Accumulation of periostin in acute exacerbation of familial idiopathic pulmonary fibrosis

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Abstract: Periostin, an extracellular matrix molecule, is associated with idiopathic pulmonary fibrosis (IPF). It is known that the frequency of familial IPF (FIPF) ranges from 0.5% to 2.2% among IPF cases. However, the relationship between periostin and FIPF has not been previously described. We report the first case of periostin accumulation in the lungs of a patient with an acute exacerbation of FIPF. A 72-year-old woman, diagnosed with FIPF, had been followed up for 5 years. The patient experienced increased dyspnea within a 1-month period and was referred to our hospital. The patient was hypoxic, and chest computed tomography showed rapidly expanding bilateral reticular shadows. Despite pulse-steroid and intravenous-cyclophosphamide therapy, the patient died 25 days after admission. On admission, serum periostin levels were not significantly elevated, while serum fibrotic marker levels were elevated. Immunohistochemical analysis of the lungs on autopsy showed marked accumulation of periostin in the active fibrotic lesions, whereas intact and burned-out areas did not show significant expression of periostin. This case might provide insight into the role of periostin in acute exacerbation of IPF.

Keywords: Familial idiopathic pulmonary fibrosis (FIPF); acute exacerbation (AE); idiopathic interstitial pneumonia; periostin

Submitted Feb 10, 2018. Accepted for publication Jun 10, 2018. doi: 10.21037/jtd.2018.06.96 **View this article at:** http://dx.doi.org/10.21037/jtd.2018.06.96

Introduction

Periostin, an extracellular matrix protein, has been identified as a novel biomarker of idiopathic interstitial pneumonia. Periostin is a mediator of chronic allergic diseases, and is involved in tissue remodeling observed in allergic inflammation in bronchial asthma, scleroderma, and atopic dermatitis (1). Periostin is highly expressed in idiopathic pulmonary fibrosis (IPF), and increased serum periostin correlates with the prognosis of IPF (2). Acute exacerbation (AE) of IPF (AE-IPF) is a life-threating event, and the prognosis of IPF is considerably dependent on the occurrence of AE (3). However, the association between periostin and AE-IPF has not been investigated. Herein, we have reported our findings in an autopsied patient with an AE of familial IPF (AE-FIPF) and revealed significant expression of periostin in the affected areas of the lungs. This is the first case report describing expression of periostin in AE-IPF.

Case presentation

In 201X, a 72-year-old woman was diagnosed with FIPF

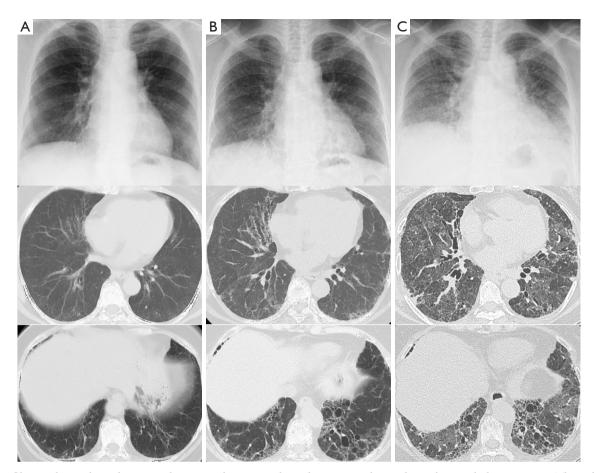


Figure 1 Chest radiographs and computed tomography images showed symmetrical reticular and ground glass opacities. The radiological features progressed slowly over 5 years from the age of 72 years (A: 72 years; B: 77 years), but then rapidly deteriorated over a month (B: 54 days and C: 25 days prior to death, respectively).

at a hospital; an older sister and younger brother had died due to similar disease conditions. There was no history of smoking. Pulmonary fibrosis slowly progressed over 5 years (*Figure 1A,B*). At 77 years of age, the patient was admitted to the same hospital for increased dyspnea of 1-month duration, and the patient was then referred to our hospital for intensive treatment.

On admission, the patient was hypoxic. Radiological examinations showed that bilateral reticular shadows rapidly progressed with traction bronchiectasis from 54 to 25 days prior to death (*Figure 1B,C*). Serum fibrotic markers were elevated (sialylated carbohydrate antigen KL-6: 2,332 U/mL, surfactant protein-D: 813 ng/mL, and surfactant protein-A: 201.6 ng/mL), while tests for autoimmune antibodies, except for autoantibody against myeloperoxidase anti-neutrophil

cytoplasmic antibody (MPO-ANCA: 13.8 U/mL), revealed negative results. Serum periostin was not markedly elevated at 108 ng/mL (upper limit: 95 ng/mL) using SS18A × 17B antibodies and 8.9 ng/mL (upper limit: 13 ng/mL) using SS20A × 19D antibodies (2). The cell count from bronchoalveolar lavage fluid was 1.6×10^5 /µL, and the differential was 48% neutrophils, 23% lymphocytes, 10% eosinophils, and 19% macrophages. The CD4/CD8 ratio was 3.44, and bacterial culture showed no growth. Intravenous cyclophosphamide and three times of pulsesteroid therapy were administered. Despite treatment, the patient developed respiratory failure and died 25 days after admission to our hospital.

Autopsy was performed at 2 hours postmortem. At autopsy, the lungs weighted 370 g (left) and 420 g (right), respectively. Histological examination revealed a diffuse

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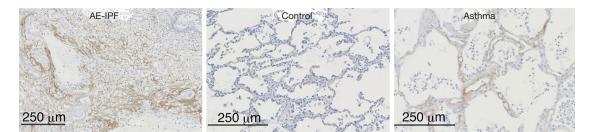


Figure 2 Immunohistochemistry of periostin in the lung. Periostin accumulation is seen in the present case (left panel), while no staining of periostin is observed in the control lung obtained from a patient with early lung cancer (middle panel). Positive staining of periostin is seen in the asthmatic lung obtained from a patient with early lung cancer complicated with bronchial asthma (right panel). AE, acute exacerbation; IPF, idiopathic pulmonary fibrosis.

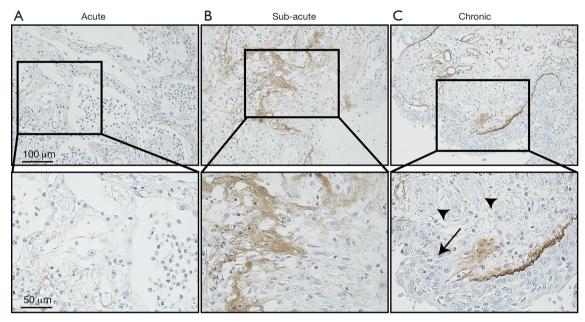


Figure 3 Periostin accumulation in sub-acute lesion after acute exacerbation of idiopathic pulmonary fibrosis. Periostin accumulation was observed in acute, sub-acute, and chronic inflammatory/fibrotic lesion of autopsy lung after acute exacerbation of idiopathic pulmonary fibrosis. Arrow indicates bronchiolar metaplasia and arrowheads indicates smooth muscle proliferation.

alveolar damage consistent with AE-IPF and a pattern of nonspecific interstitial pneumonia. Bacterial pneumonia was observed in the right upper lobe, and cytomegalovirus infection was identified in the esophageal and gastric mucosa, but not in the lungs. There was no evidence of granulomatous changes or polyangiitis indicating antineutrophil cytoplasmic antibody (ANCA)-associated vasculitis. To investigate the mechanism of AE-IPF, we performed immunohistochemical analysis using rat antihuman periostin monoclonal antibodies (clone no. SS19C). The antibodies were incubated with sections overnight at 4 °C. After washing with phosphate buffered saline, antigens were detected using an EnVisionTM horseradish peroxidase (3,3'-diaminobenzidine) system (Dako, Glostrup, Denmark) as described previously (4). Interestingly, immunohistochemical analysis showed accumulation of periostin in the fibrotic lesion of the lung parenchyma, which was greater than the periostin accumulation in control and asthmatic lung samples obtained from videoassisted thoracic surgery for early lung cancer (*Figure 2*). As shown in *Figure 3*, when comparing periostin accumulation among the acute, sub-acute, and chronic

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inflammatory/fibrotic phases in this case, periostin markedly accumulated in sub-acute fibrotic lesions developing in interstitial tissue with proliferation of fibroblasts, which was estimated approximately 7–10 days after acute exacerbation. Less accumulation of periostin was observed in the acute inflammatory phase with neutrophil invasion. The extent of periostin accumulation in the sub-acute fibrotic phase was stronger than that in the chronic phase of fibrosis with bronchiolar metaplasia and smooth muscle proliferation.

Discussion

Unlike in sporadic IPF, the typical honeycomb lung is not always observed in FIPF (5). FIPF and sporadic IPF are similar in various aspects such as results of pulmonary function test, pathologic changes, radiographic features, survival data, and treatment (6). A clinical biomarker for predicting prognosis in patients with AE-FIPF and sporadic IPF would be useful in the management of these diseases and in the treatment with anti-fibrotic agents.

Periostin combines with other extracellular matrix proteins to promote the formation of collagen fibers. It is associated with fibrotic diseases such as bronchial asthma, scleroderma, and atopic dermatitis (1,7). Periostin is highly expressed in lungs of patients with IPF, but is not expressed in patients with nonspecific interstitial pneumonia. Serum periostin levels are significantly elevated in patients with IPF when compared to healthy individuals (8). Recently, Ohta et al. reported that increased serum periostin is correlated with the prognosis of IPF (2). We measured the serum periostin level on admission and evaluated the expression of periostin in the lung obtained at autopsy by immunohistochemistry. In this case, periostin was highly expressed in lungs on autopsy, especially in the fibrotic interstitium and bronchiolar epithelium, while it was not expressed in the healthy lung tissue. On comparison of accumulation of periostin in the inflammatory/fibrotic acute, sub-acute, and chronic phases, accumulation of periostin was strong in the stroma tissue, consists of extracellular matrix. In this case, periostin may have accumulated in the fibrotic lesion in the sub-acute phase after AE-IPF.

Serum periostin levels were not clinically useful for predicting the prognosis of AE-FIPF in our case. SS18A × 17B periostin antibodies recognize inflammatory periostin produced in asthma and atopic dermatitis, while SS20A × 19D periostin antibodies specifically recognize fibrotic periostin produced by fibroblasts (2). While serum periostin may be a biomarker of IPF, it was not elevated in the patient with AE of IPF in this study. We speculated that serum periostin might be elevated at later phase of AE-IPF with increase in pulmonary accumulation. There may be a time lag between the production of periostin in the fibroblasts and its detection in blood circulation. Serum periostin may not be a sensitive biomarker for short-term changes in AE-IPF. However, it remains unclear if serum periostin level was elevated in the later phase of AE-IPF since it was not monitored during hospitalization in this case.

In summary, while serum periostin was not elevated on admission, accumulation of periostin was identified in the lungs at autopsy. Periostin might be related to the development of AE as well as chronic fibrosis. However, additional cases of FIPF or AE-IPF are required to elucidate the relationship of periostin with AE-IPF.

Acknowledgements

None.

Footnote

Conflicts of Interest: The authors have no conflicts of interest to declare.

Informed Consent: We were careful to not to identify the patient and other relevant family members when preparing the manuscript. We obtained written informed consent in Japanese from the spouse of the patient for publication.

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Cite this article as: Murata K, Koga Y, Kasahara N, Hachisu Y, Nunomura S, Nakajima N, Yokoo H, Kaira K, Maeno T, Dobashi K, Izuhara K, Hisada T. Accumulation of periostin in acute exacerbation of familial idiopathic pulmonary fibrosis. J Thorac Dis 2018;10(7):E587-E591. doi: 10.21037/jtd.2018.06.96

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