated lines, e.g., in haemodialysis.

Primary hypophosphatasia

Primary low ALP activity, called hypophosphatasia, is due to genetic mutations in the *ALPL* gene (TNSALP) and clinical symptoms can include rickets and osteomalacia, epilepsy, myopathy, respiratory difficulties, hypercalcaemia, nephrocalcinosis, and tooth loss (16). At its most severe it can be fatal in infancy, but others will present as adults (16). Despite the wide phenotype all will have dental and skeletal mineralization symptoms, with high PPi in the bone matrix (16). Deformed and painful bones in infancy or painless, early, tooth loss (with roots attached) may indicate hypophosphatasia (16). ALP deactivates active B6, PLP, to form pyridoxal, and the accumulation of pyridoxal may account for the seizures seen in some babies with hypophosphatasia, occurring a few days after birth. Although not demonstrated in humans, restoring ALP activity in deficient mice resolves the seizures (16). High urinary PEA is another useful screening tool however genetic diagnostics (more than 300 mutations have been described in the *ALPL* gene but not all are disease causing) and enzyme replacement therapy is now available (16). Hyperphosphataemia can be seen, and later hypercalcaemia, as the phosphate and calcium homeostasis mechanisms are normal promoting renal excretion which can lead to renal complications such as renal failure and nephrocalcinosis (16).

Malnutrition and malabsorption

Malnutrition can lead to low ALP activity. This may be caused by metal deficiencies such as zinc or magnesium (44,45). In one study all those with a low ALP, compared

to controls, were deficient in either zinc (47.6%) or magnesium (52.4%) (46). Nutritional rickets can reduce ALP activity, however if vitamin D deficient with low dietary calcium intake (47), ALP may be elevated (although not reliably) (48-50). A low protein state may lead to a low ALP activity (51,52) but again this observation is not consistent in kwashiorkor (53).

Vitamins C and B12 have been shown to promote bone growth, and deficiencies have a negative impact on bone growth (54) and increase the rate of osteoporosis (55). ALP is a marker for bone turnover therefore any decrease in turnover will affect the activity of ALP (55). However, it is unusual to find isolated nutritional deficiencies with, for example, a case report of scurvy demonstrating elevated ALP due to hypovitaminosis D, and coeliac disease which is often associated with elevated ALP activity (56,57).

Endocrine and metabolic links to ALP

Wilson disease, an inborn error of copper metabolism, has been associated with low ALP activity, particularly in the initial stages (58). It is believed that copper competition with zinc causes the suppression in ALP activity (25). Conditions associated with low bone turnover, such as hypoparathyroidism and hypothyroidism, may reduce ALP activity, likely not enough to cause low ALP activity commonly (16,59). In cardiopulmonary bypass ALP is presumed to be consumed by dephosphorylating inflammatory chemicals and the reduction in activity correlates with worsening outcomes (60).

Drugs and toxins

Clofibrate reduces ALP activity but is no longer used (61,62), having been replaced by other drugs in the fibrate class due to side effects. ALP activity is inhibited by theophylline, sulphonamides, arsenates, molybdates, and other agents, with active development of ALP inhibitors underway to attempt to treat ectopic calcification which are not yet on the market (8,63-65). Inhibition by cation chelation by citrate can cause a transiently low ALP, e.g., in neonates or massive transfusions (41). ALP production is reduced by bisphosphonates (66,67), denosumab (68), and proton pump inhibitors (69) although they may not result in ALP activity beneath the reference range (70) (see *Table 4*).

High ALP

The commonest tissue origins of an elevated ALP activity are liver and bone (see *Table 5*). The key differentials will

Table 4 Drugs that can lower measured ALP activity in humans (8,71,72)

Drugs	Mechanism
Proton pump inhibitors	Inhibit osteoblasts, decreases bone turnover
Steroids	Effects variable on bone turnover and can also raise ALP
Theophylline, aminophylline	Inhibits ALP activity and so test is falsely low
Bisphosphonates	Suppress osteoclast and so negative feedback to osteoblast
Denosumab	Suppress osteoclast and so negative feedback to osteoblast
Sulfonamides	Inhibits ALP activity and so test is falsely low
Cimetidine and ranitidine	Inhibits ALP activity and so test is falsely low
Imidazole and levamisole	Inhibits ALP activity and so test is falsely low
Nitrofurantoin, cyanides, arsenals	Inhibits ALP activity and so test is falsely low

ALP, alkaline phosphatase.

Table 5 Relative tissue activity of alkaline phosphatase in human pathology (2,73-77)

Disorder	Patients with abnormal ALP (%)	Mean ALP (multiple of upper reference limit)
Primary liver cancer	92	5.5
Tumour, metastatic liver	88	5.5
Extrahepatic obstruction	94	4.9
Intrahepatic obstruction	82	2.8
Acute viral hepatitis	80	2.5
Inactive cirrhosis	75	2.1
Alcoholic hepatitis	77	1.8
Chronic active hepatitis	78	1.7
Primary biliary cholangitis	>95	1.67 [†]
Osseous metastases	74	2
Pregnancy	100	Depends on stage, up to 4
Osteomalacia	80	1.5
Low vitamin D	0	–
Hyperthyroidism	44	Up to 5
Secondary hyperparathyroidism	75	–

[†], wide range of values, this figure, and above, is the trigger for treatment with ursodeoxycholic acid. ALP, alkaline phosphatase.

differ depending on how well the person is and, potentially, the degree of elevation (*Table 5*). In hospitalised patients ALP is commonly raised [causes include pyelonephritis, malignancy, congestive heart failure and renal failure (14,78,79)]; in those with underlying conditions (who are well) elevations tend to resolve within 3 months (14). The causes of a rise in ALP activity are discussed below. A

diagnostic algorithm is provided as a systematic framework that can be used to guide rather than be followed proscriptively, see *Figure 4* (80,81). It is important to note in mild elevations of ALP activity other laboratory tests may not be helpful in identifying the cause, particularly in asymptomatic patients, and there may need to be a strategy of watchful waiting to determine if the elevation

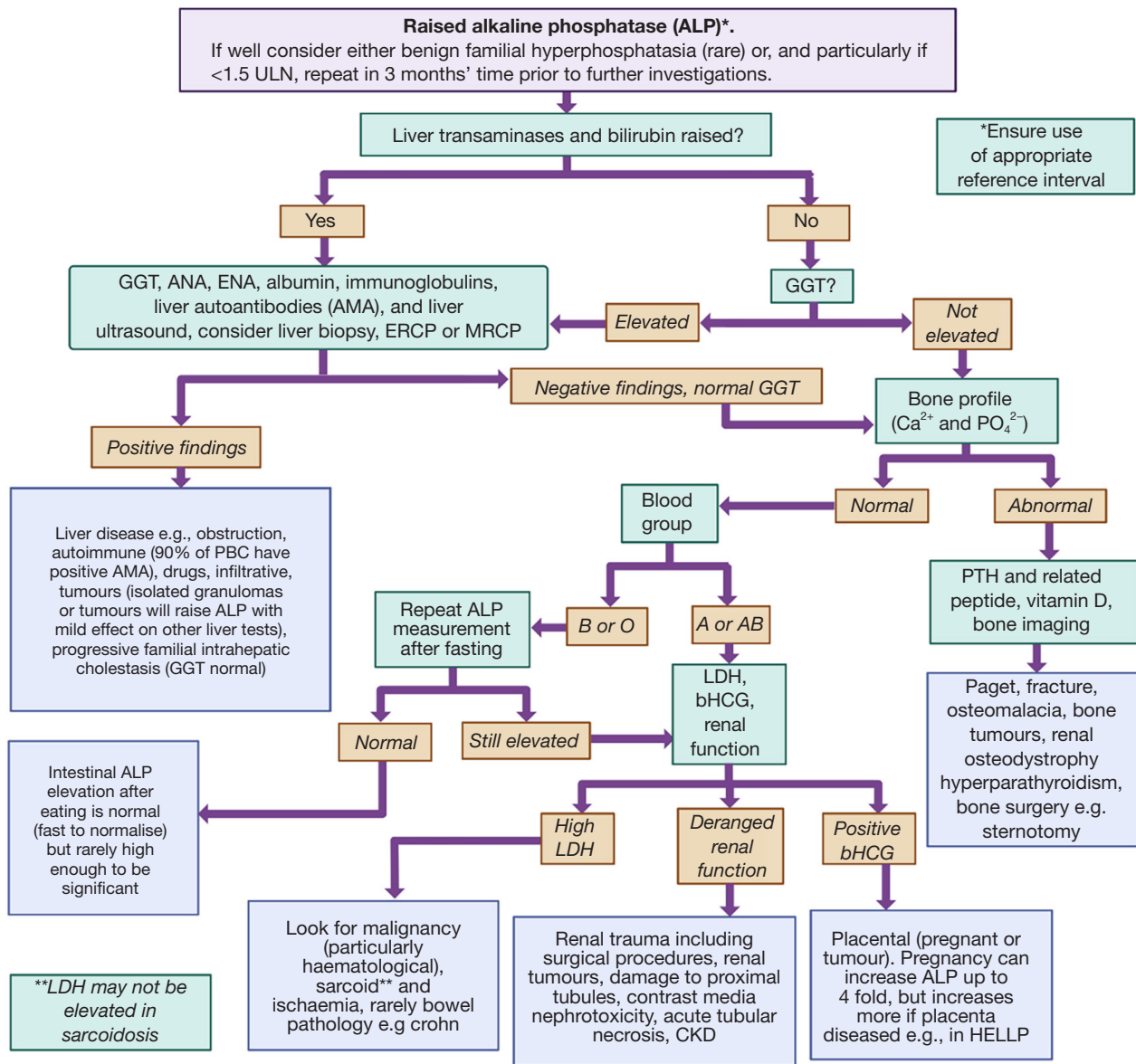


Figure 4 Algorithm and supporting information for the laboratory investigation of raised ALP. ALP, alkaline phosphatase; ULN, upper limit of normal; GGT, gamma-glutamyl transferase; ANA, anti-nuclear antibody; ENA, extractable nuclear antigen antibody; AMA, anti-mitochondrial antibody; Ca, calcium; PO, phosphate; ERCP, endoscopic retrograde cholangiopancreatography; MRCP, endoscopic retrograde cholangiopancreatography; PBC, primary biliary cholangitis; PTH, parathyroid hormone; LDH, lactate dehydrogenase; bHCG, beta human chorionic gonadotrophin; CKD, chronic kidney disease; HELLP, HELLP syndrome.

is persistent and the organ source [if less than 1.5 times the upper limit of normal (ULN) for example consider repeating at 3 months if isolated and patient is well] (82).

ALP and its association with the liver and biliary tract

Most test panels for liver function include the enzymes

ALT and ALP. A larger rise in ALT activity (compared with the ULN) may indicate hepatocellular damage whereas a relatively more significant ALP elevation (compared with the ULN) likely indicates a cholestatic pathology. Gamma-glutamyl transferase (GGT) elevation supports the diagnosis of a liver aetiology (GGT should be normal in bone causes of high ALP, unless there is

more than one pathology) (83-85). An exception is the rare familial intrahepatic cholestasis which has raised ALP and normal GGT activities (86). This can present in a benign form that occurs at any age, and lasts for several weeks to months, or a progressive form that causes severe cholestasis before 6 months of age and progressing to cirrhosis, liver failure and death, unless a liver transplant is provided (86).

When reviewing raised ALP results, an elevation of ALP of approximately four times the ULN or greater occurs in up to 75% of the patients with cholestasis, either intrahepatic or extrahepatic in one study (87). Liver diseases that principally effect parenchymal cells, such as infectious hepatitis, typically show only moderately elevated or even normal ALP activity but this depends upon study (see *Table 5*). As per the European Association for the Study of the Liver (EASL), activity thresholds for serum requiring diagnostic work-up are >1.5 times ULN (88).

This review will concentrate on the cholestatic liver diseases as the hepatocellular diseases will be covered in more detail in the companion article to this (on investigation of elevated transaminases). Cholestatic liver diseases include autoimmune e.g., primary biliary cholangitis or cirrhosis (PBC) (89). PBC is diagnosed by the presence of anti-mitochondrial antibodies (AMAs) (90) and increased concentrations of immunoglobulins [mainly immunoglobulin M (IgM)] (91). Anti-nuclear antibodies (ANAs) and anti-smooth muscle antibodies (SMAs) are found in nearly 50% of PBC patients (92). ANA, anti-glycoprotein 210 and/or anti-sp100 (nuclear membrane proteins) may be present in those who are AMAs negative (93).

Primary sclerosing cholangitis (PSC) and secondary sclerosing cholangitis are differentials for PBC (94,95). Up to 80% of people with PSC are diagnosed with inflammatory bowel disease (IBD), primarily ulcerative colitis (88). In PSC immunoglobulins are often elevated with IgM (96) and IgG raised in 45% and 61% respectively, with IgG typically exceeding 1.5 times the ULN (97). Immunoglobulin G Subclass 4 (IgG4) was found to be elevated in 9% of PSC patients in another study and has been suggested to be a disease subtype (98,99), with those with raised IgG4 tending to progress more rapidly without treatment (98,100). Other autoantibodies present include perinuclear antineutrophil cytoplasmic antibodies (pANCA) (26-94%), ANA (8-77%), and SMAs (0-83%) (101).

Imaging is relevant for all patients in whom cholestasis is suspected, cholangiography, preferentially with non-invasive magnetic resonance imaging (MRI), or endoscopically,

to exclude PSC. Transient elastography is another non-invasive tool that has shown high degree of accuracy in diagnosing advanced fibrosis in patients with PBC (102). A liver biopsy is rarely required.

Obstruction of the biliary tree, or cholestasis, will lead to an elevation of the ALP and causes for this include gallstone, biliary strictures and tumours, infiltrative processes, and medication (see *Table 6*) (82,108-111). For example, colestipol, a bile acid sequestrant, increases ALP activity but is not widely used anymore (61).

Bone

Increasing bone turnover, particularly osteoblast activity, will elevate ALP (5). Other tests including vitamin D, calcium, phosphate and PTH may be required to distinguish some of the following disease entities, and this is further illustrated in *Figure 4*. There are other, less commonly requested tests, which might also be available for the clinician to distinguish bone diseases (see sister algorithms in this special series on calcium and phosphate).

Common causes of bone pathology that increases ALP are periods of increased skeletal growth, such as during adolescence (4), fracture recovery (112), cancer including osteosarcoma or bone metastasis, hyperparathyroidism, CKD and vascular calcification (113).

Rickets, a rare condition in the UK, is the clinical consequence of impaired mineralization of the growth plate cartilage and spongiosa in the metaphysis of children and adolescents (114) due to calcium and/or vitamin D deficiency (calcipenic rickets), renal tubule dysfunction and disturbances of chondrocytes and osteoblasts (*Table 7*). Bones are resorbed to release calcium, under PTH action, which also results in hypophosphataemia (116,117). Osteomalacia is rickets occurring after the growth plates fuse, i.e., the adult version (114), and biochemically there will be raised PTH with deficiencies in phosphate and vitamin D. In osteomalacia vitamin D supplementation will return ALP activity to normal after about 6 months of treatment, therefore there is no need to remeasure activity prior to this time point (34).

Paget disease is an increasingly rare condition of disordered bone turnover with no clear single cause (118,119). Markers of bone turnover are raised including type I procollagen N-terminal peptide (P1NP), N-terminal telopeptide of type I collagen and bone ALP (120), with ALP activity used to measure treatment efficacy (121).

Table 6 Drugs that can cause an elevation in measured alkaline phosphatase activity in humans—excluding hepatotoxic medications (82,103-107)

Drugs	Mechanism
Antibiotics	
Penicillin derivatives	Intrahepatic cholestasis
Erythromycin	Intrahepatic cholestasis
Aminoglycosides	Enzyme induction
Sulfa drugs	Intrahepatic cholestasis
Antiepileptic drugs	
Carbamazepine	Intrahepatic cholestasis
Phenobarbital	Enzyme induction
Phenytoin	Enzyme induction
Sodium valproate	Enzyme induction
Antihistamines	
Cetirizine	Intrahepatic cholestasis
Cardiovascular drugs	
Captopril	Intrahepatic cholestasis
Diltiazem	Enzyme induction
Felodipine	Enzyme induction
Verapamil	Intrahepatic cholestasis
Disease modifying agents	
Penicillamine	Intrahepatic cholestasis
Sulfa drugs	Intrahepatic cholestasis
Allopurinol	Causes a granulomatous hepatitis
Polycyclic aromatic hydrocarbons	
Oral contraceptive pill (oestrogen)	Enzyme induction
Anabolic and corticosteroids	Enzyme induction but variable and can lead to low ALP
Psychotropic drugs	
Monoamine oxidase inhibitors	Intrahepatic cholestasis
Phenothiazines and chlorpromazine	Intrahepatic cholestasis
Lipid lowering	
Statins	Enzyme induction

ALP, alkaline phosphatase.

However, there is an argument that ALP normalization is an inappropriate measure of treatment success as only occurred in ~25% of those treated with bisphosphonates, though ALP activity overall did reduce by ~41% (122). Treating to normalise ALP has no reported differences in fracture rate, pain relief, hearing, need for orthopaedic surgery or quality of life when compared to treating to alleviate bone pain solely (123,124). Imaging is used to confirm a diagnosis of Paget disease (125).

Osteoporosis is not associated with an elevation of ALP, unless there is a secondary fracture, however BALP can be used to monitor the effect of treatment on bone turnover (126). Osteomyelitis does not raise ALP either. A femoral neck fracture elevates serum ALP activity by approximately 30% and trochanteric by 100% (14,127).

Note that oestrogens reduce BALP in post-menopausal women which might theoretically disguise subtle elevations in women on hormone replacement therapy (128). Steroids have an unpredictable effect on BALP as although they may be osteogenic in exogenous or endogenous hypercortisolism there may be no noticeable effects or mild suppression (66,129,130). Hyperthyroidism and thyrotoxicosis cause an elevation in ALP, likely by effects on bone metabolism (131,132).

Tumour placental ALP

In cancer there can be significant production of ALP which cannot be related to the tissue involvement and therefore represents paraneoplastic production of foetal proteins (particularly placental type ALP) from the tumour. Regan isoform, a rare variant of placental ALP, is one example; in one series (133) of 239 people with malignant disease 25.5% had elevated Regan isoenzyme detectable in the serum. Tumour types reported to demonstrate elevations of Regan isoform include kidney, stomach, uterine, lung, cervical, endometrial, testicular, ovarian, medullary thyroid, haematopoietic, prostate and germ cell (15). Other examples include Kasahara a foetal intestinal ALP isoform (134) e.g., in renal cell carcinoma (135), and Nagao a placental-like ALP isoform (134,136).

Miscellaneous

Transient hyperphosphataemia (TH) is a benign condition, of unknown aetiology, characterized by marked elevation of serum ALP activity, such as an increase of four-fold ULN. There should be a notable absence of associated diseases such as liver, bone, or kidney pathology and it

Table 7 Typical biochemical features of various forms of osteomalacia/rickets (115)

Cause	Calcium	Vitamin D	PTH	Phosphate	Other
Hypovitaminosis D	L	L	H	L	H ALP
Low 1,25D (renal failure, vitamin D dependent rickets type I caused by loss of function in 1 α hydroxylase)	L	N	H	L/H	H ALP, L 1,25D
Vitamin D dependent rickets type II (loss of function of vitamin D receptor)	L	N	H	L	H ALP, H 1,25D
Low phosphate (multiple causes see later slide)	N	N	N	L	H ALP

PTH, parathyroid hormone; L, low; H, high; N, normal; ALP, alkaline phosphatase.

settles within weeks or months of initial observation (137). However, ALP activity can remain elevated for extended periods of time, for greater than 4 months, in approximately 20% of cases (138). Diagnosis of TH is linked to the 'fast' α_2 band which is detected on agarose gel electrophoresis (139) but testing may not be necessary if the person is asymptomatic. Prevalence of TH is suggested to be around 2.8% (ALP >1,000 U/L) (140) in the classically affected population, children younger than 5 years old, but has also been described in transplant patients (141,142).

Bacterial and fungal glycoproteins can compete with the receptors involved in the excretion and elimination of ALP from the serum (14). Therefore, ALP elevations can be seen in infections without any obvious liver toxicity or cholestasis (143).

Raised ALP has also been identified in a range of other disorders including rheumatoid arthritis (144,145), atherosclerosis (146), and axial spondylarthritis (147). However, the links between elevated ALP and the diseases have not been fully established, or the mechanisms understood.

In individuals with blood groups O and B, ALP concentration increases by about 20% after consuming a fatty meal, due to contribution from the intestinal tract isoenzyme (14,148). It was found that red cells of blood group A bind almost all intestinal ALP (149), which is not replicated in those with types O or B. As this elevation can persist for up to 14 hours in the serum, the recommendation is to check the serum enzyme activity in a fasting state (150). In one case a patient's intestinal ALP activity was measured as high as 140 U/L, with a normal range suggested to be <18 U/L (151). There are also cases of benign familial conditions causing elevated intestinal ALP. Rosalki *et al.* presents examples of patients with persistent and unexplained raised ALP, above the reference range, that had a genetic component, with an autosomal dominant pattern

of inheritance suggested as the cause (152).

Special states

Serum ALP varies with age and gender. There are two peaks of ALP activity during infancy and puberty, which fall mid-childhood and towards the end of adolescence respectively (153,154). A German study of over 300,000 paediatric plasma samples demonstrated that at birth ALP activity is low. Activity increases quickly, peaking at 20 days and then decreasing again until 4 years of age with an increase during adolescence to reach adult ranges (155). For a reference source for paediatric values the CALIPER database is a useful source (156). Gender will also influence these peaks, with females shown to peak 2 years earlier than males in keeping with growth rates (112,138).

Gender also influences expected BALP activity in later life, significantly higher activities were found in postmenopausal women when compared to premenopausal (157). During pregnancy, bone turnover increases (158) contributing to an increase in BALP plus an additional increase in placental ALP activity in serum resulting in an ALP activity threefold higher at the end of term (159). Therefore, ALP activity should be reviewed against appropriate reference ranges to avoid over diagnosing or missing pathology.

Conclusions

Serum ALP activity can be affected by normal physiological states and diseases. Pathological causes of high activity commonly are attributable to bone or liver disorders. Unexpectedly low activity may be due to pre-analytical contamination with cation chelators. A set of diagnostic algorithms have been created to guide the reader's approach and provide a systematic route of testing. Each patient is unique however and the clinical picture may direct the

reader to skip steps or refer to other algorithms within the series or in the literature. These algorithms are not a replacement for experience, expert opinion or local guidelines and should instead act as a diagnostic aid.

Acknowledgments

All figures were created with BioRender.com.

Funding: None.

Footnote

Provenance and Peer Review: This article was commissioned by the editorial office, *Journal of Laboratory and Precision Medicine* for the series “Investigative Algorithms in Laboratory Medicine II: Focus on Bone and Liver”. The article has undergone external peer review.

Reporting Checklist: The authors have completed the Narrative Review reporting checklist. Available at <https://jlp.m.amegroups.org/article/view/10.21037/jlpm-23-63/rc>

Peer Review File: Available at <https://jlp.m.amegroups.org/article/view/10.21037/jlpm-23-63/prf>

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <https://jlp.m.amegroups.org/article/view/10.21037/jlpm-23-63/coif>). The series “Investigative Algorithms in Laboratory Medicine II: Focus on Bone and Liver” was commissioned by the editorial office without any funding or sponsorship. K.E.S. served as the unpaid Guest Editor of the series and serves as an unpaid editorial board member of the *Journal of Laboratory and Precision Medicine* from September 2022 to August 2024. The authors have no other conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the

original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

References

- Sharma U, Pal D, Prasad R. Alkaline phosphatase: an overview. *Indian J Clin Biochem* 2014;29:269-78.
- Lott JA, Wolf PL, Nemesánszky E, et al. Clinical enzymology: a case-oriented approach. 1986:285.
- Lowe D, Sanvictores T, Zubair M, et al. Alkaline Phosphatase. Treasure Island, FL, USA: StatPearls Publishing; 2023.
- Golub EE, Boesze-Battaglia K. The role of alkaline phosphatase in mineralization. *Curr Opin Orthop* 2007;18:444-8.
- Makris K, Mousa C, Cavalier E. Alkaline Phosphatases: Biochemistry, Functions, and Measurement. *Calcif Tissue Int* 2023;112:233-42.
- Siller AF, Whyte MP. Alkaline Phosphatase: Discovery and Naming of Our Favorite Enzyme. *J Bone Miner Res* 2018;33:362-4.
- International Confederation of Clinical Chemistry. [cited 2023 Nov 7]. Available online: <https://grid.ifcc.org/>
- Le-Vinh B, Akkuş-Dağdeviren ZB, Le NMN, et al. Alkaline Phosphatase: A Reliable Endogenous Partner for Drug Delivery and Diagnostics. *Advanced Therapeutics* 2022;5:2100219.
- Millán JL. Alkaline Phosphatases: Structure, substrate specificity and functional relatedness to other members of a large superfamily of enzymes. *Purinergic Signal* 2006;2:335-41.
- Vimalraj S. Alkaline phosphatase: Structure, expression and its function in bone mineralization. *Gene* 2020;754:144855.
- Lallès JP. Recent advances in intestinal alkaline phosphatase, inflammation, and nutrition. *Nutr Rev* 2019;77:710-24.
- Zaher DM, El-Gamal MI, Omar HA, et al. Recent advances with alkaline phosphatase isoenzymes and their inhibitors. *Arch Pharm (Weinheim)* 2020;353:e2000011.
- Zhang Z, Nam HK, Crouch S, et al. Tissue Nonspecific Alkaline Phosphatase Function in Bone and Muscle Progenitor Cells: Control of Mitochondrial Respiration and ATP Production. *Int J Mol Sci* 2021;22:1140.
- Levitt MD, Hapak SM, Levitt DG. Alkaline Phosphatase Pathophysiology with Emphasis on the Seldom-Discussed Role of Defective Elimination in Unexplained Elevations

- of Serum ALP - A Case Report and Literature Review. *Clin Exp Gastroenterol* 2022;15:41-9.
15. Bukowczan J, Pattman S, Jenkinson F, et al. Regan isoenzyme of alkaline phosphatase as a tumour marker for renal cell carcinoma. *Ann Clin Biochem* 2014;51:611-4.
 16. Linglart A, Bioso-Duplan M. Hypophosphatasia. *Curr Osteoporos Rep* 2016;14:95-105.
 17. McComb RB, Bowers GN, Posen S. *Alkaline Phosphatase*. Boston, MA, USA: Springer US; 1979.
 18. Burch J, Rice S, Yang H, et al. Systematic review of the use of bone turnover markers for monitoring the response to osteoporosis treatment: the secondary prevention of fractures, and primary prevention of fractures in high-risk groups. *Health Technol Assess* 2014;18:1-180.
 19. Sprague SM, Bellorin-Font E, Jorgetti V, et al. Diagnostic Accuracy of Bone Turnover Markers and Bone Histology in Patients With CKD Treated by Dialysis. *Am J Kidney Dis* 2016;67:559-66.
 20. Lewis R. Mineral and bone disorders in chronic kidney disease: new insights into mechanism and management. *Ann Clin Biochem* 2012;49:432-40.
 21. Moe S, Drüeke T, Cunningham J, et al. Definition, evaluation, and classification of renal osteodystrophy: a position statement from Kidney Disease: Improving Global Outcomes (KDIGO). *Kidney Int* 2006;69:1945-53.
 22. Vervloet MG. Shedding Light on the Complex Regulation of FGF23. *Metabolites* 2022;12:401.
 23. Banfi G, Salvagno GL, Lippi G. The role of ethylenediamine tetraacetic acid (EDTA) as in vitro anticoagulant for diagnostic purposes. *Clin Chem Lab Med* 2007;45:565-76.
 24. Selvakumar C, Madhubala V. Effect of sample storage and time delay (delayed processing) on analysis of common clinical biochemical parameters. *Int J Clin Biochem Res* 2017;4:295-8.
 25. Marshall W. *Alkaline phosphatase (serum, plasma)*. Analyte Monograph. 2013.
 26. Schumann G, Klauke R, Canalias F, et al. IFCC primary reference procedures for the measurement of catalytic activity concentrations of enzymes at 37 °C. Part 9: reference procedure for the measurement of catalytic concentration of alkaline phosphatase International Federation of Clinical Chemistry and Laboratory Medicine (IFCC) Scientific Division, Committee on Reference Systems of Enzymes (C-RSE) (1)). *Clin Chem Lab Med* 2011;49:1439-46.
 27. Epstein E, Kiechle FL, Zak B. Use of alkaline phosphatase isoenzyme analysis in the evaluation of cholestatic liver disease. *Ann Clin Lab Sci* 1984;14:292-7.
 28. Moss DW, Whitby LG. A simplified heat-inactivation method for investigating alkaline phosphatase isoenzymes in serum. *Clin Chim Acta* 1975;61:63-71.
 29. Mulivor RA, Plotkin LI, Harris H. Differential inhibition of the products of the human alkaline phosphatase loci. *Ann Hum Genet* 1978;42:1-13.
 30. Anderson DJ, Branum EL, O'Brien JF. Liver- and bone-derived isoenzymes of alkaline phosphatase in serum as determined by high-performance affinity chromatography. *Clin Chem* 1990;36:240-6.
 31. Le Bricon T, Gay-Bellile C, Cottu P, et al. Lectin affinity electrophoresis of serum alkaline phosphatase in metastasized breast cancer. *J Clin Lab Anal* 2010;24:20-4.
 32. Hata K, Tokuhiko H, Nakatsuka K, et al. Measurement of bone-specific alkaline phosphatase by an immunoselective enzyme assay method. *Ann Clin Biochem* 1996;33 (Pt 2):127-31.
 33. Broyles DL, Nielsen RG, Bussett EM, et al. Analytical and clinical performance characteristics of Tandem-MP Ostase, a new immunoassay for serum bone alkaline phosphatase. *Clin Chem* 1998;44:2139-47.
 34. Haji SM, Chipchase A, Fraser WD, et al. Retrospective evaluation of a local protocol used to enhance laboratory savings through minimizing the performance of alkaline phosphatase isoenzyme analysis. *Ann Clin Biochem* 2019;56:298-301.
 35. Gonzalez H, Imam Z, Wong R, et al. Normal alkaline phosphatase levels are dependent on race/ethnicity: NationalGEP Health and Nutrition Examination Survey data. *BMJ Open Gastroenterol* 2020;7:e000502.
 36. Wannamethee SG, Shaper AG. Cigarette smoking and serum liver enzymes: the role of alcohol and inflammation. *Ann Clin Biochem* 2010;47:321-6.
 37. EFLM Biological Variation. ALP Liver. [cited 2023 May 12]. Available online: [https://biologicalvariation.eu/search?query=Alkaline%20phosphatase%20\(ALP\),%20liver%20type](https://biologicalvariation.eu/search?query=Alkaline%20phosphatase%20(ALP),%20liver%20type)
 38. McTaggart MP, Rawson C, Lawrence D, et al. Identification of a macro-alkaline phosphatase complex in a patient with inflammatory bowel disease. *Ann Clin Biochem* 2012;49:405-7.
 39. Ramasamy I. Persistent Increase in Serum Alkaline Phosphatase in a Patient with Monoclonal Gammopathy of Undefined Significance. *Case Rep Hematol* 2020;2020:8406971.
 40. Lum G. Significance of low serum alkaline phosphatase activity in a predominantly adult male population. *Clin*

- Chem 1995;41:515-8.
41. Macfarlane JD, Souverijn JH, Breedveld FC. Clinical significance of a low serum alkaline phosphatase. *Neth J Med* 1992;40:9-14.
 42. Kalaria T, Ford C, Gama R. Managing ethylenediaminetetraacetic acid (EDTA) interference in EDTA contaminated samples - selectivity in reporting analytes. *Ann Clin Biochem* 2023;60:92-9.
 43. White G. Serum ethylenediaminetetraacetic acid concentrations in routine samples submitted for biochemical analysis. *Ann Clin Biochem* 2010;47:485-6.
 44. Jain A, Jadhav AA, Varma M. Relation of oxidative stress, zinc and alkaline phosphatase in protein energy malnutrition. *Arch Physiol Biochem* 2013;119:15-21.
 45. Atinmo T, Johnson A, Mbofung C, et al. Plasma zinc status of protein energy malnourished children. *Acta Trop* 1982;39:265-74.
 46. Ray CS, Singh B, Jena I, et al. Low Alkaline Phosphatase (ALP) In Adult Population an Indicator of Zinc (Zn) and Magnesium (Mg) Deficiency. *Curr Res Nutr Food Sci Jour* 2017;5:347-52.
 47. Pettifor JM. Screening for nutritional rickets in a community. *J Steroid Biochem Mol Biol* 2016;164:139-44.
 48. Vasudevan J, Jenifer A, Reddy GM, et al. Serum alkaline phosphatase for screening of hypovitaminosis D. *Indian Pediatr* 2014;51:60-1.
 49. Taylor JA, Richter M, Done S, et al. The utility of alkaline phosphatase measurement as a screening test for rickets in breast-fed infants and toddlers: a study from the puget sound pediatric research network. *Clin Pediatr (Phila)* 2010;49:1103-10.
 50. Haffner D, Leifheit-Nestler M, Grund A, et al. Rickets guidance: part I-diagnostic workup. *Pediatr Nephrol* 2022;37:2013-36.
 51. Waterlow JC. Enzyme changes in malnutrition. *J Clin Pathol Suppl (Assoc Clin Pathol)* 1970;4:75-9.
 52. Kumari R, Rao YN, Talukdar B, et al. Serum enzyme abnormalities in protein energy malnutrition. *Indian Pediatr* 1993;30:469-73.
 53. Adejuwon CA, Akinyinka OO, Ayo-ola BM. Apparent hypocalcaemia in Nigerian children with kwashiorkor. *West Afr J Med* 1994;13:168-70.
 54. Choi HK, Kim GJ, Yoo HS, et al. Vitamin C Activates Osteoblastogenesis and Inhibits Osteoclastogenesis via Wnt/ β -Catenin/ATF4 Signaling Pathways. *Nutrients* 2019;11:506.
 55. Kim GS, Kim CH, Park JY, et al. Effects of vitamin B12 on cell proliferation and cellular alkaline phosphatase activity in human bone marrow stromal osteoprogenitor cells and UMR106 osteoblastic cells. *Metabolism* 1996;45:1443-6.
 56. Fehlmann HU. Scurvy in an adult. *Schweiz Med Wochenschr* 1977;107:1199-202.
 57. Villavicencio Kim J, Wu GY. Celiac Disease and Elevated Liver Enzymes: A Review. *J Clin Transl Hepatol* 2021;9:116-24.
 58. Shaver WA, Bhatt H, Combes B. Low serum alkaline phosphatase activity in Wilson's disease. *Hepatology* 1986;6:859-63.
 59. Weber G, Mora S, Bellini A, et al. Bone mineral metabolism and thyroid replacement therapy in congenital hypothyroid infants and young children. *J Endocrinol Invest* 1995;18:277-82.
 60. Schaefer AK, Hutschala D, Andreas M, et al. Decrease in serum alkaline phosphatase and prognostic relevance in adult cardiopulmonary bypass. *Interact Cardiovasc Thorac Surg* 2020;31:383-90.
 61. Probstfield JL, Statland BE, Gorman L, et al. Alterations in human serum alkaline phosphatase and its isoenzymes by hypolipidemic agents: colestipol and clofibrate. *Metabolism* 1983;32:818-21.
 62. Whitaker KB, Costa D, Moss DW. Selective effects of clofibrate on alkaline phosphatase isoenzymes in serum. *Clin Chim Acta* 1979;94:191-6.
 63. Soma K, Izumi M, Yamamoto Y, et al. In Vitro and In Vivo Pharmacological Profiles of DS-1211, a Novel Potent, Selective, and Orally Bioavailable Tissue-Nonspecific Alkaline Phosphatase Inhibitor. *J Bone Miner Res* 2022;37:2033-43.
 64. Sajid-Ur-Rehman, Saeed A, Saddique G, et al. Synthesis of sulfadiazinyl acyl/aryl thiourea derivatives as calf intestinal alkaline phosphatase inhibitors, pharmacokinetic properties, lead optimization, Lineweaver-Burk plot evaluation and binding analysis. *Bioorg Med Chem* 2018;26:3707-15.
 65. Zizian BV, Gauthier B. Characteristics of the inhibition of serum alkaline phosphatase by theophylline. *Clin Biochem* 1978;11:57-61.
 66. Tsiantouli E, Biver E, Chevalley T, et al. Prevalence of Low Serum Alkaline Phosphatase and Hypophosphatasia in Adult Patients with Atypical Femur Fractures. *Calcif Tissue Int* 2022;110:703-11.
 67. Rosen CJ, Hochberg MC, Bonnick SL, et al. Treatment with once-weekly alendronate 70 mg compared with once-weekly risedronate 35 mg in women with postmenopausal osteoporosis: a randomized double-blind study. *J Bone Miner Res* 2005;20:141-51.

68. Nakamura Y, Suzuki T, Kato H. Serum bone alkaline phosphatase is a useful marker to evaluate lumbar bone mineral density in Japanese postmenopausal osteoporotic women during denosumab treatment. *Ther Clin Risk Manag* 2017;13:1343-8.
69. Costa-Rodrigues J, Reis S, Teixeira S, et al. Dose-dependent inhibitory effects of proton pump inhibitors on human osteoclastic and osteoblastic cell activity. *FEBS J* 2013;280:5052-64.
70. Metaye T, Mettey Y, Lehuède J, et al. Comparative inhibition of human alkaline phosphatase and diamine oxidase by bromo-levamisole, cimetidine and various derivatives. *Biochem Pharmacol* 1988;37:4263-8.
71. Gill M, Sanyal SN, Sareen ML. Interaction of H₂-receptor antagonists, cimetidine and ranitidine with microsomal drug metabolizing and other systems in liver. *Indian J Exp Biol* 1991;29:852-6.
72. Lab Tests Guide. ALP Fractionation Test. [cited 2023 Jul 5]. Available online: <https://www.labtestsguide.com/alp-fractionation-test>
73. Du WX, Duan SF, Chen JJ, et al. Serum bone-specific alkaline phosphatase as a biomarker for osseous metastases in patients with malignant carcinomas: a systematic review and meta-analysis. *J Cancer Res Ther* 2014;10 Suppl:C140-3.
74. Chinoy MA, Javed MI, Khan A, et al. Alkaline phosphatase as a screening test for osteomalacia. *J Ayub Med Coll Abbottabad* 2011;23:23-5.
75. Shaheen S, Noor SS, Barakzai Q. Serum alkaline phosphatase screening for vitamin D deficiency states. *J Coll Physicians Surg Pak* 2012;22:424-7.
76. Scappaticcio L, Longo M, Maiorino MI, et al. Abnormal Liver Blood Tests in Patients with Hyperthyroidism: Systematic Review and Meta-Analysis. *Thyroid* 2021;31:884-94.
77. Yang M, Zhang L, Huang L, et al. Risk Factors for Elevated Preoperative Alkaline Phosphatase in Patients with Refractory Secondary Hyperparathyroidism. *Am Surg* 2017;83:1368-72.
78. Lieberman D, Phillips D. "Isolated" elevation of alkaline phosphatase: significance in hospitalized patients. *J Clin Gastroenterol* 1990;12:415-9.
79. Han PKJ, Babrow A, Hillen MA, et al. Uncertainty in health care: Towards a more systematic program of research. *Patient Educ Couns* 2019;102:1756-66.
80. Kumar M, Herrera JL. Sarcoidosis and the Liver. *Clin Liver Dis* 2019;23:331-43.
81. De Mulder P, Maertens B, Hoorens A, et al. Extrapulmonary sarcoidosis primarily presenting as cholestatic liver disease. *BMJ Case Rep* 2019;12:e232618.
82. Shipman KE, Holt AD, Gama R. Interpreting an isolated raised serum alkaline phosphatase level in an asymptomatic patient. *BMJ* 2013;346:f976.
83. Lindor KD, Kowdley KV, Harrison ME, et al. ACG Clinical Guideline: Primary Sclerosing Cholangitis. *Am J Gastroenterol* 2015;110:646-59.
84. Kwo PY, Cohen SM, Lim JK. ACG Clinical Guideline: Evaluation of Abnormal Liver Chemistries. *Am J Gastroenterol* 2017;112:18-35.
85. Newsome PN, Cramb R, Davison SM, et al. Guidelines on the management of abnormal liver blood tests. *Gut* 2018;67:6-19.
86. van Ooteghem NA, Klomp LW, van Berge-Henegouwen GP, et al. Benign recurrent intrahepatic cholestasis progressing to progressive familial intrahepatic cholestasis: low GGT cholestasis is a clinical continuum. *J Hepatol* 2002;36:439-43.
87. Chapman MH, Thorburn D, Hirschfield GM, et al. British Society of Gastroenterology and UK-PSC guidelines for the diagnosis and management of primary sclerosing cholangitis. *Gut* 2019;68:1356-78.
88. EASL Clinical Practice Guidelines: management of cholestatic liver diseases. *J Hepatol* 2009;51:237-67.
89. Hirschfield GM, Dyson JK, Alexander GJM, et al. The British Society of Gastroenterology/UK-PBC primary biliary cholangitis treatment and management guidelines. *Gut* 2018;67:1568-94.
90. Myszor M, James OF. The epidemiology of primary biliary cirrhosis in north-east England: an increasingly common disease? *Q J Med* 1990;75:377-85.
91. Lindor KD, Bowlus CL, Boyer J, et al. Primary Biliary Cholangitis: 2018 Practice Guidance from the American Association for the Study of Liver Diseases. *Hepatology* 2019;69:394-419.
92. Kaplan MM, Gershwin ME. Primary biliary cirrhosis. *N Engl J Med* 2005;353:1261-73.
93. Nakamura M, Kondo H, Mori T, et al. Anti-gp210 and anti-centromere antibodies are different risk factors for the progression of primary biliary cirrhosis. *Hepatology* 2007;45:118-27.
94. Maggs JR, Chapman RW. An update on primary sclerosing cholangitis. *Curr Opin Gastroenterol* 2008;24:377-83.
95. Hirschfield GM, Gershwin ME. The immunobiology and pathophysiology of primary biliary cirrhosis. *Annu Rev Pathol* 2013;8:303-30.
96. Chapman RW, Arborgh BA, Rhodes JM, et al.

- Primary sclerosing cholangitis: a review of its clinical features, cholangiography, and hepatic histology. *Gut* 1980;21:870-7.
97. Boberg KM, Fausa O, Haaland T, et al. Features of autoimmune hepatitis in primary sclerosing cholangitis: an evaluation of 114 primary sclerosing cholangitis patients according to a scoring system for the diagnosis of autoimmune hepatitis. *Hepatology* 1996;23:1369-76.
 98. Mendes FD, Jorgensen R, Keach J, et al. Elevated serum IgG4 concentration in patients with primary sclerosing cholangitis. *Am J Gastroenterol* 2006;101:2070-5.
 99. Björnsson E, Chari ST, Smyrk TC, et al. Immunoglobulin G4 associated cholangitis: description of an emerging clinical entity based on review of the literature. *Hepatology* 2007;45:1547-54.
 100. Björnsson E, Chari S, Silveira M, et al. Primary sclerosing cholangitis associated with elevated immunoglobulin G4: clinical characteristics and response to therapy. *Am J Ther* 2011;18:198-205.
 101. Hov JR, Boberg KM, Karlsen TH. Autoantibodies in primary sclerosing cholangitis. *World J Gastroenterol* 2008;14:3781-91.
 102. Corpechot C, El Naggar A, Poujol-Robert A, et al. Assessment of biliary fibrosis by transient elastography in patients with PBC and PSC. *Hepatology* 2006;43:1118-24.
 103. Exeter Laboratory. Alkaline Phosphatase (ALP). 2019. [cited 2023 Jul 5]. Available online: <https://www.exeterlaboratory.com/test/alkaline-phosphatase/>
 104. McPherson GS, Jani BR. Isolated increase in serum alkaline phosphatase with sodium valproate therapy. *Ann Pharmacother* 1995;29:539.
 105. Cepelak I, Rekić B, Juretić D, et al. Effect of sodium valproate on renal cell brush-border enzymes in rats. *Eur J Clin Chem Clin Biochem* 1995;33:673-7.
 106. Burgunder JM, Abernethy DR, Lauterburg BH. Liver injury due to verapamil. *Hepatogastroenterology* 1988;35:169-70.
 107. Iqbal U, Siddiqui HU, Anwar H, et al. Allopurinol-Induced Granulomatous Hepatitis: A Case Report and Review of Literature. *J Investig Med High Impact Case Rep* 2017;5:2324709617728302.
 108. Math MV. Furosemide and increased serum alkaline phosphatase (hepatic isoenzyme). *Clin Chem* 1982;28:1812-3.
 109. Kwda A, Gldc P, Bauj B, et al. Effect of long term inhaled corticosteroid therapy on adrenal suppression, growth and bone health in children with asthma. *BMC Pediatr* 2019;19:411.
 110. Chamani S, Liberale L, Mobasheri L, et al. The role of statins in the differentiation and function of bone cells. *Eur J Clin Invest* 2021;51:e13534.
 111. Boks MN, Tiebosch AT, van der Waaij LA. A jaundiced bodybuilder Cholestatic hepatitis as side effect of injectable anabolic-androgenic steroids. *J Sports Sci* 2017;35:2262-4.
 112. Magnusson P, Häger A, Larsson L. Serum osteocalcin and bone and liver alkaline phosphatase isoforms in healthy children and adolescents. *Pediatr Res* 1995;38:955-61.
 113. Nizet A, Cavalier E, Stenvinkel P, et al. Bone alkaline phosphatase: An important biomarker in chronic kidney disease - mineral and bone disorder. *Clin Chim Acta* 2020;501:198-206.
 114. Uday S, Högler W. Rickets and Osteomalacia. *Encyclopedia of Endocrine Diseases* 2019;5:339-54.
 115. Whyte MP, Thakker RV. Rickets and osteomalacia. *Medicine* 2013;41:594-9.
 116. Magne D, Bluteau G, Fauchoux C, et al. Phosphate is a specific signal for ATDC5 chondrocyte maturation and apoptosis-associated mineralization: possible implication of apoptosis in the regulation of endochondral ossification. *J Bone Miner Res* 2003;18:1430-42.
 117. Sabbagh Y, Carpenter TO, Demay MB. Hypophosphatemia leads to rickets by impairing caspase-mediated apoptosis of hypertrophic chondrocytes. *Proc Natl Acad Sci U S A* 2005;102:9637-42.
 118. Ralston SH, Langston AL, Reid IR. Pathogenesis and management of Paget's disease of bone. *Lancet* 2008;372:155-63.
 119. Reid IR. Recent advances in understanding and managing Paget's disease. *F1000Res* 2019;8:F1000 Faculty Rev-1485.
 120. Alvarez L, Peris P, Pons F, et al. Relationship between biochemical markers of bone turnover and bone scintigraphic indices in assessment of Paget's disease activity. *Arthritis Rheum* 1997;40:461-8.
 121. Delmas PD, Meunier PJ. The management of Paget's disease of bone. *N Engl J Med* 1997;336:558-66.
 122. Corral-Gudino L, Tan AJ, Del Pino-Montes J, et al. Bisphosphonates for Paget's disease of bone in adults. *Cochrane Database Syst Rev* 2017;12:CD004956.
 123. Langston AL, Campbell MK, Fraser WD, et al. Randomized trial of intensive bisphosphonate treatment versus symptomatic management in Paget's disease of bone. *J Bone Miner Res* 2010;25:20-31.
 124. Tan A, Goodman K, Walker A, et al. Long-Term Randomized Trial of Intensive Versus Symptomatic Management in Paget's Disease of Bone: The PRISM-EZ

- Study. *J Bone Miner Res* 2017;32:1165-73.
125. Singer FR. The evaluation and treatment of Paget's disease of bone. *Best Pract Res Clin Rheumatol* 2020;34:101506.
 126. Kling JM, Clarke BL, Sandhu NP. Osteoporosis prevention, screening, and treatment: a review. *J Womens Health (Larchmt)* 2014;23:563-72.
 127. Nakagawa H, Kamimura M, Takahara K, et al. Changes in total alkaline phosphatase level after hip fracture: comparison between femoral neck and trochanter fractures. *J Orthop Sci* 2006;11:135-9.
 128. Dresner-Pollak R, Mayer M, Hochner-Celiniker D. The decrease in serum bone-specific alkaline phosphatase predicts bone mineral density response to hormone replacement therapy in early postmenopausal women. *Calcif Tissue Int* 2000;66:104-7.
 129. Cheng SL, Yang JW, Rifas L, et al. Differentiation of human bone marrow osteogenic stromal cells in vitro: induction of the osteoblast phenotype by dexamethasone. *Endocrinology* 1994;134:277-86.
 130. Hardy RS, Zhou H, Seibel MJ, et al. Glucocorticoids and Bone: Consequences of Endogenous and Exogenous Excess and Replacement Therapy. *Endocr Rev* 2018;39:519-48.
 131. Hyldstrup L, Clemmensen I, Jensen BA, et al. Non-invasive evaluation of bone formation: measurements of serum alkaline phosphatase, whole body retention of diphosphonate and serum osteocalcin in metabolic bone disorders and thyroid disease. *Scand J Clin Lab Invest* 1988;48:611-9.
 132. Mikosch P. Effects of thyroid disorders on the bone. *Wien Med Wochenschr* 2005;155:444-53.
 133. Lehmann FG. Immunological methods for human placental alkaline phosphatase (Regan isoenzyme). *Clin Chim Acta* 1975;65:271-82.
 134. Moss DW. Diagnostic aspects of alkaline phosphatase and its isoenzymes. *Clin Biochem* 1987;20:225-30.
 135. Hada T, Higashino K, Okochi T, et al. Kasahara-variant alkaline phosphatase in a renal cell carcinoma. *Clin Chim Acta* 1978;89:311-6.
 136. Wei SC, Doellgast GJ. Immunochemical studies of human placental-type variants of alkaline phosphatase. Structural differences between the "Nagao isoenzyme" and the placental "D-variant". *Eur J Biochem* 1981;118:39-45.
 137. Behúlová D, Bzdúch V, Holesová D, et al. Transient hyperphosphatasemia of infancy and childhood: study of 194 cases. *Clin Chem* 2000;46:1868-9.
 138. Gualco G, Lava SA, Garzoni L, et al. Transient benign hyperphosphatasemia. *J Pediatr Gastroenterol Nutr* 2013;57:167-71.
 139. Stein P, Rosalki SB, Foo AY, et al. Transient hyperphosphatasemia of infancy and early childhood: clinical and biochemical features of 21 cases and literature review. *Clin Chem* 1987;33:313-8.
 140. Huh SY, Feldman HA, Cox JE, et al. Prevalence of transient hyperphosphatasemia among healthy infants and toddlers. *Pediatrics* 2009;124:703-9.
 141. O'Riordan S, Baker AJ, Sherwood RA. Isoenzyme characterization in isolated elevation of alkaline phosphatase after liver transplantation in children. *Transplantation* 2002;74:1030-4.
 142. Sampani E, Kasimatis E, Memmos E, et al. Transient Hyperphosphatasemia in an Adolescent and an Adult Renal Transplant Patient. *Transplant Proc* 2021;53:2769-70.
 143. Maldonado O, Demasi R, Maldonado Y, et al. Extremely high levels of alkaline phosphatase in hospitalized patients. *J Clin Gastroenterol* 1998;27:342-5.
 144. Aida S. Alkaline phosphatase isoenzyme activities in rheumatoid arthritis: hepatobiliary enzyme dissociation and relation to disease activity. *Ann Rheum Dis* 1993;52:511-6.
 145. Siede WH, Seiffert UB, Merle S, et al. Alkaline phosphatase isoenzymes in rheumatic diseases. *Clin Biochem* 1989;22:121-4.
 146. Ryu WS, Lee SH, Kim CK, et al. High serum alkaline phosphatase in relation to cerebral small vessel disease. *Atherosclerosis* 2014;232:313-8.
 147. Kang KY, Hong YS, Park SH, et al. Increased serum alkaline phosphatase levels correlate with high disease activity and low bone mineral density in patients with axial spondyloarthritis. *Semin Arthritis Rheum* 2015;45:202-7.
 148. Cho SR, Lim YA, Lee WG. Unusually high alkaline phosphatase due to intestinal isoenzyme in a healthy adult. *Clin Chem Lab Med* 2005;43:1274-5.
 149. Bayer PM, Hotschek H, Knoth E. Intestinal alkaline phosphatase and the ABO blood group system--a new aspect. *Clin Chim Acta* 1980;108:81-7.
 150. Matsushita M, Harajiri S, Tabata S, et al. Alkaline phosphatase activity in blood group B or O secretors is fluctuated by the dinner intake of previous night. *Rinsho Byori* 2013;61:307-12.
 151. Verma J, Gorard DA. Persistently elevated alkaline phosphatase. *BMJ Case Rep* 2012;2012:bcr2012006768.
 152. Rosalki SB, Foo AY, Dooley JS. Benign familial hyperphosphatasemia as a cause of unexplained increase in plasma alkaline phosphatase activity. *J Clin Pathol* 1993;46:738-41.
 153. Anh DJ, Eden A, Farley JR. Quantitation of soluble

- and skeletal alkaline phosphatase, and insoluble alkaline phosphatase anchor-hydrolase activities in human serum. *Clin Chim Acta* 2001;311:137-48.
154. Magnusson P, Sharp CA, Farley JR. Different distributions of human bone alkaline phosphatase isoforms in serum and bone tissue extracts. *Clin Chim Acta* 2002;325:59-70.
155. Zierk J, Arzideh F, Haeckel R, et al. Pediatric reference intervals for alkaline phosphatase. *Clin Chem Lab Med* 2017;55:102-10.
156. Colantonio DA, Kyriakopoulou L, Chan MK, et al. Closing the gaps in pediatric laboratory reference intervals: a CALIPER database of 40 biochemical markers in a healthy and multiethnic population of children. *Clin Chem* 2012;58:854-68.
157. Zhan F, Watanabe Y, Shimoda A, et al. Evaluation of serum bone alkaline phosphatase activity in patients with liver disease: Comparison between electrophoresis and chemiluminescent enzyme immunoassay. *Clin Chim Acta* 2016;460:40-5.
158. Curtis EM, Parsons C, Maslin K, et al. Bone turnover in pregnancy, measured by urinary CTX, is influenced by vitamin D supplementation and is associated with maternal bone health: findings from the Maternal Vitamin D Osteoporosis Study (MAVIDOS) trial. *Am J Clin Nutr* 2021;114:1600-11.
159. Walker I, Chappell LC, Williamson C. Abnormal liver function tests in pregnancy. *BMJ* 2013;347:f6055.

doi: 10.21037/jlpm-23-63

Cite this article as: Irving M, Shipman AR, Shipman KE. Investigative algorithms for disorders affecting human plasma alkaline phosphatase: a narrative review. *J Lab Precis Med* 2024;9:7.