

Exploiting homologous recombination repair alterations in metastatic castrate-resistant prostate cancer: a case report and literature review

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Abstract: DNA damage can occur through a variety of processes and thus DNA repair is essential for cells to maintain their genomic integrity. Homologous recombination repair (HRR) is an important cellular mechanism by which double strand DNA breaks (DSB) are corrected with high fidelity. Proteins involved in this process include BRCA1, BRCA2, RAD51, and PALB2. Alterations in this mechanism can be found in a diverse set of malignancies and evidence exists that targeting these deficiencies can result in clinical benefit. Here we present a case of a patient with relapsed, metastatic castrate-resistant prostate cancer (mCRPC) that was refractory to standard of care therapies inducing androgen blockade and taxanes. He subsequently was found to have a PALB2 mutation on molecular analysis (next generation sequencing) of his tumor which was later confirmed to be germline. Treatment with carboplatin followed by a PARP-inhibitor led to rapid control of his advanced disease. This article will review the patient's course along with the mechanisms of DNA damage repair with a focus on HRR and the rationale for therapies targeted to exploit alterations of this process.

Keywords: Metastatic castrate-resistant prostate cancer (mCRPC); homologous recombination repair (HRR); PALB2; PARP-inhibition; case report

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Introduction

An elderly male patient with heavily pretreated metastatic castrate-resistant prostate cancer (mCRPC) was found to have a germline PALB2 alteration along with other somatic alterations in HRR pathway genes and had a significant response to platinum followed by PARP inhibitor therapy.

Case presentation

Our patient is a 70-year-old African-American male with a history of asthma, hyperlipidemia, and a spindle cell neoplasm of his mesentery treated with right hemicolectomy at age 52 who was diagnosed at age 60 with localized prostate cancer. Family history was notable for a mother diagnosed with breast cancer in her 60's and a grandchild diagnosed with brain cancer at 18 months. His pre-treatment PSA was 7.5 ng/mL and a metastatic workup was negative. He underwent radical prostatectomy which showed a pathologic stage T3b/N0 adenocarcinoma with a Gleason score of 4+4 (tertiary pattern 5). He then completed a course of adjuvant radiation for positive surgical margins. He experienced a biochemical relapse over the next year with a PSA of 5 that was treated with androgen deprivation therapy (ADT) consisting of leuprolide injections given every 3 months along with denosumab. After an initial response, his PSA steadily increased over the next three years (PSA 4.3, doubling time 1.7 months) and imaging revealed bone metastases. Given progression to mCRPC enzalutamide was added to ADT followed by another prolonged biochemical response.



Figure 1 Patient timeline.



Figure 2 PSA curve during treatment with carboplatin and PARPi.

Later his PSA began to increase and radium-223 was given without clear benefit. The PSA continued to rise and imaging showed progression of bone metastases and new pelvic and retroperitoneal lymphadenopathy. Subsequently he received 5 cycles of docetaxel and 3 cycles of cabazitaxel but again progressed radiographically (new liver and lung metastases), biochemically (peak PSA 2,628) and clinically with worsening fatigue, bone pain, weight loss, and bulky adenopathy-related leg edema. See *Figure 1* for a timeline of significant events.

Tumor management

Next generation sequencing (NGS) was ordered from a prior biopsy specimen and revealed a microsatellite stable

tumor with a tumor mutational burden of 8 mutations/ Mb (intermediate by FoundOne), loss of FAS and PTEN, and mutations in CDK12, FANCA, KEL, and PALB2 [c.2878delC(p.Leu960Tyrfs*2)] (see Table S1). After consultation with the NGS service (Foundation One) the PALB2 mutation was deemed likely germline heterozygous and the patient was referred to Genetic Counseling. There an Integrated BRCA Analysis with Myriad myRisk Genetic testing was ordered and confirmed the heterozygous PALB2 mutation along with three variants of undetermined significance in ATM, BRIP1 and PMS2 (see Table S2). The patient was discussed at Molecular Tumor Board and the decision was made to proceed with carboplatin (AUC 6, every 3 weeks) due to initial cost issues with PARP inhibitors (PAPRi). He eventually received 7 doses with improvement in clinical symptoms and a decrease in his PSA from 2,628 to 62 (see Figure 2). Imaging at 4 months showed a mixed response then at 6 months marked improvement of osseous lesions on nuclear bone scan with decrease in size of retroperitoneal adenopathy and pulmonary nodules on CT (see Figures 3-5). Carboplatin was eventually discontinued due to fatigue and cytopenias. During this break from carboplatin PSA increased to 95. The PARPi olaparib was obtained with the help of a pharmaceutical company prescription savings program (AstaZeneca, AZ&ME). The patient tolerated olaparib 300 mg BID well and while at one month his PSA had risen from 95 to 224 by month two it had returned to 99 followed by a further decline to 57 at three months. Imaging at three months post PARPi initiation revealed a partial response with stable pulmonary and mediastinal nodules and further decreased abdominal nodules and bone lesions. He continues on the medication as of this writing.

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Figure 3 Sequential CT scans of pelvis highlighting treatment responses.



Figure 4 Sequential CT scans of abdomen highlighting treatment responses.



Figure 5 Sequential CT scans of chest highlighting treatment responses.

Case-related literature review

Homologous recombination involves the use of a DNA template, such as a sister chromatid or homologous chromosome, to exchange nucleotide bases between DNA molecules. This process contributes to genetic variation in meiosis and is crucial in the repair of damaged DNA. Eukaryotic DNA is constantly undergoing damage, with as many as 10⁵ DNA lesions of various kinds per day occurring in one cell (1). DNA damage repair (DDR) encompasses a variety of cellular mechanisms that, when defective, can lead to developmental issues, tumorigenesis, and cellular death.

Homologous recombination repair (HRR) has been called a conservative process in that it can recapitulate the

original genetic sequence with high fidelity as opposed to some other process of DDR such as single strand annealing (SSA) or non-homologous end joining (NHEJ) (2). HHR is a means for cells to restore their genomic integrity in the setting of double stranded breaks (DSBs) but is also involved in repair of inter-strand crosslinks (3). Inter-strand crosslinking is one of the mechanisms via which alkylating agents, such as the nitrogen mustards, and platinum agents affect cellular toxicity.

DSBs are significantly toxic to cells and entail concurrent fractures of the phosphate backbones of complimentary DNA strands (4). These breaks can be caused by ionizing radiation, chemical or drug effects (i.e., bleomycin), through cell intrinsic processes such as when DNA polymerase encounters nicks in DNA or other means of stress related to genomic replication (5).

Recognition of genetic damage is the first step in DDR and the activation of oxidative-stress related proteins such as SIRT6 allow for recruitment of PARP1 to DSBs (6). This is followed rapidly (within seconds) by PARP1 regulated chromatin remodeling to allow for access to the affected strands of DNA (7). ATM is auto-phosphorylated in the setting of DBSs and, once activated, affects multiple downstream molecules including BRCA1 and histone variant H2AX, contributing to de-condensation of chromatin (1,8).

In HRR the MRN complex (nuclease Mre11-Rad50-Xrs2 with cofactor Sae2) binds to each broken end of DNA (9). The 5' ends are resected to create 3' overhangs of the complimentary DNA strands. Further 5 to 3 prime resections occur followed by binding of RPA to each 3' overhanging single strand. With the help of mediators RAD51 recombinase overtakes the RPA binding and produces RAD51 filaments in an ATP dependent fashion that act as scaffolds (10). These mediators involve both positive regulators such as other RAD proteins (i.e., RAD52), BRCA1 and 2, and PALB2 and negative regulators such as FANCJ (9). These nucleoprotein filaments (often referred to as the presynaptic complex) then search for and locate a complimentary sequence of DNA scaffolds (10). Strand invasion occurs as the free 3' filament interposes itself within the selected double stranded DNA thus displacing its homolog and forming base pairs with its complementary strand. This is known as the displacement loop or D-loop as it involves breaking of the previously connecting double stranded DNA base pairs (9). With the help of other proteins DNA polymerases are then able to extend the 3' prime invading strand with the use of this

complimentary sequence. The strand invasion process, D-loop extension, and eventual D-loop dissociation are likely influenced by many factors including the topography of the potential DNA partner strand, with a preference for negatively super-coiled domains (9).

Subsequently two primary models for completion of the DSB repair process exist, double strand break repair (DSBR) and synthesis-dependent strand annealing (SDSA). Briefly SDSA involves release of the extended invading strand, annealing of this strand to its original strand's broken end, removal of excess DNA, further nucleotide synthesis to fill gaps followed by ligation without producing any crossover events (11). DSBR involves the creation of double Holiday junctions involving both the invading strand and the original 3' DNA non-invading strand with the selected DS homologues. This is followed by further DNA synthesis and ligation, which, depending on the sites of cutting and homing of the recombination products, will most commonly lead to chromosomal crossover (11). In human cell DDR there is an apparent preference for SDSA, which is unsurprising given it does not lead to crossover events thus maintaining original genetic integrity (12).

There are different pathways by which DSBs can be repaired and competition between these processes exists intracellularly. Early binding of the NHEJ complex prevents HRR initialization due HRR's requirement for a resected DNA end (2). The timing of the cell cycle also contributes to the type of DDR mechanism that cells utilize. HHR occurs once mammalian cells have produced sister chromatids so that these can be used as templates, thus it appears to be primarily active in the later S phase and G2 phase of the cell cycle when these chromatids are available. This temporal regulation may occur via decreased CDK meditated phosphorylation of BRCA2's RAD51 binding site (13). Other repair processes, specifically the afore-mentioned NHEJ, predominate during other phases of the cell cycle and, given their mechanistic flexibility and template independence (14), are the primary means of DDR when cellular HHR is deficient. HHR may be more dynamic however, with some evidence that it may be able to overcome cell cycle restrictions (13).

Relevant to our patient PALB2 (Partner And Localizer of BRCA2) appears have multiple functions important to HRR including recruitment of RAD51 and BRCA2 to DSBs (15), binding BRCA2 and promoting the activity of the complex to promote the polymerization of the RAD51 filament (16), and assisting RAD51 polymers with strand invasion (17). The mutation found in our patient [c.2878delC(p.

Leu960Tyrfs*2)] appears to result in a frameshift variant and the location of the deletion (WD40 domain) is likely involved in the binding of PALB2 to BRCA2, RAD51, and RAD51C thus affecting HRR (17,18). The finding that RAD51 is often up-regulated in BRCA1 deficient breast tumors suggests that the molecules in this process may compensate for each other in some malignancies (19). HRR defects promote tumorigenesis by forcing cells to rely on non-conservative and error-prone DDR mechanisms thus increasing mutational rate potentially leading to alterations that promote growth and cell survival. However the resultant malignant cells must balance the benefits of this increased rate of mutation with the potential for cellular death that can result from these errors.

Poly ADP-ribose polymerases (PARPs) compromise a group of enzymes that can catalyze the synthesis of PAR chains. The PARP enzymes (best characterized are PARP1 and PARP2) are relevant to DDR through their ability to recognize damage in DNA, bind to strand breaks and recruit enzymes that continue the DDR process. Their involvement in DDR was recognized after PARP levels were found to be increased after radiation damage to DNA and in the setting of increased DNA lesions in general (20). PARPs are involved in the processes of base excision repair and nucleotide excision repair in response to single stranded breaks (SSBs) in DNA (20). While SSBs in DNA are the thought to be the main target of PARPs, PARP1, as described earlier, is also implicated in the early stages DSB repair. Following DNA repair it is also necessary for the PARP complex to leave the site of DNA damage and there is evidence that PARP inhibitors may cause further cellular toxicity by inhibiting this post-repair dissociation thus stalling the replication fork (21). Synthetic lethality is the concept that while one defective process may not be enough to compromise cellular survival the additional inhibition of another related process may induce death. Thus the targeting of SSB repair (which if un-repaired can cause DSBs) and DDR sensing/initiation by PARP inhibition in the setting of deficient HHR may overwhelm the ability of tumor cells to survive (21).

The frequency of DDR and more specifically HRR alterations have varied across studies. An analysis of the 333 primary prostate carcinoma samples revealed inactivation of DNA repair genes in 19% (22) with a more recent investigation showing an association between pathogenic DDR mutations in tumors and more advanced tumor stages and Gleason grade groups.(23) An examination of men with metastatic prostate cancer, unselected for family history or age, found germline mutations in DDR processes of 11.8% and that they were associated with worse prognoses (24,25). The most frequent germline mutations were in BRCA2 (5.35%), CHEK2 (1.87%) and ATM (1.59%) with BRCA1 (0.87%), RAD51D (0.43%) and PALB2 (0.43%) seen less often. Another study focusing on HRR in advanced prostate cancer included a subset of 221 patients with germline testing; 19% harbored a known or likely pathogenic germline mutation in an HRR gene. Of these 221 patients 27% were found to have any alteration (germline and/ or somatic) in BRCA2, BRCA1, ATM or CHEK2 with germline analysis capturing only about half of those with likely HHR deficiency. PALB2 and ATM specifically were enriched in metastatic castrate-resistant versus metastatic castrate sensitive disease (26). For comparison in an Ashkenazi Jewish population of men with localized prostate cancer BRCA1 & 2 mutations were found in 3% (27). Another important point is that loss of function alterations in the HRR pathway often affects multiple alleles. Robinson et al reported biallelic BRCA2 loss in 90% of patients with BRCA2 alterations usually resulting from a somatic point mutation combined with loss of heterozygosity (28). The NCCN currently recommends germline evaluation of HHR genes (BRCA1, BRCA2, ATM, PALB2, CHEK2) and those involved in Lynch Syndrome/mismatch repair (MHL1, MSH2, MSH6, PMS2) for patients with high risk, very high risk, regionally advanced (N1) or metastatic prostate cancer; it states molecular testing of tumor tissue for these alterations should be considered (NCCN Guidelines Version 4.2019). See Figures 6 and 7 for HRR alteration frequency in data combined from multiple studies in mCRPC.

Our patient's tumor sample also harbored mutations in CDK12 and FANCA both of which are involved in the process of DNA damage repair. A patient with metastatic prostate cancer with a FANCA germline mutation and somatic loss of heterozygosity of FANCA was reported to have a dramatic long term response to cisplatin-docetaxel with further in vitro studies confirming this mutation's increased sensitivity to DNA damaging agents (29). Whether our patient's sensitivity to platinum and PARPinhibitor therapy was due to the germline PALB2 mutation combined with LOH versus the mutations in CDK12 and FANCA is unclear. The germline abnormalities of undetermined significance in ATM, BRIP1 and PSM2, which were not found in his tumor sample and have DDR functions, are also of interest in light of his personal and family cancer history.



Data combined from 3 studies: de Bono Annals of Oncology 2019*, Robinson Cell 2015**, Mateo NEJM 2015***
* BRCA1 & 2, ATM, CDK12, CHEK2, PPP2R2A, PALB2, BRIP1, RAD54L, BARD1,

RAD51B, RAD51D, CHEK1, FANCL, RAD51C ** BRCA1 & 2, ATM, CDK12, FANCA, RAD51B, RAD51C,

*** BRCA 1 & 2, ATM, FANCA, CHEK2, PALB2, HDAC2, RAD51

Figure 6 Combined frequency of homologous recombination repair alterations in mCRPC. mCRPC, metastatic castrateresistant prostate cancer.



Robinson Cell 2015, Mateo NEJM 2015

Figure 7 Frequency of germline HRR mutations in mCRPC patients with identified HRR alterations. HRR, homologous recombination repair; mCRPC, metastatic castrate-resistant prostate cancer.

Prior studies in unselected patients with metastatic prostate cancer treated with platinum agents resulted in modest clinical benefit without an increase in overall

survival (30,31). As knowledge of DDR and HRR pathways and their sensitives has grown populations with these abnormalities have received more focus. A retrospective analysis targeting patients with HRR deficiency treated with carboplatin/docetaxel revealed extremely encouraging results with PSA responses in 75% of patients with germline BRCA2 mutations compared to 17% in those without (32). High response rates (88%) with PARPi have also been seen in patients with refractory prostate cancer who harbor DDR defects and were seen in patients with both germline and somatic alterations (33). More recent studies add further support to this treatment strategy. The TOPARP-B phase II trial treated mCRPC patients, selected for DDR alterations, with two different dose of olaparib. Varying radiologic response rates were seen depending on the genetic lesion with the highest rates in the BRCA1/2 (80%) and PABL2 (57%) mutated subgroups (34). The phase III PROfound trial compared olaparib to enzalutamide or abiraterone in patients with mCRPC with HRR alterations with the one group containing only those with BRCA1/2 or ATM mutations and the other including alterations of any HRR gene. Significant improvements in 12 month radiographic progression-free survival were seen in both groups (35). See Table 1 for preliminary PARPi Phase II trial results and Table 2 for ongoing Phase III clinical trials.

Conclusions

Once a defect is detected in the HHR pathway there is a clear rationale for use of inter-strand cross-linking agents or PARP inhibition as well as other potential methods that, via DNA damage, would force cells to rely on more error prone repair mechanisms. Responses appear to occur regardless of whether the alterations in the HHR pathway are germline or somatic thus lending support for the evaluation of both in patients with advanced disease. In metastatic CRPC refractory to standard treatments a combination of germline and somatic testing seems a reasonable clinical algorithm based on mounting evidence for actionable mutations in this disease.

Future areas to explore include new targets for simultaneous inhibition of different DDR mechanisms. For example would concurrently targeting topoisomerase be useful as it is likely necessary for the strand invasion process in HHR or would it be limited by excessive toxicity (36,37)? What are the ways in which tumors harboring DDR defects evolve to balance the positive and negative selection

Table 1 Notable rep	orted clinical	l trials of PA	ARPi in prostate cance	er				
PARP inhibitor	Name	Phase	Condition	Mutation selection	Patients	Primary outcome	Results	NCT
Olaparib	TOPARP-E	3 Phase II	mCRPC Post progression on one or more taxanes	Selected for DDR a alterations (germline or somatic, mono- or bi-allelic) by tumor sequencing	98 patients randomized, 92 treated and evaluable	Response rate (RR): RECIST 1.1 and/or PSA decline >50% and/or circulating turnor cell count (CTC) conversion to less than 5 after 4 weeks	Olaparib 400 mg BID: RR 54%; olaparib 300 mg BID: RR 37%; with median f/u 17.6 months (mo), overall median PFS of 5.4 mo	NCT01682772
Olaparib (300 mg BID) + Abiraterone vs. Abiraterone + placebo		Phase II	mCRPC; post treatment with docetaxel and candidates for abiraterone	Unselected for DDR or HRR alterations	142 patients randomized, 71 to each arm	Investigator-assessed radiographic progression-free survival (rPFS)	Olaparib + Abiraterone: rPFS 13.8 mo; abiraterone + placebo: rPFS 8.2 mo; hazard Ratio 0.65	NCT01972217
Olaparib (300 mg BID) vs. Abirateron¢ or Enzalutimade (Physicians Choice or PC)	PROfound	Phase III	mCRPC; post progression on prior new hormonal agent therapy	HRR alterations centrally and prospectively collected (foundation medicine)	Cohort A: 245 patients w/BRCA1/2 or ATM alterations; Cohort B: 142 patients with other HRR alterations	rPFS by blinded independent central review	Cohort A: Olaparib rPFS 7.39 vs. 3.55 mo for PC; Cohort A + B: Olaparib rPFS 5.82 vs. 3.52 mo for PC	NCT02987543
Niraparib (300 mg daily)	GALAHAD	Phase II	mCRPC; post progression on androgen signaling inhibitors and taxanes	Selected for biallelic DNA-repair gene j defects (DRD) by plasma assay	123 patients with DRD, 39 with bi-allelic DRD mutations	Composite response rate (RR): RECIST 1.1, CTC conversion to less than 5/7.5 mL, PSA decline >50%	RR 65% in biallelic BRCA1/2 (n=23); RR 31% in biallelic non- BRCA (n=16)	NCT02854436
Rucaparib (600 mg BID)	TRITON-2	Phase II	mCRPC; post progression on 1–2 lines of androgen receptor therapy and 1 taxane	Germline or somatic alterations in BRCA1/2 or 13 other HRR genes	52 patients treated	Centrally assessed objective response rate per modified RECIST v1.1/PCWG3 in patients with measurable disease and PSA decrease >50% in those without	Median f/u 3.7 mo; 11 of 23 (48%) evaluable BRCA altered patients had PSA response; 5 of 11 (46%) evaluable BRCA altered patients had radiographic response	NCT02952534
mCRPC, metastatic	: castrate-re	sistant pros	state cancer.					

prostate cancer	Comparator
Ongoing phase III studies of PARPi in	PARP inhibitor
Table 2	Name

Name	PARP inhibitor	Comparator	Condition	Primary outcome	NCT
PROpel	Olaparib + Abiraterone	Placebo + Abiraterone	First line in mCRPC	Radiologic Progression Free Survival	NCT0373280
KEYLINK-010	Olaparib + Pembrolizumab with Abiraterone or Enzalutamide	Abiraterone or Enzalutamide	mCRPC, unselected for HRR defects Post progression on one next gen hormonal agent	. Overall Survival	NCT03834519
I	Niraparib + Abiraterone	Abiraterone + Placebo	Metastatic Prostate Cancer, DDR alterations assessed pre-treatment	Radiographic Progression Free Survival	NCT03748641
I	Talazoparib + Enzalutamide	Enzalutamide + Placebo	mCRPC, first line treatment once documented metastatic	Radiographic Progression Free Survival	NCT03395197
TRITON3	Rucaparib	Physician's Choice: Abiraterone, Enzalutamide, or Docetaxel	mCRPC, selected for HRR alterations	Radiographic Progression Free Survival	NCT02975934
mCRPC, metast	atic castrate-resistant prostate cancer.				

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effects of mutagenesis from increased error rate? Recent evidence has shown improved radiographic progression free survival in patients with mCRPC treated with olaparib and abiraterone who were not selected for HRR alterations (38). In the potential setting of general benefit from PARP inhibition combinations in mCRPC, will HRR alterations select for better or longer responses? How can the relationships between androgen receptor signaling, the PI3K-AKT pathway and HRR deficiencies be further exploited with therapy (39)? Beyond specific mutations will BRCAness or BRCA-like genetic scars also correlate with response to platinum agents and PARPi as seen in ovarian cancer? Lastly the ability of malignancies to adapt to these therapies is becoming more apparent and offers an area ripe for further inquiry (40).

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Supplementary

Table S1 Foundation one testing of prior tumor sample				
Microsatellite status	MS-Stable			
Tumor mutational burden	8 Mut/Mb			
CDK12	L836fs*16			
FANCA	S251*			
FAS	Loss			
KEL	Splice site 348_400+158del211			
PALB2	L960fs*2			
PTEN	Loss			

Table S2 Integrated BRCA analysis Myriad myRisk genetic result

Gene	Mutation	Interpretation
PALB2	c.2878delC (p.Leu960Tyrfs*2) Heterozygous	High cancer risk
ATM	c.290T>C (p.lle97Thr)	Uncertain clinical significance
BRIP1	c.1057>C (p.Tyr353His)	Uncertain clinical significance
PMS2	c.2179C>g (p.Gln727Glu) (aka Q727E (2179C>G))	Uncertain clinical significance