



# Etiological diagnostic performance of probe capture-based targeted next-generation sequencing in bloodstream infection

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**Background:** A rapid and precise etiological diagnosis is crucial for the effective treatment of bloodstream infection (BSI). In this study, the performance of probe capture-based targeted next-generation sequencing (tNGS) was compared to that of blood culture and metagenomic next-generation sequencing (mNGS) in detecting potential pathogens in patients with BSI.

**Methods:** A total of 80 patients with suspected BSI were prospectively enrolled from 24 November 2023 to 30 December 2023 at Zhongshan Hospital, Shanghai, China. All 80 participants underwent simultaneous blood culture, blood mNGS, and blood tNGS after admission when febrile, and the results were compared.

**Results:** Among the 80 participants, 11 were clinically diagnosed with noninfectious fever, and 69 were diagnosed with BSI. Blood tNGS had a higher sensitivity for the diagnosis of BSI than blood culture (91.3% *vs.* 23.2%,  $P < 0.001$ ) and blood mNGS (91.3% *vs.* 69.6%,  $P = 0.001$ ). There was no significant difference in specificity between blood mNGS and tNGS (81.8% *vs.* 100.0%,  $P = 0.13$ ). Blood tNGS demonstrated a faster turnaround time than blood culture and blood mNGS. In 22 (31.9%) patients with BSI, targeted adjustment of the anti-infectious therapy according to the blood tNGS results resulted in clinical improvement.

**Conclusions:** Blood tNGS may be a promising tool for detecting potential pathogens in patients with BSI. The application of blood tNGS for BSI could guide anti-infectious treatment strategies and might improve clinical outcomes.

**Keywords:** Probe capture-based targeted next-generation sequencing (probe capture-based tNGS); metagenomic next-generation sequencing (mNGS); blood culture; bloodstream infection (BSI); diagnostic

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## Introduction

Bloodstream infection (BSI) is a life-threatening disease associated with high mortality rates (1). Appropriate antibiotic treatment is recommended for BSI at the earliest

opportunity (2,3). Delayed diagnosis and ineffective treatment may lead to septic shock, multiple organ failure, disseminated intravascular coagulation (DIC), and even death. Thus, rapid and precise etiological diagnosis is crucial

for the optimal treatment and prognosis of BSI. Blood culture is the most widely used conventional etiological diagnostic method (4). However, the sensitivity of blood cultures is far from ideal. Just over one-third of the patients with sepsis have positive blood cultures, mainly because of inadequate sampling volumes and prior administration of antibiotics (5). Blood culture is also time-consuming, usually taking 3–7 days or longer to identify potential pathogens, which is not feasible in clinical practice.

In recent years, culture-independent methods such as polymerase chain reaction (PCR) and metagenomic next-generation sequencing (mNGS) have been used for the etiological diagnosis of BSI (6–8). Regarding PCR, a presumptive diagnosis is necessary and misdiagnosis is common. The sensitivity of mNGS varies depending on the specimen type and disease severity (6–8). On account of the large amount of host human DNA and low biomass of pathogens, the positive rate of mNGS in serum samples is still not ideal. In addition, mNGS is expensive, limiting its clinical application. In Shanghai, the cost of a single mNGS test is around 3,500 RMB. If mNGS covers DNA and RNA dual processes, the price will increase to 4,500–6,500 RMB. Therefore, a more sensitive, rapid, and cost-efficient method for diagnosing BSI is urgently needed.

Target enrichment strategies are commonly used in NGS workflows to eliminate genomic DNA regions that are not of interest (9–11). Probe capture-based targeted next-generation sequencing (tNGS) has been applied for the molecular diagnosis of human infectious diseases (12,13). However, the application of probe capture-based tNGS for the etiological diagnosis of BSI has not yet been explored. Our study innovatively applied probe capture-based

tNGS in patients with BSI with the aim of evaluating its etiological diagnostic performance. We present this article in accordance with the STARD reporting checklist (available at <https://jtd.amegroups.com/article/view/10.21037/jtd-24-400/rc>).

## Methods

### Study design

This prospective study was performed at Zhongshan Hospital, Shanghai, China, from 24 November 2023 to 30 December 2023. A total of 88 patients with suspected BSI were screened and 80 were enrolled in the present study, as shown in *Figure 1*. The inclusion criteria were as follows: (I) age  $\geq 14$  years; (II) matching the diagnostic criteria of systemic inflammatory response syndrome (SIRS); (III) febrile, with temperature  $\geq 38.5$  °C; (IV) at least one biomarker increased among white blood cell count (WBC), C-reactive protein (CRP), erythrocyte sedimentation rate (ESR), and procalcitonin (PCT); and (V) severe infection clinically suspicious of BSI which could not be explained by focal infection. As Zhongshan Hospital does not have the Pediatric Department, all the patients admitted must be  $\geq 14$  years of age. The exclusion criteria were as follows: (I) the initial diagnosis of fever on admission was a non-infectious disease, such as tumors and autoimmune diseases; (II) unclear clinical diagnosis at discharge/death; and (III) refusal to undergo blood culture, mNGS, or tNGS. All patients enrolled in the study underwent simultaneous blood culture, blood mNGS, and blood tNGS during fever after admission. Other specimens such as pus, pleural fluid, peritoneal fluid, bile, cerebrospinal fluid, urine, and sputum were also collected from some patients for culture and mNGS. This study was registered with the Chinese Clinical Trial Registry at [www.chictr.org.cn](http://www.chictr.org.cn), with registration number ChiCTR2300077934. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by the Ethical Review Committee of Zhongshan Hospital, Fudan University, Shanghai, China (No. B2023-356R) and informed consent was taken from all the patients.

A clinical diagnosis of BSI or noninfectious fever was made at discharge/death by a clinician team consisting of at least two senior clinicians/professors and four attending doctors, according to the patient's medical history, signs and symptoms, laboratory and imaging examinations, and response to antibiotic treatment.

### Highlight box

#### Key findings

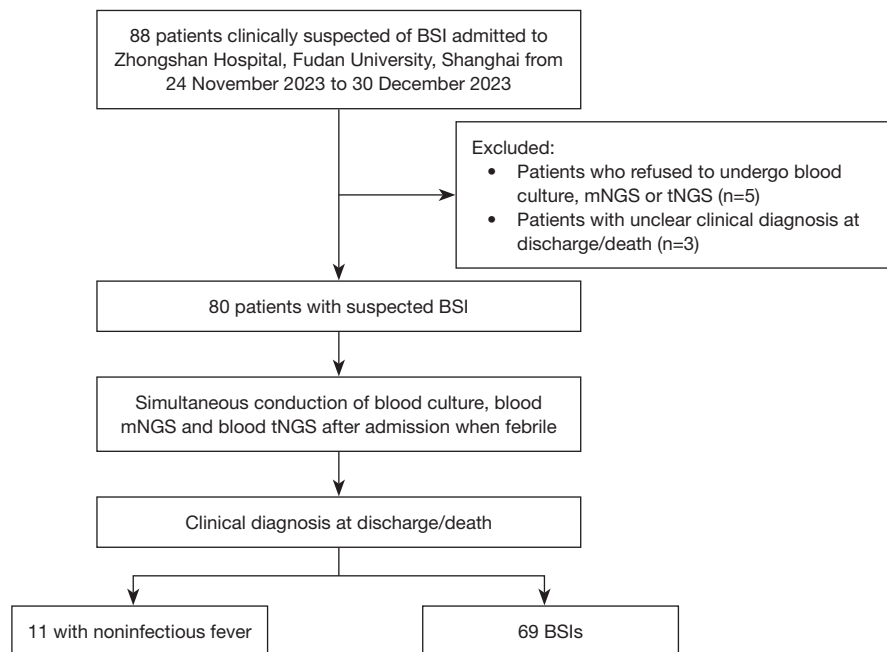
- Blood targeted next-generation sequencing (tNGS) may be a promising tool for detecting potential pathogens in patients with bloodstream infection (BSI).

#### What is known and what is new?

- BSI is a life-threatening disease. Rapid and precise etiological diagnosis is crucial for the treatment of BSI.
- Blood tNGS had a higher sensitivity and faster turnaround time than culture and metagenomic next-generation sequencing.

#### What is the implication, and what should change now?

- The application of blood tNGS for BSI could guide anti-infectious treatment strategies and might improve clinical outcomes.



**Figure 1** Flowchart of patient recruitment and sample collection. A total of 88 patients with suspected BSI were screened from 24 November 2023 to 30 December 2023 at Zhongshan Hospital, Shanghai, China and 80 were enrolled in the study. At discharge/death, 11 patients were clinically diagnosed with noninfectious fever and 69 were diagnosed with BSI. BSI, bloodstream infection; mNGS, metagenomic next-generation sequencing; tNGS, targeted next-generation sequencing.

Demographic information, signs and symptoms of infection, laboratory and imaging examinations, antibiotic treatments, and responses were recorded. Targeted adjustment of anti-infectious therapy was defined as the adjustment of anti-infectious medications by a clinician according to the results of tNGS, mNGS, or culture, including the de-escalation or replacement of anti-infectious medications. Clinical improvement was defined as an improvement in the infectious signs and symptoms or a decrease in inflammatory biomarkers.

### *Etiological testing*

All samples were tested using microbiological culture, mNGS, and tNGS following strict procedures. Culture, mNGS, and tNGS were conducted by different technicians not aware of the results of each other, which ensured adequate blinding between different testing methods. The results were reviewed by at least two clinicians to discriminate between infection and contamination or colonization. If a virus was detected but not related to the disease course and the patient had recovered with antibacterial/antifungal/antiparasitic drugs only and without

antiviral drugs, the virus was considered colonized. mNGS or tNGS results were recognized as positive if the following conditions were met: (I) the read number of the potential pathogen identified by mNGS or tNGS was significantly higher than that of any other microbe; and (II) mNGS or tNGS results were consistent with the clinical course of the patient according to the judgment of the clinicians. If more than one potential pathogen was identified by mNGS or tNGS with approximate read number (within one-fold) and all the identified potential pathogens were consistent with the clinical course of the patient, the mNGS or tNGS results were recognized as polymicrobial infection. Details of the microbiological cultures are provided in [Appendix 1](#). The details of mNGS and tNGS testing are as follows.

### *mNGS workflow*

#### **Sample processing and DNA extraction**

Blood samples of 3 mL were drawn from patients and stored at room temperature for 3–5 minutes before plasma separation, followed by centrifugation at 4,000 rpm for 10 minutes at 4 °C. The plasma samples were then transferred to new sterile tubes. Three mL of pus, sputum,

or body fluids were collected from the patients according to standard procedures. Sputum was liquefied by 0.1% DTT (dithiothreitol) for 30 minutes at room temperature. Pus or body fluids could go directly to the next operation. DNA was extracted from 300  $\mu$ L of plasma using the TIANamp Micro DNA Kit (DP316, TIANGEN BIOTECH, Beijing, China), following the operational manual of the manufacturer. The extracted DNA samples were used to construct DNA libraries (14).

### Construction of DNA libraries and sequencing

DNA libraries were constructed using DNA fragmentation, end repair, adapter ligation, and PCR amplification by MetaCAP Pathogen Capture Metagenomic Assay Kit (KingCreate, Guangzhou, China). Agilent 2100 (Agilent, Santa Clara, CA, USA) was used for quality control of the DNA libraries. Quality-qualified libraries were pooled. Then, a DNA nanoball (DNB) was prepared and sequenced using the MGISEQ-2000 platform (MGI Tech, Shenzhen, China) set to 50-bp single end with average of 100 million reads per sample (15).

### tNGS workflow

#### Sample processing and nucleic acid extraction

Blood samples of 3 mL were centrifuged at 1,600 rcf for 10 minutes at 4 °C. Nucleic acid was extracted from 300  $\mu$ L of plasma using the MasterPure DNA & RNA Extraction Kit (KingCreate) following the manufacturer's operational manual and was used for the construction of libraries.

#### Construction of libraries, enrichment, and sequencing

After complementary DNA (cDNA) was converted from RNA, libraries were constructed through fragmentation, end repair, adapter ligation, and PCR amplification. Then, eight uniquely barcoded libraries were pooled for hybridization and captured using specific biotinylated probes for 2 hours after library generation using the MetaCAP Pathogen Capture Metagenomic Assay Kit (KingCreate), following the manufacturer's instructions. The enrichment panel targeted >3,000 pathogens, including viruses, bacteria, fungi, and parasites. The pathogen target was screened according to the following rules: (I) the target had at least one completed genome sequence; (II) the target had at least one article related to human infection; (III) the target had research reports. Finally, this capture range was 1,060 bacteria, 1,402 viruses, 388 fungi and 210 parasites. Probe design selected the conserved regions such

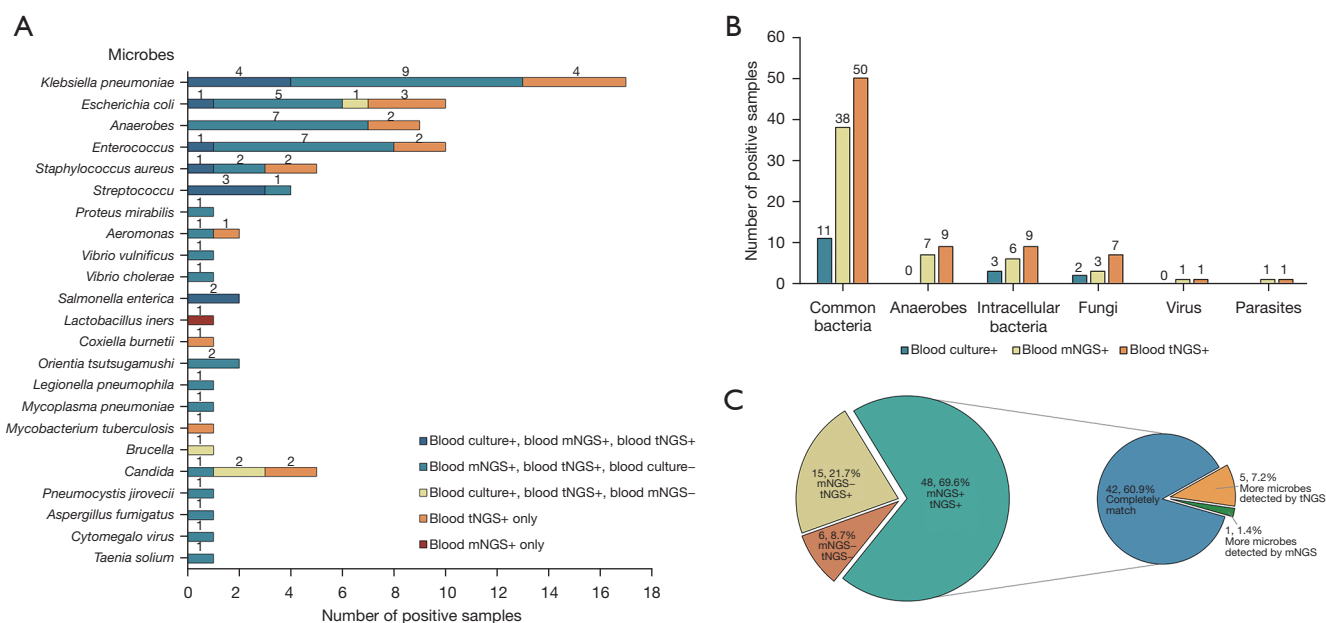
as ribosomal RNA (rRNA) gene (16S rRNA, 18S rRNA or internal transcribed spacer), house-keeping genes which allowed for pathogen identification down to the genus or species level. Agilent 2100 was used for the quality control of the libraries and showed peaks near 350 bp. The Qubit dsDNA HS Assay Kit (Thermo Fisher Scientific Inc., Waltham, MA, USA) was used to construct quality-qualified libraries, and sequencing was performed on an Illumina MiniSeq platform set to 100-bp single end with average of 1 million reads per sample.

### Bioinformatic analysis

Bioinformatic analyses of both mNGS and tNGS data were conducted according to the following process. Clean reads were obtained by removing sequencing adapters, reads of low quality, excessive N bases, or reads with length below 35 bp using fastp (version 0.23.1) (16). The remaining reads were aligned to the human reference (hg38) using Burrows-Wheeler aligner (BWA; version 0.7.17-r1188; Linux, San Francisco, CA, USA), and human reads were filtered (17). Subsequently, the reads were compared to the classification reference genome database. The scope of the database contained all targeted pathogens and other species in their genus. In addition, it also included species outside the genus with genome average nucleotide identity >80%. Database contained 11,958 bacteria, 7,373 viruses, 1,714 fungi and 343 parasites. The species genomes were collected and downloaded from the National Center for Biotechnology Information (NCBI) genome (<ftp://ftp.ncbi.nlm.nih.gov/genomes/>). Genome assemble level at complete or scaffold was selected and shielded contamination introduced by assembly, such as host sequences. Self-developed software was used to calculate the number of reads per million (RPM) at the species and genus levels.

### Statistical analysis

Continuous data were expressed as the mean  $\pm$  standard deviation or median with interquartile range (IQR, 25th–75th percentiles), depending on the normality of the data distribution. The final clinical diagnosis at discharge or death was the gold standard. The sensitivity and specificity of blood culture, blood mNGS, and blood tNGS, determined based on whether the test reflected the clinical diagnosis of BSI, were compared using the Chi-square test. Data analysis was performed using SPSS software (version 21.0; IBM Corp., Armonk, NY, USA). All tests were two-



**Figure 2** Comparison of blood culture, blood mNGS, and blood tNGS in detecting microbes for BSI patients. (A) Different microbes detected by blood culture, blood mNGS, and blood tNGS in 69 patients with BSI. (B) The ability of blood culture, blood mNGS, and blood tNGS to detect specific types of microbes in 69 patients with BSI. Blood tNGS identified more common bacteria, anaerobes, intracellular bacteria, and fungi than blood culture and blood mNGS. (C) Consistency between blood tNGS and blood mNGS results. In 69 patients with BSI, 15 cases (21.7%) were blood mNGS negative and blood tNGS positive. Six cases (8.7%) were both negative for blood mNGS and blood tNGS. Forty-eight cases (69.6%) were both positive for blood mNGS and blood tNGS, among which 42 cases (60.9%) had tNGS results completely concordant with mNGS. In the other 5 cases (7.2%), tNGS detected more potential pathogens than mNGS. In the last 1 case (1.4%), mNGS detected more potential pathogen than tNGS. mNGS, metagenomic next-generation sequencing; tNGS, targeted next-generation sequencing; BSI, bloodstream infection.

tailed, and statistical significance was set at  $P < 0.05$ .

## Results

### Characteristics of the study participants

The demographic and clinical characteristics of the patients are provided in Table S1. A total of 80 patients were enrolled in this study, of whom 11 were diagnosed with noninfectious fever and 69 were diagnosed with BSI. Among the 69 patients with BSI, 50 (72.5%) were male, and the mean age was  $59.4 \pm 15.8$  years. A total of 19 BSIs (27.5%) were nosocomial infections, and 59 patients with BSI (85.5%) received empirical antibiotic treatment within 4 weeks before admission. Nosocomial infections referred to infections acquired during hospitalization. Infections occurred after discharge but acquired during hospitalization still belonged to nosocomial infections. The underlying infectious syndromes of the 69 patients

with BSI were classified into 11 types: abdominal infections (26 cases, 37.7%), infective endocarditis (9 cases, 13.0%), catheter/device related infections (8 cases, 11.6%), urinary tract infections (7 cases, 10.1%), skin and soft tissue infections (7 cases, 10.1%), respiratory infections (6 cases, 8.7%), infectious aneurysm (2 cases, 2.9%), osteoarticular infections (1 case, 1.4%), septic thrombophlebitis (1 case, 1.4%), central nervous system infections (1 case, 1.4%), and other/unknown infections (1 case, 1.4%). The underlying risk factors in the 69 patients with BSI were as follows: active malignancy in 13 cases (18.8%), diabetes mellitus in 12 cases (17.4%), and long-term use of immunosuppressive therapy in 7 cases (10.1%).

Table S2 lists the results of blood culture, mNGS, and tNGS for the 80 participants. For the 69 patients with BSI, the numbers of different types of microbes detected by blood culture, blood mNGS, and blood tNGS are shown in Figure 2A, 2B. The consistency between blood mNGS and blood tNGS is described in Figure 2C.



**Table 1** Diagnostic performance of blood culture, blood mNGS, and blood tNGS

Methods	Positive number		Sensitivity (%)	Specificity (%)
	Noninfectious fever (n=11)	BSI (n=69)		
Blood culture	0	16	23.2	100.0
Blood mNGS	2	48	69.6	81.8
Blood tNGS	0	63	91.3	100.0

mNGS, metagenomic next-generation sequencing; tNGS, targeted next-generation sequencing; BSI, bloodstream infection.

**Table 2** Comparison of blood culture, blood mNGS, and blood tNGS

Case classification	Number of samples	Identical findings	New microbes detected by tNGS	Microbes not identified by tNGS
Blood tNGS versus blood culture				
Noninfectious fever	11	11 (100.0)	NA	NA
Blood culture-positive BSI	16	13 (81.3)	3 (18.8) <sup>a</sup>	0
Blood culture-negative BSI	53	6 (11.3)	47 (88.7) <sup>b</sup>	NA
Blood tNGS versus blood mNGS				
Noninfectious fever	11	9 (81.8)	NA	NA
Blood mNGS-positive BSI	48	42 (87.5)	5 (10.4) <sup>c</sup>	1 (2.1) <sup>d</sup>
Blood mNGS-negative BSI	21	6 (28.6)	15 (71.4) <sup>e</sup>	NA

Data shown are the n (%) of samples in which there are identical or discrepant findings between blood culture, blood mNGS and blood tNGS. <sup>a</sup>, 5 new microbes were detected by tNGS in 16 blood culture-positive BSIs; <sup>b</sup>, 56 new microbes were detected by tNGS in 53 blood culture-negative BSIs; <sup>c</sup>, 5 new microbes were detected by tNGS in 48 blood mNGS-positive BSIs; <sup>d</sup>, 1 microbe was missed by tNGS in 48 blood mNGS-positive BSIs; <sup>e</sup>, 17 new microbes were detected by tNGS in 21 blood mNGS-negative BSIs. mNGS, metagenomic next-generation sequencing; tNGS, targeted next-generation sequencing; NA, not applicable; BSI, bloodstream infection.

### Diagnostic performance of blood culture, blood mNGS, and blood tNGS

Blood tNGS had a higher sensitivity for the diagnosis of BSI than blood culture (91.3% vs. 23.2%,  $P < 0.001$ ). Additionally, blood tNGS had a higher sensitivity than blood mNGS (91.3% vs. 69.6%,  $P = 0.001$ ), as shown in Table 1. The specificities of blood culture, blood mNGS, and blood tNGS for the diagnosis of BSI were 100.0%, 81.8%, and 100.0%, respectively. There was no significant difference in the specificity between blood mNGS and tNGS ( $P = 0.13$ ).

#### Blood tNGS versus blood culture

Of the 16 blood culture-positive BSIs, microbes were detected by tNGS in 16 cases (100.0%), including a single microbe detected by tNGS in 13 cases and multiple

microbes in three cases. The most common microbes identified using both methods were *Enterobacteriaceae*, including *Klebsiella pneumoniae* (*K. pneumoniae*) and *Escherichia coli* (*E. coli*). The results of tNGS were concordant with the blood culture in 13 cases (81.3%), with additional potential pathogens detected in three cases (18.8%), including *Enterococcus* in two cases, and *Staphylococcus aureus* (*S. aureus*) and *Candida* each in one case, as shown in Table 2.

Of the 53 patients with blood culture-negative BSIs, 6 (11.3%) had negative tNGS results, whereas 56 additional microbes were detected by tNGS among the remaining 47 (88.7%) patients. A total of eight participants presented with polymicrobial infections. Additional microbes detected by tNGS included *K. pneumoniae* (13 cases), *Anaerobes* (9 cases), *E. coli* (8 cases), *Enterococcus* (6 cases), *S. aureus* (2 cases), *Aeromonas* (2 cases), *Orientia tsutsugamushi* (*O. tsutsugamushi*) (2 cases), *Vibrio vulnificus* (1 case), *Vibrio*

*cholerae* (1 case), *Streptococcus* (1 case), *Coxiella burnetii* (*C. burnetii*) (1 case), *Mycobacterium tuberculosis* (*M. tuberculosis*) (1 case), *Legionella pneumophila* (*L. pneumophila*) (1 case), *Mycoplasma pneumoniae* (*M. pneumoniae*) (1 case), *Proteus mirabilis* (1 case), *Candida* (2 cases), *Aspergillus fumigatus* (1 case), *Pneumocystis jirovecii* (1 case), *Cytomegalovirus* (1 case), and *Taenia solium* (1 case).

### Blood tNGS versus blood mNGS

Among the 48 patients with blood mNGS-positive BSIs, the blood tNGS results were also all positive, which were concordant with the blood mNGS results in 42 cases (87.5%), with additional potential pathogens detected in 5 (10.4%) cases, including *Candida* in two cases and *S. aureus*, *K. pneumoniae*, and *Enterococcus* in one case each, as shown in Table 2. In one case (2.1%), *Lactobacillus iners* was detected by mNGS but missed by tNGS. The most common microbe identified using both methods was *K. pneumoniae*, followed by *Enterococcus*.

Of the 21 patients with blood mNGS-negative BSIs, 6 (28.6%) had negative tNGS results, whereas 17 additional microbes were detected by tNGS in the remaining 15 cases (71.4%). Polymicrobial infections were detected in two participants. New microbes detected by tNGS included *E. coli* (4 cases), *K. pneumoniae* (3 cases), *Anaerobes* (2 cases), *Candida* (2 cases), *Enterococcus* (1 case), *S. aureus* (1 case), *Aeromonas* (1 case), *C. burnetii* (1 case), *M. tuberculosis* (1 case), and *Brucella* (1 case).

### Concordance between blood tNGS results and microbes detected by culture or mNGS in other specimens

Among the 69 patients with BSIs, 23 patients underwent culture or mNGS of other specimens such as pus, sputum, pleural fluid, peritoneal fluid, bile, cerebrospinal fluid, or urine, as shown in Table S3. Among the 23 patients, 21 (91.3%) had blood tNGS results consistent with the microbes detected by culture or mNGS in other specimens. In one patient (P56), blood tNGS identified *Enterococcus faecalis* (*E. faecalis*), *Enterococcus faecium* (*E. faecium*), and *K. pneumoniae*, whereas the bile culture grew *E. faecalis* and bile mNGS identified *E. faecalis* and *K. pneumoniae*. In another patient (P73), blood tNGS identified *Bacteroides thetaiotaomicron* (*B. thetaiotaomicron*) and *E. faecium*, whereas culture of the peritoneal fluid grew *E. faecium* and *E. coli*, and mNGS of the peritoneal fluid identified *B. thetaiotaomicron*, *E. faecium*, *K. pneumoniae*, and *Candida albicans*.

### Analysis of cases with noninfectious fever

Of the 11 patients with noninfectious fever, microbes were detected by blood culture in 0 patients, mNGS in two cases, and tNGS in 0 cases. The two microbes detected by mNGS were *Streptococcus mitis* and *Haemophilus parainfluenzae*, which were suspected to be contaminant. These two patients were respectively diagnosed with Takayasu arteritis and polyarteritis nodosa. After transferred to the Rheumatology Department, they received glucocorticoid and immunosuppressant treatment and clinically improved.

### Turnaround time (TAT) of blood culture, mNGS, and tNGS

TAT was defined as the time from sample reception to identification of potential pathogens. Among the 12 patients who were simultaneously positive by blood culture, blood tNGS, and blood mNGS, TAT of blood culture, blood mNGS, and blood tNGS were 50.1±9.9, 30.5, and 16 hours, respectively. Blood tNGS had a faster TAT than blood culture and blood mNGS.

### Targeted adjustment of anti-infectious therapy

Among the 69 patients with BSI, 22 (31.9%) underwent targeted adjustment of anti-infectious therapy according to the blood tNGS results and showed clinical improvement, as shown in Table 3. Among these 22 patients, 6 (27.3%) were blood culture positive, 17 (77.3%) were blood mNGS-positive, and 22 (100.0%) were blood tNGS-positive. Due to having a faster TAT compared to that of culture and mNGS, blood tNGS guided the clinical anti-infectious treatment in the first place. Through the rapid and broad detection of potential pathogens, blood tNGS minimizes the time required for empirical treatment and allows for precise anti-infectious therapy. For the other 47 patients with BSI, tNGS results were concordant with the clinician's initial evaluation of the patient and the initial anti-infectious therapy continued.

## Discussion

Despite the great advances in medical science in the past century, BSI remains a growing public health concern, with an incidence ranging between 113 and 204 per 100,000 people, including nosocomial BSIs that account for approximately 30–50% of the cases in high-income

**Table 3** Targeted adjustment of anti-infectious therapy according to blood tNGS in patients with BSI

Patient number	Anti-infectious therapy before tNGS testing	Anti-infectious therapy after adjustment	Microbes detected by tNGS	Underlying infectious syndromes
P15	Ceftriaxone + levofloxacin	Piperacillin/tazobactam	<i>Treponema denticola</i> , <i>Fusobacterium vincentii</i>	Catheter/device related infections <sup>a</sup>
P19	Ceftazidime/avibactam	Ceftazidime/avibactam + daptomycin	<i>Klebsiella pneumoniae</i> , <i>Staphylococcus aureus</i>	Infective endocarditis <sup>b</sup> (PVE)
P24	Meropenem	Ceftriaxone + levofloxacin	<i>Aeromonas veronii</i>	Skin and soft tissue infections
P26	Meropenem + vancomycin	Ceftriaxone + levofloxacin	<i>Vibrio vulnificus</i>	Skin and soft tissue infections
P33	Vancomycin	Ceftriaxone + amikacin	<i>Streptococcus sanguinis</i>	Infective endocarditis (NVE)
P35	Meropenem	Piperacillin/tazobactam+ caspofungin	<i>Escherichia coli</i> , <i>Candida parapsilosis</i>	Urinary tract infections
P39	Ertapenem	Caspofungin	<i>Candida parapsilosis</i>	Other/unknown infections
P42	Meropenem + levofloxacin	Sulfamethoxazole and trimethoprim, ganciclovir	<i>Pneumocystis jirovecii</i> , <i>Cytomegalo virus</i>	Respiratory infections
P44	Piperacillin/tazobactam	Doxycycline	<i>Orientia tsutsugamushi</i>	Skin and soft tissue infections
P45	Cefoperazone/sulbactam	Cefoperazone/sulbactam + metronidazole	<i>Bacteroides fragilis</i>	Abdominal infections
P50	Levofloxacin	Isoniazid + rifampin + ethambutol + pyrazinamide	<i>Mycobacterium tuberculosis</i>	Respiratory infections
P52	Ceftriaxone	Doxycycline	<i>Orientia tsutsugamushi</i>	Skin and soft tissue infections
P53	Meropenem	Ceftriaxone + levofloxacin	<i>Salmonella enterica</i>	Infectious aneurysm
P55	Piperacillin/tazobactam	Doxycycline	<i>Mycoplasma pneumoniae</i>	Respiratory infections
P57	Meropenem + vancomycin	Caspofungin	<i>Candida metapsilosis</i>	Infective endocarditis (PVE)
P59	Isoniazid + rifampin + ethambutol + pyrazinamide	Albendazole + praziquantel	<i>Taenia solium</i>	Central nervous system infections
P61	Ceftriaxone	Doxycycline + rifampin	<i>Brucella</i>	Osteoarticular infection
P62	Vancomycin	Doxycycline + moxifloxacin	<i>Coxiella burnetii</i>	Catheter/device related infections
P70	Piperacillin/tazobactam	Moxifloxacin	<i>Legionella pneumophila</i>	Respiratory infections
P72	Piperacillin/tazobactam	Vancomycin + caspofungin	<i>Enterococcus faecalis</i> , <i>Candida glabrata</i>	Abdominal infections
P73	Tigecycline	Vancomycin + piperacillin/tazobactam	<i>Enterococcus faecium</i> , <i>Bacteroides thetaiotaomicron</i>	Abdominal infections
P75	Meropenem	Voriconazole	<i>Aspergillus fumigatus</i>	Septic thrombophlebitis

<sup>a</sup>, including infections related to central line catheters, artificial blood vessels, and stent implantation; <sup>b</sup>, including NVE, PVE, and pacemaker-related endocarditis. tNGS, targeted next-generation sequencing; BSI, bloodstream infection; NVE, native valve endocarditis; PVE, prosthetic valve endocarditis.

countries (18). Overall, *E. coli* was the most common pathogen causing BSI, followed by *Klebsiella* spp. and *S. aureus*. Other key pathogens of BSI include coagulase-negative staphylococci, *Pseudomonas aeruginosa*, *Acinetobacter*

*baumanni*, *Salmonella enterica*, *Streptococcus* spp., and *Enterococcus* spp., the first three of which are almost always healthcare-associated (18). The short-term case fatality rate following BSI ranges from 13.5% to 20.6%, whereas the



1-year mortality ranges from 8% to 48% (1,19). Prompt etiological diagnosis and anti-infectious therapy are essential for the effective treatment of BSI.

Blood cultures remain an imperfect diagnostic tool for deadly BSIs; 30–50% of positive blood cultures are contaminated, whereas false-negative blood cultures occur in 40–60% of cases with suspected sepsis, leading to a diagnostic conundrum (6). Novel molecular diagnostics, mainly nucleic acid amplification-based testing, have dramatically changed the clinical approach to the diagnosis of BSI, with significant improvements in sensitivity and speed. mNGS has been used to diagnose BSI. In a previous study conducted by Sun *et al.*, the positivity rate of blood mNGS was higher than that of blood culture (67.74% *vs.* 19.35%,  $P < 0.001$ ) in intensive unit care patients with sepsis (20). However, due to the interference of host DNA and low biomass of circulating pathogens, mNGS in serum samples is still not sufficiently sensitive. Furthermore, owing to the complexity of data processing and analysis, high cost, and difficulty to reduce TAT, the expansion of mNGS applications for BSI has encountered limitations.

Various target enrichment strategies have been used for the sequencing and analysis of a number of targets of interest (9–11). Hernández-Neuta *et al.* adopted molecular inversion probes as a cost-effective target-enrichment approach to identify pathogens and antimicrobial resistance markers in blood samples using short-read sequencing (13). Li *et al.* applied tNGS for target detection of 153 pathogens in adults with pneumonia, and observed that tNGS was effective and comparable to mNGS in detecting respiratory pathogens (21). Fida *et al.* prospectively investigated 16S rRNA gene-based tNGS in 60 adults with sepsis and compared tNGS results with blood cultures. They concluded that 16S rRNA gene-based tNGS may be useful for detecting bacteria in the plasma of patients with sepsis (22). In a study conducted by Hong *et al.*, 16S rRNA gene-based tNGS was found to be a potential diagnostic tool for the identification of periprosthetic joint infection pathogens. However, there was no difference in the positive percent agreement between 16S rRNA gene-based tNGS and shotgun metagenomic sequencing (sNGS) (72.1% *vs.* 73.1%,  $P = 0.83$ ) for detecting microorganisms in the sonicate fluid of periprosthetic joint infection (23). We assume that this result was attributable to the retrospective study design in which sonicate fluid sNGS had been conducted between 2011 and 2016, whereas  $-80\text{ }^{\circ}\text{C}$  frozen clinical samples were collected for tNGS testing in 2022. Samples stored for several years may have affected the detection efficiency of tNGS. The results of this study may

also be affected by the inherent limitations of the 16S rRNA gene-based tNGS methodology.

However, the application of probe capture-based tNGS for the etiological diagnosis of BSI has not been explored. Compared with mNGS, probe capture-based tNGS has fewer data burdens and simplified data interpretation, facilitating a lower cost and faster TAT (12). In this study, we demonstrated that blood tNGS has significantly higher sensitivity and faster TAT than blood mNGS for the diagnosis of BSI. However, the specificities of blood tNGS and mNGS were not statistically different. tNGS especially outperformed mNGS in the detection of fungi and intracellular bacteria, including *M. pneumoniae*, *L. pneumophila*, *O. tsutsugamushi*, *Salmonella enterica*, *Brucella*, *C. burnetii*, and *M. tuberculosis*. The biomass of fungi and intracellular bacteria in the serum is relatively low, which may explain the false negative results of mNGS to some extent. Probe capture and enrichment in tNGS increases the ability to detect fungi and intracellular bacteria.

Furthermore, mNGS can generally detect either DNA or RNA pathogens, but not both. If mNGS covers DNA and RNA dual processes, testing will become more complicated and double in cost, increasing the economic burden on patients. In contrast, tNGS inherently covers both DNA and RNA pathogens and is much more convenient and cost-effective than mNGS.

Compared with tNGS, mNGS has much more data burdens and the data interpretation process is more complex. The data size is approximately 100–120 M per sample for mNGS. Fifty bp is recommended by BGI to balance the diagnosis performance and clinical TAT. If 50 bp increases to 100 bp, time spent on sequencing will double, consuming almost 30 hours on the sequencing platform. TAT of 100 bp in mNGS cannot meet clinical needs. However, in tNGS, target capture and enrichment can reduce data burdens. The data size is approximately 1 M per sample for tNGS and sequencing needs only 6 hours on Illumina Miniseq platform. One hundred bp is more suitable for tNGS because specific target for pathogen identification requires certain length. These are the inherent differences between mNGS and tNGS.

In this study, 23 patients with BSI underwent culture or mNGS of specimens other than blood, such as pus, sputum, or body fluids. Among these, 21 (91.3%) had blood tNGS results consistent with those of microbes detected by culture or mNGS of other specimens, demonstrating the accuracy of blood tNGS for the etiological diagnosis of BSI. Compared with the culture or mNGS of other specimens,

blood tNGS is much more convenient as blood samples can be easily collected at admission. In contrast, the collection of other specimens is more complicated, requiring procedures such as percutaneous puncture and drainage, which are more time consuming.

Among the 69 patients with BSI, 22 underwent targeted adjustment of anti-infectious therapy according to the tNGS results and clinically improved. tNGS may help clinicians to downsize antibiotic regimens and decrease the empirical usage of broad-spectrum antibiotics, which might reduce antibiotic abuse and affect the clinical prognosis.

Despite these findings, this study has some limitations. As this was a single-center study, a patient selection bias may have existed. The gold standard was the clinical evaluation at death/discharge which would follow the testing and thus further introduce bias as evaluations would be done retrospectively. The number of patients enrolled was relatively small, and we could not divide the patients into subgroups according to the underlying infectious syndromes. Additionally, no RNA microbes were detected in this study and no patient with viremia was diagnosed, perhaps because of the small sample size.

## Conclusions

In conclusion, blood tNGS may be a promising tool for the etiological diagnosis of BSI. Owing to its higher sensitivity and faster TAT than blood mNGS and blood culture, blood tNGS is a valuable tool for identifying the pathogens responsible for BSI and guiding anti-infectious treatments.

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## Footnote

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**Ethical Statement:** The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by the Ethical Review Committee of Zhongshan Hospital, Fudan University, Shanghai, China (No. B2023-356R) and informed consent was taken from all the patients.

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## Appendix 1

*Details of the microbiological cultures*

All the patients underwent blood culture by peripheral venous puncture at least once during fever after admission. Blood samples (8–10 mL) were injected into aerobic, anaerobic, and fungal blood culture bottles (BD BACTECTM, Becton, Dickinson, and Co., Franklin Lakes, NJ, USA) and then loaded into an automated continuous monitoring system (BD BACTECTM, USA) for 7 days. Other specimens such as pus, pleural fluid, peritoneal fluid, bile, cerebrospinal fluid, urine and sputum were cultured onto blood agar plates, chocolate agar plates, and fungal chromogenic plates for 7 days. If culture showed bacterial or fungal growth, strain identification was conducted by VITEK MALDI-TOF mass spectrometry (bioMérieux, Craponne, France). Antimicrobial susceptibility tests were conducted using Vitek 2 (bioMérieux, France).

**Table S1** Demographic and clinical characteristics of the study participants

Characteristics	Noninfectious fever (n=11)	BSI (n=69)
Age (years)	60.6±12.3	59.4±15.8
Gender, male	7 (63.6)	50 (72.5)
Underlying infectious syndromes		
Abdominal infections	NA	26 (37.7)
Infective endocarditis <sup>a</sup>	NA	9 (13.0)
Catheter/device related infections <sup>b</sup>	NA	8 (11.6)
Urinary tract infections	NA	7 (10.1)
Skin and soft tissue infections	NA	7 (10.1)
Respiratory infections	NA	6 (8.7)
Infectious aneurysm	NA	2 (2.9)
Osteoarticular infections	NA	1 (1.4)
Septic thrombophlebitis	NA	1 (1.4)
Central nervous system infections	NA	1 (1.4)
Other/unknown infections	NA	1 (1.4)
Underlying risks		
Active malignancy	5 (45.5)	13 (18.8)
Diabetes mellitus	1 (9.1)	12 (17.4)
Long-term use of immunosuppressive therapy	0	7 (10.1)
Nosocomial infection	NA	19 (27.5)
Antibiotics within 4 weeks before admission	10 (90.9)	59 (85.5)
Duration of fever on admission (days)	17 [14–30]	20 [7–30]
Laboratory		
WBC (10 <sup>9</sup> /L)	10.4 [6.3–13.2]	8.9 [6.4–12.5]
ESR (mm/h)	80 [40–120]	41 [18–65]
CRP (mg/L)	136 [87–300]	94 [49.5–184.2]
PCT (ng/mL)	0.6 [0.1–1.1]	0.5 [0.2–3.3]

Data are expressed as mean ± standard deviation, n (%) or median [IQR]. <sup>a</sup>, including native valve endocarditis, prosthetic valve endocarditis and pacemaker-related endocarditis; <sup>b</sup>, including infections related to central line catheters, artificial blood vessels and stent implantation. BSI, bloodstream infection; WBC, white blood cell count; ESR, erythrocyte sedimentation rate; CRP, C-reactive protein; PCT, procalcitonin; NA, not applicable; IQR, interquartile range.

**Table S2** Results of blood culture, blood mNGS and blood tNGS of the 80 patients enrolled in this study

Patient number	Blood culture	Blood mNGS	Blood tNGS	Underlying infectious syndromes
Patients with noninfectious fever (n=11)				
P1	Negative	Negative	Negative	NA
P2	Negative	<i>Streptococcus mitis</i>	Negative	NA
P3	Negative	Negative	Negative	NA
P4	Negative	Negative	Negative	NA
P5	Negative	Negative	Negative	NA
P6	Negative	<i>Haemophilus parainfluenzae</i>	Negative	NA
P7	Negative	Negative	Negative	NA
P8	Negative	Negative	Negative	NA
P9	Negative	Negative	Negative	NA
P10	Negative	Negative	Negative	NA
P11	Negative	Negative	Negative	NA
Patients with BSI (n=69)				
P12	<i>Streptococcus oralis</i> ( <i>S. oralis</i> )	<i>S. oralis</i>	<i>S. oralis</i>	Infective endocarditis <sup>a</sup>
P13	<i>Staphylococcus aureus</i> ( <i>S. aureus</i> )	<i>S. aureus</i>	<i>S. aureus</i>	Infective endocarditis
P14	<i>Escherichia coli</i> ( <i>E. coli</i> )	Negative	<i>E. coli</i> , <i>Enterococcus faecium</i>	Abdominal infections
P15	Negative	<i>Treponema denticola</i> ( <i>T. denticola</i> ), <i>Fusobacterium nucleatum</i> ( <i>F. nucleatum</i> )	<i>T. denticola</i> , <i>F. nucleatum</i>	Catheter/device related infections <sup>b</sup>
P16	Negative	<i>Klebsiella pneumoniae</i> ( <i>K. pneumoniae</i> )	<i>K. pneumoniae</i>	Abdominal infections
P17	Negative	Negative	Negative	Abdominal infections
P18	Negative	Negative	<i>K. pneumoniae</i>	Abdominal infections
P19	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>	<i>K. pneumoniae</i> , <i>S. aureus</i>	Infective endocarditis
P20	Negative	Negative	Negative	Abdominal infections
P21	Negative	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>	Abdominal infections
P22	<i>E. coli</i>	<i>E. coli</i>	<i>E. coli</i>	Urinary tract infections
P23	Negative	Negative	<i>S. aureus</i>	Skin and soft tissue infections
P24	Negative	<i>Aeromonas veronii</i> ( <i>A. veronii</i> )	<i>A. veronii</i>	Skin and soft tissue infections
P25	Negative	Negative	<i>K. pneumoniae</i>	Abdominal infections
P26	Negative	<i>Vibrio vulnificus</i> ( <i>V. vulnificus</i> )	<i>V. vulnificus</i>	Skin and soft tissue infections
P27	Negative	Negative	<i>E. coli</i>	Urinary tract infections
P28	Negative	<i>K. pneumoniae</i> , <i>Lactobacillus iners</i>	<i>K. pneumoniae</i>	Abdominal infections
P29	Negative	Negative	<i>E. coli</i>	Urinary tract infections

**Table S2** (Continued)



Table S2 (Continued)

Patient number	Blood culture	Blood mNGS	Blood tNGS	Underlying infectious syndromes
P30	Negative	<i>Enterococcus raffinosus</i> ( <i>E. raffinosus</i> )	<i>E. raffinosus</i> , <i>K. pneumoniae</i>	Abdominal infections
P31	Negative	Negative	<i>K. pneumoniae</i>	Catheter/device related infections
P32	Negative	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>	Abdominal infections
P33	<i>Streptococcus sanguinis</i> ( <i>S. sanguinis</i> )	<i>S. sanguinis</i>	<i>S. sanguinis</i>	Infective endocarditis
P34	Negative	<i>E. coli</i>	<i>E. coli</i>	Abdominal infections
P35	Negative	<i>E. coli</i>	<i>E. coli</i> , <i>Candida parapsilosis</i> ( <i>C. parapsilosis</i> )	Urinary tract infections
P36	Negative	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>	Abdominal infections
P37	Negative	Negative	<i>E. coli</i>	Abdominal infections
P38	Negative	Negative	Negative	Infective endocarditis
P39	<i>C. parapsilosis</i>	Negative	<i>C. parapsilosis</i>	Other/unknown infections
P40	Negative	<i>E. coli</i>	<i>E. coli</i>	Urinary tract infections
P41	Negative	Negative	Negative	Catheter/device related infections
P42	Negative	<i>Pneumocystis jirovecii</i> ( <i>P. jirovecii</i> ), <i>Cytomegalo virus</i>	<i>P. jirovecii</i> , <i>Cytomegalo virus</i>	Respiratory infections
P43	Negative	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>	Respiratory infections
P44	Negative	<i>Orientia tsutsugamushi</i> ( <i>O. tsutsugamushi</i> )	<i>O. tsutsugamushi</i>	Skin and soft tissue infections
P45	Negative	<i>Bacteroides fragilis</i> ( <i>B. fragilis</i> )	<i>B. fragilis</i>	Abdominal infections
P46	Negative	<i>Streptococcus agalactiae</i> ( <i>S. agalactiae</i> )	<i>S. agalactiae</i>	Infective endocarditis
P47	Negative	Negative	Negative	Abdominal infections
P48	Negative	<i>E. coli</i>	<i>E. coli</i>	Abdominal infections
P49	Negative	<i>Prevotella oris</i> ( <i>P. oris</i> )	<i>P. oris</i>	Respiratory infections
P50	Negative	Negative	<i>Mycobacterium tuberculosis</i>	Respiratory infections
P51	<i>Enterococcus faecalis</i>	<i>E. faecalis</i>	<i>E. faecalis</i>	Catheter/device related infections
P52	Negative	<i>O. tsutsugamushi</i>	<i>O. tsutsugamushi</i>	Skin and soft tissue infections
P53	<i>Salmonella enterica</i>	<i>S. enterica</i>	<i>S. enterica</i>	Infectious aneurysm
P54	Negative	<i>Bacteroides dorei</i> ( <i>B. dorei</i> )	<i>B. dorei</i>	Abdominal infections
P55	Negative	<i>Mycoplasma pneumoniae</i> ( <i>M. pneumoniae</i> )	<i>M. pneumoniae</i>	Respiratory infections
P56	Negative	<i>E. faecalis</i> , <i>E. faecium</i> , <i>K. pneumoniae</i>	<i>E. faecalis</i> , <i>E. faecium</i> , <i>K. pneumoniae</i>	Abdominal infections

Table S2 (Continued)

Table S2 (Continued)

Patient number	Blood culture	Blood mNGS	Blood tNGS	Underlying infectious syndromes
P57	<i>Candida metapsilosis</i> ( <i>C. metapsilosis</i> )	Negative	<i>C. metapsilosis</i>	Infective endocarditis
P58	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>	Catheter/device related infections
P59	Negative	<i>Taenia solium</i> ( <i>T. solium</i> )	<i>T. solium</i>	Central nervous system infections
P60	Negative	<i>Vibrio cholerae</i> ( <i>V. cholerae</i> )	<i>V. cholerae</i>	Skin and soft tissue infections
P61	<i>Brucella</i>	Negative	<i>Brucella</i>	Osteoarticular infection
P62	Negative	Negative	<i>Coxiella burnetii</i>	Catheter/device related infections
P63	Negative	Negative	<i>Aeromonas caviae</i>	Skin and soft tissue infections
P64	<i>S. sanguinis</i>	<i>S. sanguinis</i>	<i>S. sanguinis</i>	Infective endocarditis
P65	Negative	<i>S. aureus</i>	<i>S. aureus</i>	Catheter/device related infections
P66	Negative	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>	Abdominal infections
P67	Negative	Negative	Negative	Infectious aneurysm
P68	Negative	Negative	<i>Bacteroides thetaiotaomicron</i> ( <i>B. thetaiotaomicron</i> ), <i>Parabacteroides distasonis</i>	Catheter/device related infections
P69	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>	Abdominal infections
P70	Negative	<i>Legionella pneumophila</i> ( <i>L. pneumophila</i> )	<i>L. pneumophila</i>	Respiratory infections
P71	Negative	<i>Prevotella bivia</i> ( <i>P. bivia</i> )	<i>P. bivia</i>	Abdominal infections
P72	Negative	<i>E. faecalis</i>	<i>E. faecalis</i> , <i>Candida glabrata</i> ( <i>C. glabrata</i> )	Abdominal infections
P73	Negative	<i>B. thetaiotaomicron</i>	<i>B. thetaiotaomicron</i> , <i>E. faecium</i>	Abdominal infections
P74	Negative	<i>E. coli</i>	<i>E. coli</i>	Urinary tract infections
P75	Negative	<i>Aspergillus fumigatus</i> ( <i>A. fumigatus</i> )	<i>A. fumigatus</i>	Septic thrombophlebitis
P76	Negative	<i>K. pneumoniae</i> , <i>E. faecalis</i> , <i>E. faecium</i> , <i>C. glabrata</i>	<i>K. pneumoniae</i> , <i>E. faecalis</i> , <i>E. faecium</i> , <i>C. glabrata</i>	Abdominal infections
P77	<i>S. enterica</i>	<i>S. enterica</i>	<i>S. enterica</i>	Infective endocarditis
P78	Negative	<i>E. faecium</i>	<i>E. faecium</i>	Abdominal infections
P79	Negative	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>	Abdominal infections
P80	Negative	<i>Proteus mirabilis</i>	<i>P. mirabilis</i>	Urinary tract infections

<sup>a</sup>, including native valve endocarditis, prosthetic valve endocarditis and pacemaker-related endocarditis; <sup>b</sup>, including infections related to central line catheters, artificial blood vessels and stent implantation. mNGS, metagenomic next-generation sequencing; tNGS, targeted next-generation sequencing; NA, not applicable; BSI, bloodstream infection.

**Table S3** Consistency between blood tNGS results and microbes detected by culture or mNGS in other specimens

Patient number	Underlying infectious syndromes	Blood tNGS results	Other specimens		
			Specimen type	Culture results	mNGS results
P16	Abdominal infections	<i>Klebsiella pneumoniae</i> ( <i>K. pneumoniae</i> )	Pus	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>
P18	Abdominal infections	<i>K. pneumoniae</i>	Pus	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>
P21	Abdominal infections	<i>K. pneumoniae</i>	Pus	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>
P23	Skin and soft tissue infections	<i>Staphylococcus aureus</i> ( <i>S. aureus</i> )	Pus	<i>S. aureus</i>	<i>S. aureus</i>
P24	Skin and soft tissue infections	<i>Aeromonas veronii</i> ( <i>A. veronii</i> )	Pus	Negative	<i>A. veronii</i>
P26	Skin and soft tissue infections	<i>Vibrio vulnificus</i> ( <i>V. vulnificus</i> )	Pus	Negative	<i>V. vulnificus</i>
P27	Urinary tract infections	<i>Escherichia coli</i> ( <i>E. coli</i> )	Urine	Negative	<i>E. coli</i>
P43	Respiratory infections	<i>K. pneumoniae</i>	Sputum	Negative	<i>K. pneumoniae</i>
P48	Abdominal infections	<i>E. coli</i>	Pus	Negative	<i>E. coli</i>
P49	Respiratory infections	<i>Prevotella oris</i> ( <i>P. oris</i> )	Pleural fluid	Negative	<i>P. oris</i>
P50	Respiratory infections	<i>Mycobacterium tuberculosis</i> ( <i>M. tuberculosis</i> )	Pleural fluid	<i>M. tuberculosis</i>	<i>M. tuberculosis</i>
P52	Skin and soft tissue infections	<i>Orientia tsutsugamushi</i> ( <i>O. tsutsugamushi</i> )	Pus	Negative	<i>O. tsutsugamushi</i>
P55	Respiratory infections	<i>Mycoplasma pneumoniae</i> ( <i>M. pneumoniae</i> )	Sputum	Negative	<i>M. pneumoniae</i>
P56	Abdominal infections	<i>Enterococcus faecalis</i> ( <i>E. faecalis</i> ), <i>Enterococcus faecium</i> ( <i>E. faecium</i> ), <i>K. pneumoniae</i>	Bile	<i>E. faecalis</i>	<i>E. faecalis</i> , <i>K. pneumoniae</i>
P59	Central nervous system infections	<i>Taenia solium</i> ( <i>T. solium</i> )	Cerebrospinal fluid	Negative	<i>T. solium</i>
P60	Skin and soft tissue infections	<i>Vibrio cholerae</i> ( <i>V. cholerae</i> )	Pus	Negative	<i>V. cholerae</i>
P62	Catheter/device related infections <sup>a</sup>	<i>Coxiella burnetii</i> ( <i>C. burnetii</i> )	Pus	Negative	<i>C. burnetii</i>
P63	Skin and soft tissue infections	<i>Aeromonas caviae</i> ( <i>A. caviae</i> )	Pus	<i>A. caviae</i>	<i>A. caviae</i>
P70	Respiratory infections	<i>Legionella pneumophila</i> ( <i>L. pneumophila</i> )	Sputum	Negative	<i>L. pneumophila</i>
P73	Abdominal infections	<i>Bacteroides thetaiotaomicron</i> ( <i>B. thetaiotaomicron</i> ), <i>E. faecium</i>	Peritoneal fluid	<i>E. faecium</i> , <i>E. coli</i>	<i>B. thetaiotaomicron</i> , <i>E. faecium</i> , <i>K. pneumoniae</i> , <i>Candida albicans</i>
P78	Abdominal infections	<i>E. faecium</i>	Peritoneal fluid	<i>E. faecium</i>	NA
P79	Abdominal infections	<i>K. pneumoniae</i>	Pus	<i>K. pneumoniae</i>	<i>K. pneumoniae</i>
P80	Urinary tract infections	<i>Proteus mirabilis</i> ( <i>P. mirabilis</i> )	Urine	Negative	<i>P. mirabilis</i>

<sup>a</sup>, including infections related to central line catheters, artificial blood vessels and stent implantation. tNGS, targeted next-generation sequencing; mNGS, metagenomic next-generation sequencing; NA, not applicable.