



EGFR T790M testing through repeated liquid biopsy over time: a real-world multicentric retrospective experience

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Background: About 15% of non-small cell lung cancers (NSCLCs) harbor epidermal growth factor receptor (*EGFR*) mutations. Upfront treatment with first and second generation *EGFR* tyrosine kinase inhibitors (1-2gen TKIs) is superior to chemotherapy. The most frequent resistance mechanism to 1-2gen TKIs is *EGFR* T790M mutation, which is targeted by osimertinib. T790M mutation can be revealed by liquid biopsy (LB) or by tissue rebiopsy (TB). LB is easily feasible but less sensitive than TB. We focused on repeated LBs and analyzed clinical features associated with *EGFR* T790M detection.

Methods: This is a retrospective multicenter observational study including *EGFR*-mutant NSCLC consecutive patients with disease progression (PD) after 1-2gen TKIs and with a first *EGFR* LB negative for T790M mutation, referred between 2016 and 2019. Aims of the study were to determine the prevalence of T790M mutation using LB in a real-life setting and the prevalence of T790M mutation by repeated LBs. We explored the association of T790M with clinical-pathological features and, through a survey, we evaluated the decision-making process behind LB request. Data on TBs were also collected.

Results: One hundred and ten patients were included in the study, for a total of 326 LBs. Median number of LB per patient was 3.0. The T790M prevalence through LB was 34.5%. Over time, significantly more LBs were requested “at clinical and radiological PD” and “at radiological PD” compared to “arbitrarily”. The probability of finding the T790M mutation for a patient across each subsequent LB did not significantly change. Liver and lymph node PD were significantly correlated to T790M positivity. Notably, “at PD” compared to “arbitrarily” LB request and liver, bone or lymph node PD were correlated to the detection of any *EGFR* mutation in cfDNA. TB was performed in 59.7% of patients with a T790M negative LB and 18.8% of them were T790M positive. In most cases, TB was not feasible due to anatomical reasons. In our study population, the overall T790M prevalence—detected with both LB and TB—was 42.7%.

Conclusions: Repeated LB testing can be useful in a real-life scenario to detect *EGFR* T790M mutation.

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Introduction

Lung cancer is the leading cause of cancer-related death and non-small cell lung cancer (NSCLC) represents about 85% of lung cancers (1). About 15% of patients with advanced NSCLC harbor epidermal growth factor receptor (*EGFR*) mutations, the most common being exon 19 deletion and the exon 21 L858R substitution (2). *EGFR* mutation testing is recommended at diagnosis for patients with lung adenocarcinoma and for light smokers regardless of histology, considering the survival benefit shown by *EGFR* tyrosine kinase inhibitors (TKIs) over chemotherapy for *EGFR*-positive patients (3-6).

EGFR exon 20 T790M mutation constitutes the predominant acquired resistance mechanism to first and second generation TKIs (7-10). Osimertinib is a third generation TKI that targets activating mutations and the T790M resistance mutation. This compound irreversibly binds to the C797 residue in the *EGFR* mutant protein ATP-binding pocket, sparing wild-type *EGFR* (11). At disease progression after first or second generation TKIs treatment, the analysis of circulating tumor DNA (ctDNA) through liquid biopsy (LB) or tissue rebiopsy (TB), when feasible, is carried out to identify T790M mutation and, whenever present, switch to osimertinib (6,11).

Recently, osimertinib has become the standard of care first line treatment for *EGFR*-positive patients (12,13). Notably, resistance mechanisms to first line osimertinib do not include T790M mutation (14-16). Nonetheless, characterizing tumor genomics after progression to TKIs remains crucial, in order to find druggable alterations and personalize further treatments. After osimertinib failure, extensive ctDNA analysis through next generation sequencing (NGS) could be performed through liquid and tissue biopsy to evaluate resistance mechanisms (14-16). No targeted treatments are currently approved by European Medicines Agency for upfront osimertinib acquired resistance, although several clinical trials are ongoing, especially in the setting of *MET*-amplified patients (17-20).

Compared to tissue biopsy, LB is a minimally invasive technique that can better picture tumor heterogeneity, be safely repeated over time, and overcome feasibility issues (21,22). However, tissue biopsy remains the only technique providing histological information and the gold standard for re-assessing mutational status (23). Since LB sensitivity is 50–90%, a negative LB result does not exclude the presence of a molecular alteration, but might be due to the lack of ctDNA or to presence of mutant allele at lower frequency than the limit of detection (24-28). Extrathoracic disease and bone metastases have been correlated with a higher probability to detect *EGFR* T790M mutation on ctDNA (28,29), while detection rates of genomic alterations is lower in patients with exclusively intracranial disease (30).

To date, it is unknown whether a specific timing of testing or clinical features could enhance LB sensitivity, whether a resistance mutation could be found in ctDNA before radiological progression, and whether repeating LBs after a negative result would be clinically meaningful (31,32). Therefore, optimizing LB procedures and ctDNA workflow is still an unmet clinical need (33). The clinical data derived from the use of LB to detect T790M mutation can help fill these knowledge gaps and may be translated into other clinical scenarios in which the analytic methods rely on ctDNA, such as NGS after progression to first line osimertinib (14-20).

The primary aims of this study were the description of the prevalence of *EGFR* T790M mutation at LB and in repeated LBs over time in a real-life cohort of *EGFR* mutant advanced NSCLC patients progressing to upfront first or second generation *EGFR* TKIs. Secondary aims were to assess the correlation of *EGFR* activating mutations and T790M mutation at LB with clinical-pathological features and the prevalence of *EGFR* T790M mutation in TB. Finally, we described the decision-making process for LB request and for TB. We present the following article in accordance with the STROBE reporting checklist (available at <https://jtd.amegroups.com/article/view/10.21037/jtd-22-745/rc>).

Methods

We designed an observational retrospective study at two Italian Oncology Centers. The inclusion criteria were histologically confirmed diagnosis of advanced NSCLC between 2016 and 2019, the presence of *EGFR* activating mutation at diagnosis, upfront treatment with first or second generation *EGFR* TKIs (erlotinib, gefitinib, or afatinib) and the presence of at least two LBs after starting first or second generation *EGFR* TKI, with the first LB resulting as T790M negative. Patients with T790M mutation at diagnosis were excluded, as patients with only one LB test.

This study was conducted according to current rules of Good Clinical Practice and the principles of Helsinki Declaration (as revised in 2013). Informed consent form for diagnostic procedures and treatment was signed by all patients, according to clinical practice and to the Regulation EU 2016/679 of the European Parliament and of the Council on personal data protection. Patients' data collected in the study were anonymized. Ethics Committee approval by our Institutions was not required because of the observational intent of the data collection from the diagnostic routine practice.

An electronic questionnaire was submitted to medical oncologists and biologists at each involved Oncology Center [Supplementary file ([Appendix 1](#))]. The survey was distributed in November 2020 and was collected in January 2021. The following clinical data were collected for each LB: date and result of LB, reason for LB request, progression type, progression sites, subsequent systemic therapy after LB; tissue biopsy after LB, tissue biopsy site and result, and, if tissue biopsy was not done, the reason for not performing it.

The possible pre-set reasons for LB request included “at radiological progression”, “at clinical and radiological progression” or “arbitrarily”. Progression type was classified as “only radiological and not meeting Response Evaluation Criteria version 1.1 (RECIST 1.1) for progression”, “only radiological and meeting RECIST 1.1 criteria for progression”, and “both clinical and radiological progression”. Progression sites included central nervous system, intrathoracic, liver, adrenal gland, metastatic lymph nodes and bone. Subsequent therapies were classified as TKI, chemotherapy, immunotherapy, or best supportive care. The possible pre-set reasons for which tissue biopsy was not carried out included “T790M positive liquid biopsy

outcome”, “tissue biopsy not feasible”, “patient's decision”, “only radiological progression”, or “clinical deterioration”.

EGFR mutational analysis

At the time of histologic diagnosis, mutational analysis of *EGFR* gene exons 18 to 21 was performed according to standard clinical practice.

At disease progression, LB was performed to assess the *EGFR* T790M mutational status in ctDNA. In T790M negative patients at LB, a tissue biopsy was carried out, when feasible. If test was still T790M negative, new plasma samples were collected.

In case of LB, cell free-DNA was isolated from 2 mL of plasma using the cobas[®] cf-DNA Sample Preparation kit (Roche, Basel, Switzerland) or the Helix[®] circulating Nucleic Acid assay (Diatech Pharmacogenetics, Jesi, Italy), and the *EGFR* mutational analysis was performed by polymerase chain reaction (PCR)-based methods (cobas[®] *EGFR* Mutation Test v2, Roche, Basel, Switzerland, or Easy *EGFR* kit, Diatech Pharmacogenetics, Jesi, Italy).

In case of tissue biopsy, DNA was extracted from formalin-fixed paraffin-embedded tissue sections using the QIAamp DNA Mini kit (Qiagen, Hilden, Germany) and analyzed by mass spectrometry-based methods (Sequenom MassARRAY, Diatech Pharmacogenetics, Jesi, Italy).

Results of liquid and tissue biopsies were classified as: positive (when *EGFR* T790M mutation was present); negative (when neither T790M nor activating mutations were found); other (when only *EGFR* activating mutations but not T790M were reported).

Treatment and tumor assessment

First-line systemic treatment was administered according to national and international guidelines. Patients received a first (oral erlotinib 150 mg daily or gefitinib 250 mg daily) or second generation TKI (oral afatinib 40 mg daily) until radiological disease progression (PD), clinical treatment failure, unacceptable toxicity, physician and/or patient decision, or death.

Tumor assessment was performed through chest and abdomen computerized tomography (CT) scan; brain CT scan or nuclear magnetic resonance was performed as clinically indicated. Response to treatment was classified according to RECIST, version 1.1.

Statistical analysis

Statistical analysis was performed through R software (version 4.0.5; the R foundation, Vienna, Austria). Mean and standard deviation, median, first (Q1) and third (Q3) quartile were reported for quantitative variables. The Cochran-Mantel-Haenszel test for monotonic trend was used to evaluate statistically significant changes in the reasons for LB request or in the systemic therapy, over time or through repeated LBs, and changes in conditional probability of T790M mutation over time. Associations of *EGFR* T790M positivity by LB and any *EGFR* mutation positivity by LB with clinical factors were evaluated through logistic and mixed effects logistic models, respectively. Odds

ratios (ORs) with 95% confidence intervals (95% CIs) were reported. Significance level was set at $P < 0.05$.

Results

Study population and repeated LBs

Between January 2016 and December 2019, 110 consecutive patients with progression to a first or second generation *EGFR* TKI and a first T790M negative LB were included in this study. A second LB was performed in all patients. Data cut-off was carried out on 5 January 2021 (Figure 1).

Twenty-four patients out of 110 (21.8%) were enrolled in 2016, 25 (22.7%) in 2017, 29 (26.4%) in 2018 and 32 (29.1%) in 2019.

Overall, 326 LBs were carried out (Figure 1). Twenty-nine (8.9%) out of 326 were collected in 2016, 74 (22.7%) in 2017, 72 (35.7%) in 2018, 110 (33.7%) in 2019 and 41 (12.6%) in 2020.

The mean number of LBs per patient was 3.0 (standard deviation: 1.2). The median number of LBs per patient was 3 (Q1: 2; Q3: 6). The median time between subsequent blood draws was 124 days (Q1: 70 days; Q3: 230 days). The median time between each subsequent LB is detailed in Table 1.

LBs results and prevalence of *EGFR* T790M mutation

Seventy-seven out of 110 patients (70.0%) showed any *EGFR* mutation at LB. The T790M mutation was detected through LB in 38 of 110 patients (34.5%); in 22 of 38 cases both T790M and baseline *EGFR* activating mutation were present, whereas 16 of 38 cases were T790M positive and baseline *EGFR* activating mutation negative. Thirty-nine of 110 patients (35.5%) displayed only the baseline *EGFR* activating mutations and were

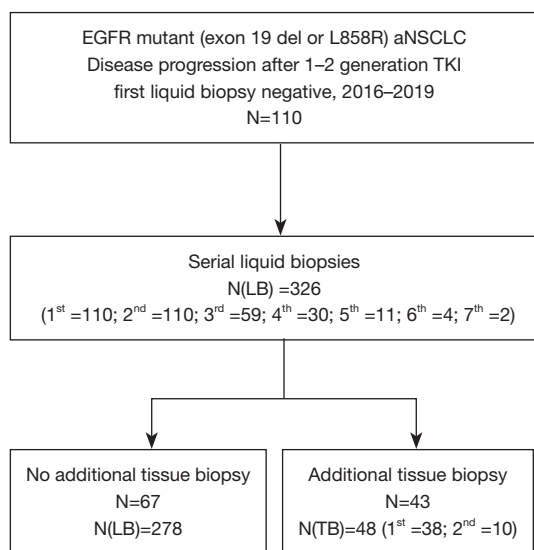


Figure 1 Study design. EGFR, epidermal growth factor receptor; Del, deletion; aNSCLC, advanced non-small cell lung cancer; TKI, tyrosine kinase inhibitor; N, number of patients; N(LB), number of liquid biopsies; N(TB), number of tissue biopsies.

Table 1 Timing of serial liquid biopsies

Number of repeated liquid biopsy per patient	First	Second	Third	Fourth	Fifth	Sixth	Seventh	Total
Number of liquid biopsies	110	110	59	30	11	4	2	326
Time from previous liquid biopsy (days)								
Median	–	119	128	137	104	90	116	124
Q1, Q3	–	70, 197	66, 291	82, 242	88, 152	61, 118	72, 159	70, 230
Range	–	3–784	14–892	21–423	30–386	28–146	28–203	3–892

Q1, first quartile; Q3, third quartile.

Table 2 Reasons for liquid biopsy request over time

Year of liquid biopsy	2016	2017	2018	2019	2020	Total	P value (trend test)
Reason for liquid biopsy request, n (%)							0.0114
At clinical and radiological progression	2 (6.9)	13 (17.6)	18 (25.0)	20 (18.3)	12 (30.8)	65 (20.1)	
At radiological progression	15 (51.7)	47 (63.5)	40 (55.6)	73 (67.0)	22 (56.4)	197 (61.0)	
Arbitrarily	12 (41.4)	14 (18.9)	14 (19.4)	16 (14.7)	5 (12.8)	61 (18.9)	
Missing	0	0	0	1	2	3	
Total ^a	29	74	72	110	41	326	

^a, percent values were calculated excluding missing data.

Table 3 Reasons for liquid biopsy request through sequential liquid biopsies

Number of repeated liquid biopsy	First	Second	Third	Fourth	Fifth	Sixth ^b	Seventh ^b	Total	P value (trend test)
Reason for liquid biopsy request, n (%)									0.0651
At clinical and radiological progression	17 (15.5)	23 (21.1)	14 (23.7)	6 (21.4)	3 (27.3)	1 (25.0)	1 (50.0)	65 (20.1)	
At radiological progression	80 (72.7)	60 (55.0)	34 (57.6)	16 (57.2)	5 (45.4)	2 (50.0)	0 (0.0)	197 (61.0)	
Arbitrarily	13 (11.8)	26 (23.9)	11 (18.7)	6 (21.4)	3 (27.3)	1 (25.0)	1 (50.0)	61 (18.9)	
Missing	0	1	0	2	0	0	0	3	
Total ^a	110	110	59	30	11	4	2	326	

^a, percent values were calculated excluding missing data; ^b, sixth and seventh liquid biopsies were excluded from the trend evaluation because of limited numerosity.

T790M negative. Finally, thirty-three patients out of 110 (30.0%) had a negative LB.

Reasons for LB testing over time and through LBs

We reported the reason for requesting each LB. A significant reduction in the “arbitrarily” reason, compared to “at clinical and radiological progression” and “at radiological progression”, was shown over time ($P=0.0114$) (Table 2). The “arbitrarily” motivation increased through subsequent biopsies, although not significantly (Table 3).

We collected information about the treatment that patients received after each LB (Figure 2). Forty-four patients out of 110 (40%) changed systemic therapy, whereas 66 (60%) patients did not. A significant increase in the probability of administering chemotherapy after LB compared to other therapies was found throughout subsequent LBs ($P<0.001$) (Table S1).

Prevalence of EGFR T790M mutation throughout LBs

We analyzed the prevalence of T790M mutation across each subsequent LB (Table 4). The probability of finding the T790M mutation for a patient across subsequent LBs did not significantly change (Table 4).

Features associated with EGFR T790M positivity and ctDNA positivity

We evaluated the association between EGFR T790M positivity at LB and the reason for LB request. A trend was shown for association of T790M positivity when “clinical and radiological progression” motivation was present, compared to “radiological progression” and “arbitrarily” reason (Figure 3, Table S2). Conversely, liver and lymph node progression were significantly correlated to T790M positivity (OR 4.13, 95% CI: 1.67–10.2, $P=0.002$; and OR 4.2, 95% CI: 1.45–12.1, $P=0.008$, respectively) (Figure 3,

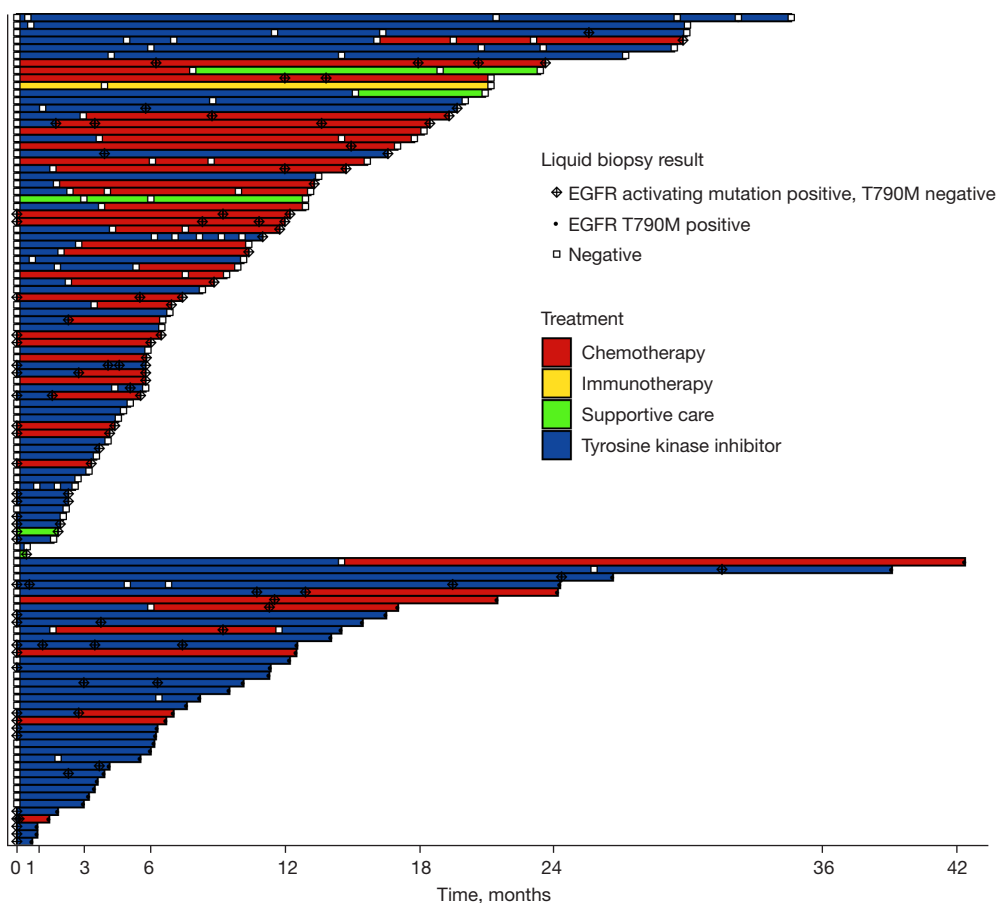


Figure 2 Swimmer plot of *EGFR* T790M negative and T790M positive patients. Each row represents a single patient. Points represent the liquid biopsy result (black bullet, *EGFR* T790M mutation positive; ringed cross, *EGFR* activating mutation positive, T790M negative; white square: *EGFR* activating mutation and T790M negative). Bars represent the treatment ongoing after the liquid biopsy (red: chemotherapy; yellow: immunotherapy; green: supportive care; blue: tyrosine kinase inhibitor). *EGFR*, epidermal growth factor receptor.

Table 4 Prevalence of *EGFR* T790M mutation through repeated liquid biopsies

Number of repeated liquid biopsy	First ^a	Second	Third	Fourth	Fifth	Sixth	Seventh ^a	Total	P value (trend test)
T790M, n (%)									0.7164
Negative	110 (100.0)	89 (80.9)	49 (83.1)	26 (86.7)	9 (81.8)	3 (75.0)	2 (100.0)	288 (88.3)	
Positive	0 (0.0)	21 (19.1)	10 (16.9)	4 (13.3)	2 (18.2)	1 (25.0)	0 (0.0)	38 (11.7)	
Total	110	110	59	30	11	4	2	326	

^a, first and seventh liquid biopsies were excluded from the trend evaluation. *EGFR*, epidermal growth factor receptor.

Table S2).

Moreover, the association between LB positivity for any *EGFR* mutation (i.e., T790M or other *EGFR* mutations) and the reason prompting LB were evaluated. Compared to “arbitrarily” reason, the reason “at clinical and radiological

progression” was significantly associated to increased odds of finding any mutation (OR 6.52, 95% CI: 2.13–19.9, P=0.001) (Figure 4, Table S2). When the metastatic disease spread to the liver, bone or lymph nodes, the probability of detecting any mutation at LB was significantly higher

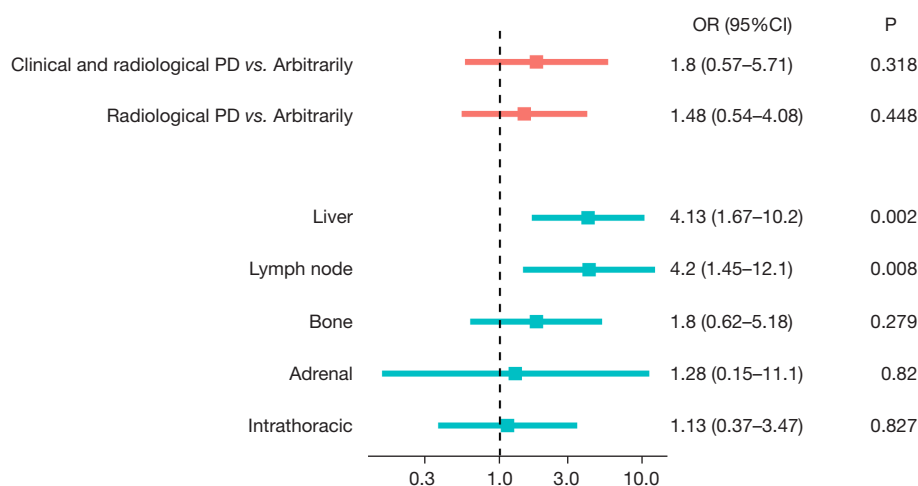


Figure 3 Features associated with *EGFR* T790M positivity at liquid biopsy. Squares represent odds ratios and bars 95% confidence intervals for the odds ratios. Features positively associated with *EGFR* T790M mutation show an odds ratio >1, whereas features negatively associated an odds ratio <1. PD, progression; OR, odds ratio; 95% CI, 95% confidence interval; *EGFR*, epidermal growth factor receptor.

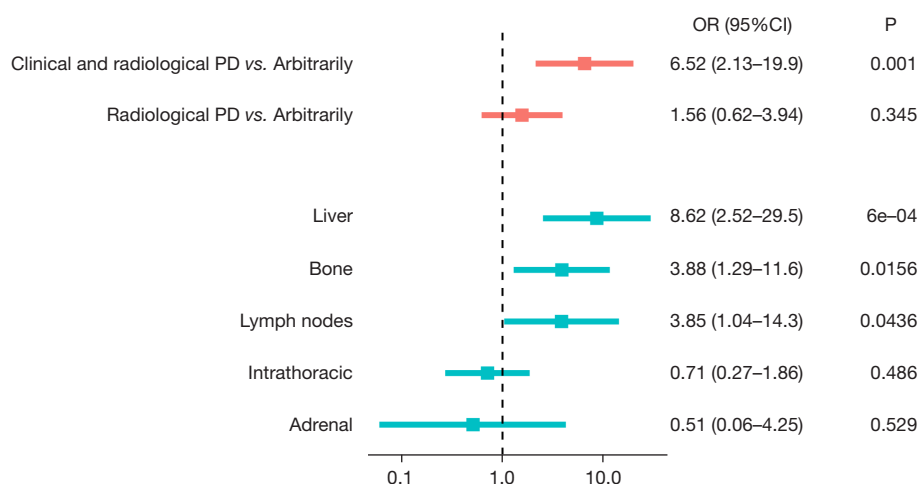


Figure 4 Features associated with *EGFR* liquid biopsy positivity (any mutation). Squares represent odds ratios and bars 95% confidence intervals for the odds ratios. Features positively associated with *EGFR* liquid biopsy positivity (any mutation) show an odds ratio >1, whereas features negatively associated an odds ratio <1. PD, progression; OR, odds ratio; 95% CI, 95% confidence interval; *EGFR*, epidermal growth factor receptor.

(OR 8.62, 95% CI: 2.52–29.5, $P=0.0006$; OR 3.88, 95% CI: 1.29–11.6, $P=0.0156$, and OR 3.85, 95% CI: 1.04–14.3, $P=0.0436$, respectively) (Figure 4, Table S2).

TB as a salvage procedure

When LB was T790M negative and the procedure was feasible, TBs were performed in 43 out of 72 patients

(59.7%) (Figure 1). Thirty-eight out of 43 (88.4%) patients had one tissue biopsy, while 5 (11.6%) patients had two tissue biopsies, for a total of 48 procedures. The site of tissue biopsy was intrathoracic in 40 out of 48 (83.3%) biopsies, liver in 4 cases (8.3%), lymph nodes in 3 cases (6.2%) and breast in 1 case (2.1%). Data on biopsy procedure was available for 47 cases: in 22 out of 47 cases (46.8%) the procedure was CT or ultrasound-guided, in 20

(42.5%) was in bronchoscopy, in 5 (10.7%) in thoracoscopy. Nine out of 48 tissue biopsies (18.8%) were both *EGFR* T790M and activating mutation positive, 16 (33.3%) were positive for *EGFR* activating mutation and T790M negative, and 23 (47.9%) were negative. The prevalence of T790M at tissue biopsy in our study was 20.9% (9 out of 43 patients).

Notably, in most patients (67 out of 110, 60.9%), tissue biopsy was not carried out. For two hundred seventy-three out of 278 LBs we recorded the reason for which tissue biopsy was not done: in 135 out of 273 blood draws (49.5%) there was no clinical progression, in 67 cases (24.6%) tissue biopsy was not feasible due to the biopsy site, in 38 cases (13.9%) LB was T790M positive, in 22 cases (8.0%) the patient's conditions worsened and for patient refusal in 11 cases (4.0%).

The overall T790M prevalence in the study population by liquid and tissue biopsy was 47/110 (42.7%).

Discussion

This real-world, multicenter study evaluated the use of *EGFR* LB for detecting resistance mechanisms in a cohort of patients with *EGFR* positive advanced NSCLC, after progression to first or second generation *EGFR* TKI. The impact of LB on the diagnostic-therapeutic pathway was also described.

Randomized clinical trials and several real-world studies demonstrated osimertinib efficacy in second and in further lines in *EGFR* T790M mutation positive patients (11,34-39). Thus, international guidelines include *EGFR* mutation testing through LB at disease progression after first or second generation TKIs treatment to find T790M acquired mutation (6). When LB is T790M negative, tissue biopsy is recommended to identify T790M mutation and, if present, switch to osimertinib (6). However, TB is frequently not feasible due to technical issues or worsening conditions of the patient (40-42). In this case, a common clinical strategy is to repeat LB to seek for T790M mutation. Few real-world studies described this medical oncologist attitude (43-46) and, to the best of our knowledge, this is the first work to specifically focus on this topic in a large patient population, and to include only patients with repeated *EGFR* testing at LB and a first T790M negative LB.

Among the real-world experiences, a work by Spence *et al.* included 343 sequential patients with progression after first of second generation TKI, of which 82 patients were T790M positive at initial LB testing. Eighty-four patients received additional follow-up testing. Fifty-one patients

had serial blood specimens (a total of 59 LBs), 34 patients TB (for a total of 35 procedures) and 16 patients cytology testing (18 specimens in total). Eight of 59 (13.6%) repeated LBs were T790M positive, 15 of 35 (42.9%) tissue biopsies and 7 of 18 (38.8%) cytology specimens (43).

The work by Reale and colleagues reported data on 308 consecutive patients with progression to upfront first or second generation TKI, of which 169 underwent a LB and 66 (39%) were T790M positive. Forty-eight patients underwent further testing with liquid or tissue biopsy, 24 (50%) of these patients were T790M positive (44).

A third study by Minari *et al.* included 120 consecutive patients with progression to upfront first or second generation TKI, 23 (19.2%) of which tested T790M positive at first LB. Forty patients underwent a second LB with 5 (12.5%) positive samples for T790M. Twelve patients underwent a third LB, with 3 (25.0%) patients positive for T790M and, finally, two patients underwent a fourth LB with no T790M positive samples. Fifty-four (60.7%) of 89 T790M negative patients underwent TB, with 28 (56%) patients positive for T790M (45).

Finally, Silveira and colleagues evaluated 111 consecutive patients, with 40 (36.0%) patients T790M positive at LB. Eighteen T790M negative patients received further testing: 6 patients had another LB, with the detection of T790M in 3 cases (50%), while 12 had tissue biopsy, with 2 T790M positive cases (16.7%) (46).

The prevalence of *EGFR* T790M mutation using LB in our cohort was 34.5%, consistently with other real-life studies where prevalence ranged from 24% to 52% (43-46). This value is less than the overall proportion of T790M mutation as a resistance mechanism (about 50%), as expected, due to the limited sensitivity of LB compared to tissue biopsy.

Furthermore, we described the decision-making process prompting LB request. The most reported reasons for testing were "at radiological progression" and "at radiological and clinical progression", compared to "arbitrarily". We showed a significantly reduction of the "arbitrarily" reason over time. This evolution over time can be explained by the improvement of clinicians' learning curve about LB and increasing data showing better sensitivity for LB in patients with high disease burden (28,29,47).

Interestingly, we showed that the prevalence of *EGFR* T790M mutation across repeated LBs did not significantly change over time. This original finding and the observation that osimertinib is effective also beyond second line (34-38)

suggest the utility of repeated LB in T790M negative patients at every disease progression event in order to detect T790M mutation.

Moreover, we explored the correlation of *EGFR* mutations and T790M mutation at LB with clinical and pathological features. The features associated with the presence of any *EGFR* mutation at LB, an estimate of ctDNA “shedding” status, was clinical and radiological progression, or, similarly, the presence of liver, bone or nodal metastases. These factors correlate to disease burden and spread. Conversely, only liver and lymph node progression were associated to T790M positivity. These results are coherent with other published works (28,29,43,47).

We finally collected data on TBs. TB was carried out in 59.7% of T790M negative patients. The most frequent reason accounting for no TB was technical feasibility, followed by the patient's clinical deterioration. Other real-life experiences reported similar or lower proportions of patients undergoing TB and the same issues we found for not carrying out the procedure (40,41,45,46). In our study, the prevalence of *EGFR* T790M mutation in TB was 20.9%. This relatively low value could possibly be explained by tumor spatial heterogeneity, which could be overcome by LB.

Last, the overall prevalence of T790M mutation in our study, considering both liquid and tissue biopsy, was 42.7%, which is in line with previous studies (9,10).

The main limitations of our work are its retrospective nature and the relatively small number of patients with data on TBs.

Future research should aim to prospectively clarify the best timing, strategies and factors driving serial LB testing, in order to optimize the diagnostic yield and the clinical utility of this test. These findings could be translated to the management of patients receiving upfront osimertinib and experiencing acquired resistance, monitored through serial NGS, and, similarly, to patients receiving other targeted treatments.

Conclusions

Repeated *EGFR* LB mutational analysis can be used in a real-life scenario to detect *EGFR* T790M mutation acquisition.

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Footnote

Reporting Checklist: The authors have completed the STROBE reporting checklist. Available at <https://jtd.amegroups.com/article/view/10.21037/jtd-22-745/rc>

Data Sharing Statement: Available at <https://jtd.amegroups.com/article/view/10.21037/jtd-22-745/dss>

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <https://jtd.amegroups.com/article/view/10.21037/jtd-22-745/coif>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study complies with the Declaration of Helsinki (as revised in 2013). Ethics Committee approval by our Institutions was not required because of the observational intent of the data collection from the diagnostic routine practice. Informed consent form for diagnostic procedures and treatment was signed by all patients, according to clinical practice and to the Regulation EU 2016/679 of the European Parliament and of the Council on personal data protection.

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Appendix 1 Questionnaire

1. Liquid biopsy number
2. Liquid biopsy date
3. Liquid biopsy result
 - a. EGFR T790M positive
 - b. EGFR activating mutation positive, T790M negative
 - c. Negative
4. Motivation for liquid biopsy
 - a. At clinical and radiological progression
 - b. At radiological progression
 - c. Arbitrarily
5. In case of progression, specify progression type
 - a. Only radiological progression and not meeting RECIST criteria
 - b. Only radiological progression and meeting RECIST criteria
 - c. Both clinical and radiological progression
6. Specify progression sites
 - a. central nervous system
 - b. intrathoracic
 - c. liver
 - d. adrenal gland
 - e. metastatic lymph nodes
 - f. bone
7. Systemic therapy after liquid biopsy
 - a. TKI
 - b. Chemotherapy
 - c. Immunotherapy
 - d. Supportive care
8. Has a tissue rebiopsy been performed after liquid biopsy?
 - a. Yes
 - b. No
9. Tissue biopsy site (specify)
10. Type of procedure
 - a. CT or ultrasound-guided
 - b. Bronchoscopy
 - c. Thoracoscopy.
11. Tissue biopsy result
 - a. EGFR T790M positive
 - b. EGFR activating mutation positive, T790M negative
 - c. Negative
12. If tissue biopsy was not carried out, specify the reason
 - a. positive liquid biopsy
 - b. tissue biopsy not feasible
 - c. patient's decision
 - d. only radiological progression
 - e. clinical deterioration

Table S1 Systemic therapy following liquid biopsy

Number of repeated liquid biopsy	First	Second	Third	Fourth	Fifth	Sixth ^a	Seventh ^a	Total	P value (trend test)
Treatment									<0.001
Chemotherapy	9 (8.2%)	19 (17.3%)	30 (50.8%)	14 (46.7%)	4 (36.4%)	1 (25.0%)	1 (50.0%)	78 (23.9%)	
Immunotherapy	1 (0.9%)	1 (0.9%)	1 (1.7%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	3 (0.9%)	
Supportive therapy	0 (0.0%)	3 (2.7%)	3 (5.1%)	2 (6.6%)	0 (0.0%)	0 (0.0%)	0 (0.0%)	8 (2.5%)	
TKI	100 (90.9%)	87 (79.1%)	25 (42.4%)	14 (46.7%)	7 (63.6%)	3 (75.0%)	1 (50.0%)	237 (72.7%)	
Total	110	110	59	30	11	4	2	326	

^a, sixth and seventh liquid biopsies were excluded from the trend evaluation because of limited numerosity. TKI, tyrosine kinase inhibitor.

Table S2 Features associated with *EGFR* T790M mutation or other *EGFR* mutations

		<i>EGFR</i> T790M mutation	Other <i>EGFR</i> mutations ^a	Negative	Total
Reason for liquid biopsy request	At clinical and radiological progression	9 (24.3%)	32 (30.5%)	24 (13.3%)	65 (20.1%)
	At radiological progression	23 (62.2%)	58 (55.2%)	116 (64.1%)	197 (61.0%)
	Arbitrarily	5 (13.5%)	15 (14.3%)	41 (22.7%)	61 (18.9%)
	Missing	1	2	0	3
Type of disease progression	Radiological progression according to RECIST criteria	16 (50.0%)	29 (32.2%)	30 (21.4%)	75 (28.6%)
	Clinical and radiological progression	8 (25.0%)	30 (33.3%)	24 (17.1%)	62 (23.7%)
	Radiological progression (non-RECIST)	8 (25.0%)	31 (34.4%)	86 (61.4%)	125 (47.7%)
	Missing	1	2	0	2
Progression site	Intracranic and extracranic	3 (9.4%)	6 (6.7%)	15 (10.7%)	24 (9.2%)
	Extracranic only	25 (78.1%)	79 (87.8%)	114 (81.4%)	218 (83.2%)
	Intracranic only	4 (12.5%)	5 (5.6%)	11 (7.9%)	20 (7.6%)
	Missing	1	2	0	3
Site of extracranic progression	Intrathoracic	24 (85.7%)	69 (81.2%)	111 (86.0%)	204 (84.3%)
	Liver	9 (32.1%)	16 (18.8%)	6 (4.7%)	31 (12.8%)
	Adrenal	1 (3.6%)	2 (2.4%)	4 (3.1%)	7 (2.9%)
	Lymph node	6 (21.4%)	8 (9.5%)	5 (3.9%)	19 (7.9%)
	Bone	5 (17.9%)	15 (17.9%)	8 (6.2%)	28 (11.6%)
Treatment	Chemotherapy	7 (18.9%)	40 (38.1%)	30 (16.6%)	77 (23.8%)
	Immunotherapy	0 (0.0%)	1 (1.0%)	2 (1.1%)	3 (0.9%)
	Supportive therapy	0 (0.0%)	2 (1.9%)	6 (3.3%)	8 (2.5%)
	TKI	30 (81.1%)	62 (59.0%)	143 (79.0%)	235 (72.8%)
	Missing	1	2	0	3
Tissue biopsy	No	38 (100.0%)	87 (81.3%)	153 (84.5%)	278 (85.3%)
	Yes	0 (0.0%)	20 (18.7%)	28 (15.5%)	48 (14.7%)
<i>EGFR</i> mutation analysis at tissue biopsy	Negative/not informative	–	12 (60.0%)	11 (39.3%)	23 (47.9%)
	T790M+	–	1 (5.0%)	8 (28.6%)	9 (18.8%)
	Other ^a	–	7 (35.0%)	9 (32.1%)	16 (33.3%)
Total		38	107	181	326

^a, other *EGFR* mutation: *EGFR* T790M mutation negative, activating mutation positive. *EGFR*, epidermal growth factor receptor; TKI, tyrosine kinase inhibitor; RECIST, Response Evaluation Criteria in Solid Tumors.