



Promoter methylation of *p16* and *DAPK* genes in brushing, blood, and tissue samples from patients with nasopharyngeal carcinoma: a systematic meta-analysis

Lifeng Xiao, Li Jiang, Qi Hu, Yuru Li

Department of Otolaryngology Head and Neck Surgery, First Affiliated Hospital of Harbin Medical University, Harbin 150001, China

Contributions: (I) Conception and design: L Xiao, Y Li; (II) Administrative support: Y Li; (III) Provision of study materials or patients: L Xiao, L Jiang, Q Hu; (IV) Collection and assembly of data: L Xiao, Q Hu, Y Li; (V) Data analysis and interpretation: L Xiao, L Jiang, Q Hu; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

Correspondence to: Yuru Li. Department of Otolaryngology Head and Neck Surgery, First Affiliated Hospital of Harbin Medical University, No23, Youzheng Street, Nangang District, Harbin 150001, China. Email: liyuru3131@sina.com.

Background: Promoter methylation of tumor suppressor genes (TSGs) *p16* and death-associated protein kinase (*DAPK*) has been reported in various carcinomas. However, the correlation between *p16* or *DAPK* promoter methylation and the detection of nasopharyngeal carcinoma (NPC) in different sample types remains unclear.

Methods: A systematic literature research was conducted to identify available studies. The pooled odds ratios (ORs) and their 95% confidence intervals (CIs) were calculated and analyzed.

Results: Eleven eligible papers on *p16* promoter methylation and seven eligible papers on *DAPK* promoter methylation were included in this analysis. The pooled OR of *p16* and *DAPK* promoter methylation was significantly higher in samples of NPC tissues than in nontumorous (OR =5.49, $P<0.001$; OR =17.51, $P<0.001$; respectively). Moreover, the pooled OR of *p16* promoter methylation was significantly higher in NPC than in normal blood and noncancerous brushing samples (OR =19.37, $P<0.001$; OR =15.03, $P<0.001$; respectively). In NPC samples, the OR of *DAPK* promoter methylation were significantly higher than in normal blood and normal brushing samples (OR =7.23, $P<0.001$; OR =42.00, $P=0.001$; respectively). No significant correlation was found between *p16* or *DAPK* promoter methylation and clinical stage ($P>0.1$).

Conclusions: Our findings suggested that *p16* and *DAPK* promoter methylation may be associated with the carcinogenesis of NPC. Promoter methylation of *p16* or *DAPK* may become a noninvasive biomarker for NPC detection in blood and brushing samples. *DAPK* or *p16* promoter methylation was not correlated with tumor stage. Conducting additional studies is essential in the future to confirm our results.

Keywords: *p16*; death-associated protein kinase (*DAPK*); methylation; nasopharyngeal carcinoma (NPC); biomarker; blood; brushing

Submitted Aug 18, 2016. Accepted for publication Oct 31, 2016.

doi: 10.21037/tcr.2016.12.08

View this article at: <http://dx.doi.org/10.21037/tcr.2016.12.08>

Introduction

Nasopharyngeal carcinoma (NPC) is a unique epithelial cancer of the head and neck with an extremely geographical and ethnic distribution. NPC is a prevalent malignancy in Southeast Asia, where it occurs in 20–30 per 100,000 persons every year; it is particularly prevalent in Southern China and ranges from 15 to 50 cases per 100,000

individuals, whereas it is rare in the Western countries, with an annual incidence rate of less than 1 per 100,000 people (1,2). Radiotherapy alone or in combination with chemotherapy has been used as the primary treatment strategy, providing a generally satisfactory disease control in patients with early-stage NPC (3). However, approximately 60–70% of the patients are diagnosed at later stages (III–IV)

with loco-regional lymph node metastases (4). Currently, distant metastasis is considered a typical failure pattern, particularly in advanced patients, with poor prognosis (5-7).

Previous studies have shown that certain etiological factors contribute to the initiation and development of NPC, including infection with Epstein-Barr virus, inflammation, dietary and lifestyle factors, and genetics and epigenetic alterations (8-11). Aberrant DNA methylation of tumor suppressor genes (TSGs), such as deleted in liver cancer 1 (*DLCL1*), is a frequent event in NPC (12,13). The human *p16* gene, mapped to chromosome 9p21 and consisting of 3 exons and 2 introns, is a cyclin-dependent kinase inhibitor that plays a key role in cell cycle regulation (14,15). Localized on human chromosome 9p34, death-associated protein kinase (*DAPK*), a serine/threonine kinase, participates in the modulation of a variety of cellular processes, including apoptosis, autophagy, and inflammation (16,17). The loss of *p16* and *DAPK* expression as TSGs through promoter methylation has been shown to be associated with many carcinomas (18-20).

Nonetheless, there are conflicting findings concerning the relationship between *p16* promoter methylation and NPC. For example, Challouf *et al.* reported the absence of significant association between *p16* promoter methylation and NPC (21), whereas the results of Nawaz *et al.* evidenced the presence of a significant association between *p16* promoter methylation and NPC (22). Similarly, there are controversial results on *DAPK* promoter methylation and its relation to NRC occurrence. For instance, Chang *et al.* established that the *DAPK* promoter methylation rate in NPC patients was similar to or even lower than that in healthy subjects (23). In contrast, the investigation of Tian *et al.* revealed that the level of *DAPK* promoter methylation was significantly higher in NPC cases than in healthy subjects (24). It is important to note that studies with a small sample size may lack strong statistical power (25,26). Therefore, to address these inconsistencies and evaluate the relationship between *p16* and *DAPK* promoter methylation and NPC, basing on several selection criteria, we integrated all eligible publications on this subject. In addition, we also determined whether *p16* and *DAPK* promoter methylation was linked with the clinical stage of NPC.

Methods

Search strategy and selection criteria

We conducted a comprehensive literature search of four

online electronic databases, including PubMed, EBSCO, Cochrane Library, and EMBASE, to identify eligible studies published before August 8th, 2016. We used the following combination of keywords and free words: 'nasopharyngeal cancer OR nasopharyngeal neoplasm OR nasopharyngeal carcinoma OR nasopharyngeal tumor OR NPC', 'p16 OR INK4A OR CDKN2A OR cyclin-dependent kinase inhibitor 2A', 'DAPK OR death-associated protein kinase OR DAP-kinase', 'methylation OR hypermethylation OR promoter methylation OR epigenetic'.

To be included in our meta-analysis, the articles had to meet the following inclusion criteria: (I) the diagnosis of NPC was based on histopathological examination; (II) case-control or cohort design studies published in English; (III) studies provided sufficient information with regard to the methylation rate of *p16* or *DAPK* promoter methylation to calculate the pooled odds ratios (ORs) and corresponding 95% confidence intervals (CIs); (IV) when identical or overlapping data were used in multiple publications, only the most recent paper or the paper with the largest population was included; (V) the sample type was without restriction and included tissue, blood, and brushing samples from patients with NPC. A tumor stage of ≤ 1 was considered as an early stage, whereas a tumor stage of ≥ 2 was defined as a later stage.

Data extraction

Two independent reviewers extracted the following data from each available study using selection standards, including the last name of the first author, publication year, country, ethnicity, sample type, the participants of cases and controls, method for detection of methylation, the frequency of *p16* or *DAPK* promoter methylation, *p16* or *DAPK* expression status. Nontumorous specimens were used as control samples. Any disagreements concerning study selection and data extraction were resolved by consensus or by a third reviewer.

Statistical analysis

In the current meta-analysis, statistical analysis of the pooled data was performed using Stata software, version 12.0 (STATA Corp., College Station, TX, USA). The pooled OR and corresponding 95% CI were calculated to determine the strength of the association between *p16* or *DAPK* promoter methylation and NPC risk. In

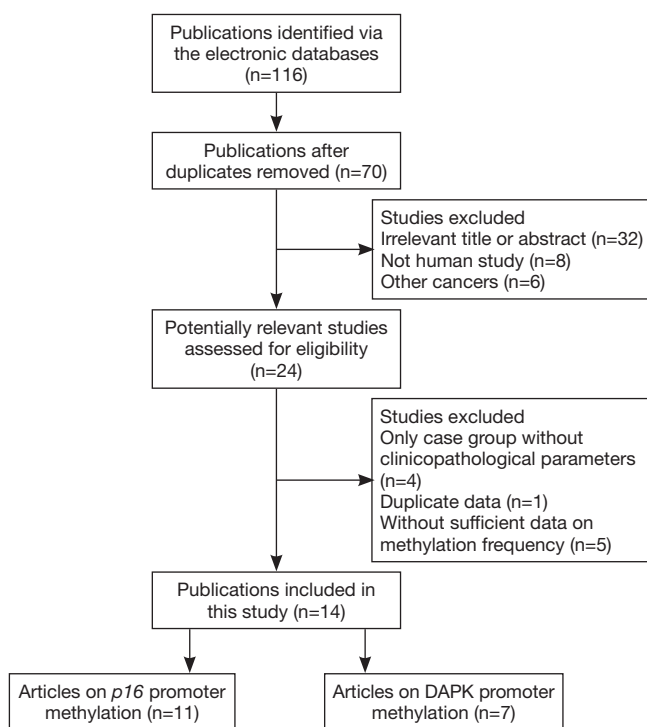


Figure 1 Flow diagram of the literature selection process.

addition, the correlation between *p16* or *DAPK* promoter methylation and the clinical stage of NPC was also established. The statistical heterogeneity among the studies was tested based on the Cochran's Q and I^2 tests (27). The random-effects model was employed when the P value was less than 0.1 in the Q -test, indicating the presence of substantial heterogeneity; otherwise, the fixed-effects model was applied when no evidence of the heterogeneity was observed (28,29). P value <0.05 was considered statistically significant.

Results

Study characteristics

Initially, a total of 116 potential articles were collected by an extensive search of the databases used (Figure 1). After selection based on the eligibility criteria, eleven available articles on *p16* promoter methylation (21-26,30-34) and seven eligible articles on *DAPK* promoter methylation (21,23,24,34-37) were eventually identified in this analysis. Seven of these studies evaluated the association between *p16* promoter methylation and NPC in NPC *vs.* nontumorous tissues (21-23,25,26,31,34). Three studies evaluated the

relationship between *p16* promoter methylation and NPC in NPC *vs.* normal blood samples (23,24,32). In addition, three studies estimated the connection between *p16* promoter methylation and NPC in NPC *vs.* nontumorous brushing samples (23,30,33). Six other studies assessed the correlation between *DAPK* promoter methylation and NPC in NPC *vs.* nontumorous tissues (21,23,34-37), and two studies evaluated the association between *DAPK* promoter methylation and NPC in NPC *vs.* normal blood samples (23,24). The relationship between *DAPK* promoter methylation and NPC in NPC *vs.* normal brushing samples was estimated in one study (23) and the correlation of *p16* promoter methylation with tumor stage in NPC in four (24,31-33). Two studies assessed the association of *DAPK* promoter methylation with tumor stage in NPC (24,37). Table 1 presents the main characteristics of the included studies.

Correlation of *p16* promoter methylation in cancer *vs.* controls

No obvious heterogeneity was observed for *p16* promoter methylation in cancer *vs.* controls (P value of the heterogeneity >0.1), and the fixed-effects model was applied. A higher frequency of *p16* promoter methylation in NPC than in nontumorous tissues (OR =5.49; 95% CI, 2.39–12.63; $P<0.001$) was established in seven studies with samples from 244 NPC and 61 nontumorous tissues (Figure 2).

We also evaluated the association between *p16* promoter methylation and the risk of NPC in fluid samples (blood and brushing samples). The results from three studies including 111 NPC and 127 normal blood samples demonstrated that *p16* promoter methylation in NPC samples was notably higher than in normal blood samples (OR =19.37; 95% CI, 4.45–84.26; $P<0.001$) (Figure 2). Three other studies with 111 NPC and 102 noncancerous brushing samples evidenced that *p16* promoter methylation was significantly higher in NPC samples than in noncancerous brushing samples (OR =15.03; 95% CI, 5.92–38.17; $P<0.001$) (Figure 2).

Therefore, our results revealed that *p16* promoter methylation was significantly correlated with the increased risk of NPC in tissue, blood, and brushing specimens.

Correlation of *DAPK* promoter methylation in cancer *vs.* controls

There was no substantial evidence of heterogeneity in

Table 1 The general characteristics of eligible studies

Gene	First author	Country	Ethnicity	Method	Sample	Control Sample	Case, N (M, %)	Control, N (M, %)	Stage 1-2, N [M]	Stage 3-4, N [M]	Expression
<i>p16</i>	Lo 1996 (26)	China	Asians	MPCR	Tissue	Normal	27 (22.2)	4 (0)	-	-	Loss
	Tong 2002 (33)	China	Asians	MSP	Brushing	Non-tumor	28 (46.4)	12 (0)	17 [7]	11 [6]	ND
	Kwong 2002 (34)	China	Asians	MSP	Tissue	Normal	33 (51.5)	6 (0)	-	-	ND
	Chang 2003 (23)	China	Asians	MSP	Blood	Normal	30 (0)	43 (0)	-	-	ND
	Chang 2003 (23)	China	Asians	MSP	Brushing	Normal	30 (16.7)	43 (0)	-	-	ND
	Chang 2003 (23)	China	Asians	MSP	Tissue	Normal	30 (33.3)	6 (0)	-	-	ND
	Wong 2003 (25)	China	Asians	MSP	Tissue	Normal	30 (23.3)	5 (0)	-	-	ND
	Wong 2004 (32)	China	Asians	MethylLight	Blood	Normal	41 (41.5)	43 (2.3)	24 [12]	17 [5]	ND
	Ayadi 2008 (31)	Tunisia	Caucasians	MSP	Tissue	Normal	44 (61.4)	3 (0)	32 [20]	12 [7]	ND
	Hutajulu 2011 (30)	The Netherlands	Caucasians	MSP	Brushing	Non-tumor	53 (66)	47 (12.8)	-	-	ND
	Challouf 2012 (21)	Tunisia	Caucasians	MSP	Tissue	Non-tumor	36 (33.3)	19 (21.0)	-	-	ND
	Tian 2013 (24)	China	Caucasians	MSP	Blood	Normal	40 (22.5)	41 (2.4)	36 [7]	4 [2]	ND
	Nawaz 2015 (22)	Sweden	Caucasians	MSP	Tissue	Non-tumor	44 (45.5)	18 (0)	-	-	ND
	<i>DAPK</i>	Kwong 2002 (34)	China	Asians	MSP	Tissue	Normal	33 (72.7)	6 (0)	-	-
Wong 2002 (37)		China	Asians	MSP	Tissue	Normal	32 (75.0)	3 (0)	25 [18]	7 [6]	ND
Chang 2003 (23)		China	Asians	MSP	Blood	Normal	30 (3.3)	43 (2.3)	-	-	ND
Chang 2003 (23)		China	Asians	MSP	Brushing	Normal	30 (50.0)	43 (2.3)	-	-	ND
Chang 2003 (23)		China	Asians	MSP	Tissue	Normal	30 (76.7)	6 (0)	-	-	ND
Kong 2006 (36)		China	Asians	MSP	Tissue	Non-tumor	46 (76.1)	6 (0)	-	-	Loss
Fendri 2009 (35)		Tunisia	Caucasians	MSP	Tissue	Normal	68 (88.2)	9 (0)	-	-	Loss
Challouf 2012 (21)		Tunisia	Caucasians	MSP	Tissue	Non-tumor	36 (47.2)	19 (15.8)	-	-	ND
Tian 2013 (24)		China	Caucasians	MSP	Blood	Normal	35 (51.4)	41 (9.8)	36 [16]	4 [2]	ND

DAPK, death-associated protein kinase; “-” stands for no data; MSP: methylation-specific polymerase chain reaction; MPCR, multiplex PCR; M: methylation positive status; N, the number of methylation and unmethylation; ND, not done.

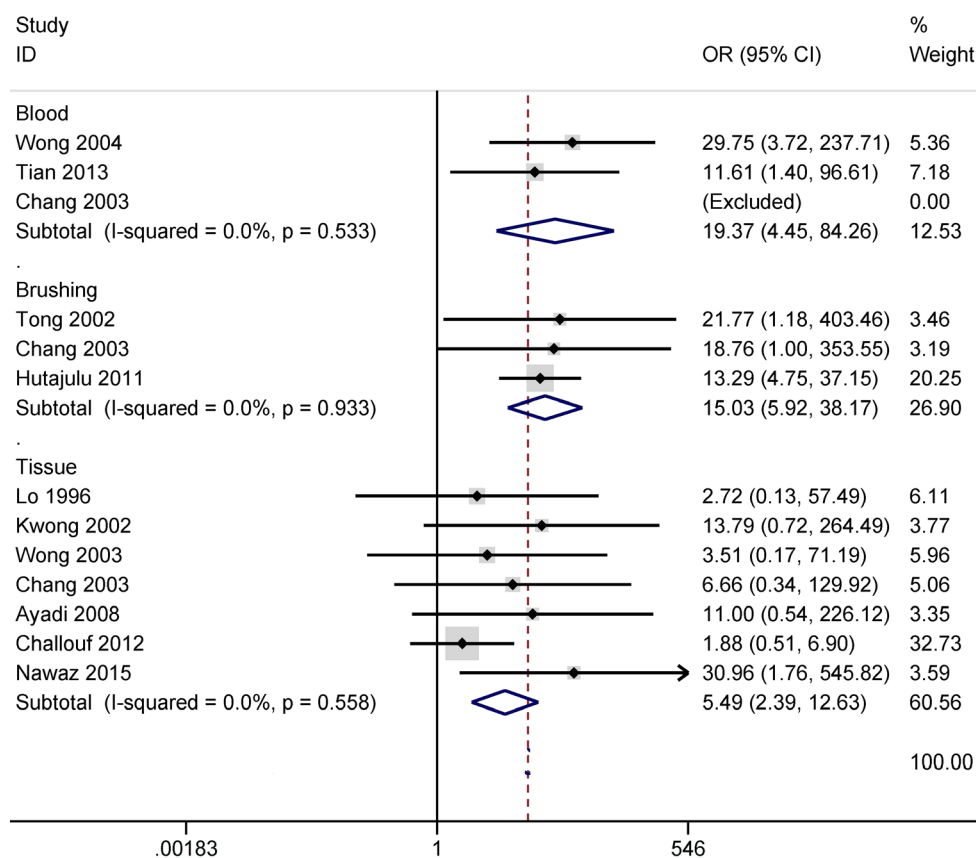


Figure 2 Forest plot for the correlation of *p16* promoter methylation the risk of NPC showing the pooled OR obtained by the fixed-effects model in cancer vs. controls. Tissue: OR =5.49, P<0.001, including seven studies with 244 NPC and 61 nontumorous tissues; blood: OR =19.37, P<0.001, including three studies with 111 NPC and 127 normal blood samples; brushing: OR =15.03, P<0.001, including three studies with 111 NPC and 102 noncancerous brushing samples.

cancer vs. controls; thus, the fixed-effects model was used for *DAPK* promoter methylation (P value of the heterogeneity >0.1). The results concerning six studies with 245 NPC and 49 nontumorous tissue samples indicated that a higher *DAPK* promoter methylation rate was observed in NPC tissues in comparison to that in nontumorous tissues (OR =17.51; 95% CI, 7.08–43.32; P<0.001) (Figure 3).

Additionally, the pooled OR of two studies with 65 NPC and 84 normal blood samples showed that *DAPK* promoter methylation was significantly greater in NPC than in normal blood samples (OR =7.23; 95% CI, 2.44–21.45; P<0.001) (Figure 3). The combined OR in one study exhibited a significant association between *DAPK* promoter methylation and NPC based on the results from the analysis of 30 NPC and 43 normal brushing samples (OR =42.00; 95% CI, 5.10–345.85; P=0.001) (Figure 3).

Our findings revealed that the methylation status of

DAPK promoter was significantly associated with the increased risk of NPC in tissue, blood, and brushing specimens. However, the analyses of brushing and blood samples should be cautiously interpreted as only one or two studies with small sample sizes were involved in this meta-analysis.

Subgroup analysis by ethnic population in cancer vs. nontumorous tissues

Further, we conducted subgroup analysis by ethnicity to evaluate the different degree of association of the stratified population in tissue samples. Subgroup analysis based on ethnicity showed that *p16* promoter methylation was significantly correlated with an increased risk of NPC in both among both populations investigated, Asians (OR =5.90; 95% CI, 1.35–25.82; P=0.018) and Caucasians

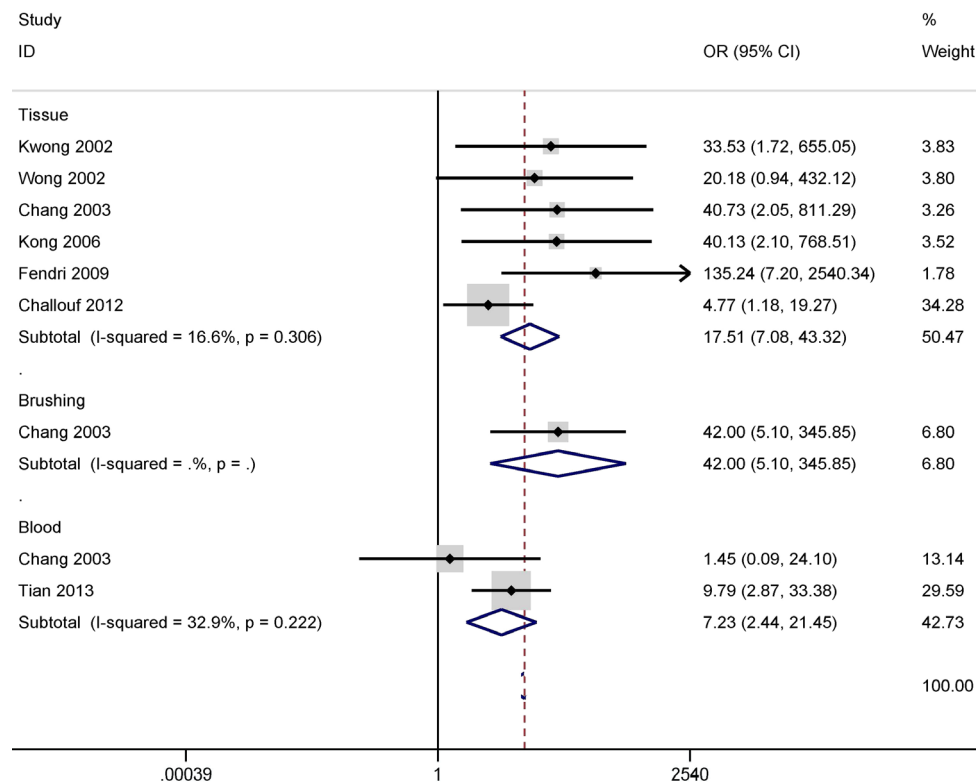


Figure 3 Forest plot for the association of *DAPK* promoter methylation with the risk of NPC displaying the pooled OR under the fixed-effects model in cancer *vs.* controls. Tissue: OR =17.51, $P < 0.001$, including 6 studies with 245 NPC and 49 nontumorous tissues; blood: OR =7.23, $P < 0.001$, including 2 studies with 65 NPC and 84 normal blood samples; brushing: OR =42.00, $P = 0.001$, including 1 study with 30 NPC and 43 normal brushing samples.

(OR =5.28; 95% CI, 1.93–14.43; $P = 0.001$) (Figure 4).

DAPK promoter methylation also was significantly correlated with the increased risk of NPC in the Asian (OR =33.25; 95% CI, 7.48–147.89; $P < 0.001$) and the Caucasian population (OR =11.21; 95% CI, 3.58–35.09; $P < 0.001$) (Figure 5). However, the results of subgroup analysis should be carefully considered as only small sample sizes were included in this study, especially in the Caucasian population subgroup.

Correlation of *p16* or *DAPK* promoter methylation with tumor stage in NPC

We also determined whether *p16* or *DAPK* promoter methylation was correlated with tumor stage in NPC using the fixed-effects model (Figure 6). The overall OR from four studies involving 109 advanced NPC patients and 44 early NPC patients demonstrated that *p16* promoter methylation was not correlated with tumor stage (OR =1.06; 95% CI,

0.51–2.20; $P = 0.876$). The overall OR from two other studies including 61 advanced NPC patients and 11 early NPC patients indicated that *DAPK* promoter methylation was not associated with tumor stage (OR =0.59; 95% CI, 0.13–2.65; $P = 0.491$). Nonetheless, the results regarding the relationship between *p16* or *DAPK* promoter methylation and tumor stage should be cautiously interpreted as only a small sample size of NPC patients were analyzed in the current research.

Discussion

The hypermethylation of TSGs and hypomethylation of oncogenes are two important molecular mechanisms in epigenomic regulation that play key roles in the initiation and progression of cancer (38–40). The promoter methylation of TSGs may affect cell proliferation, cell death, cell migration, and cell invasion (41). Aberrant promoter methylation of *p16* and *DAPK* genes has been

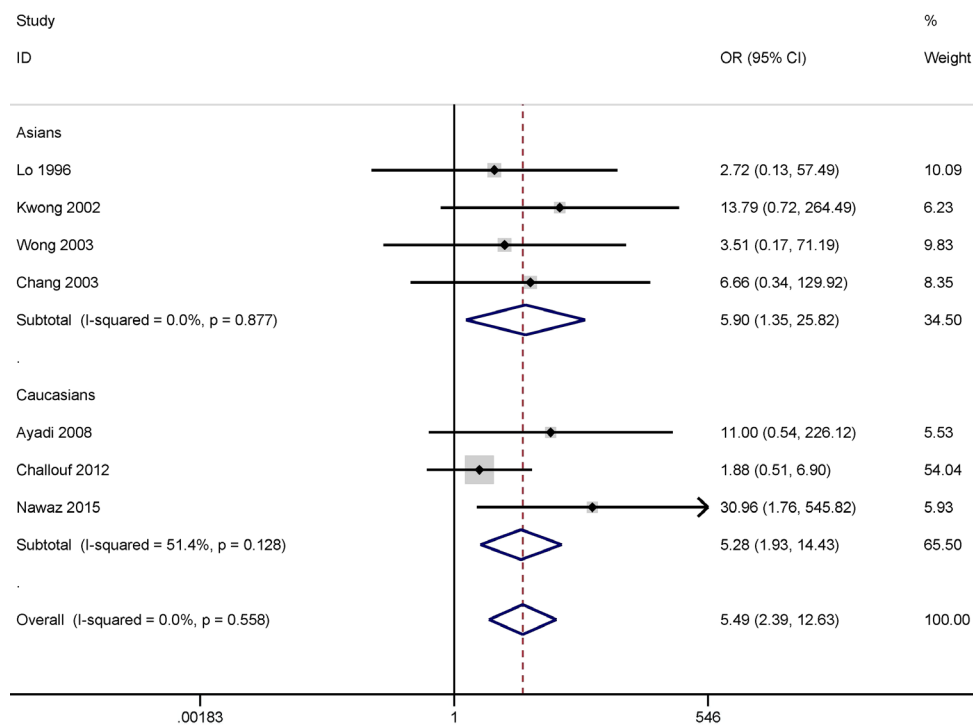


Figure 4 Forest plot for the correlation of *p16* promoter methylation with the risk of NPC by ethnic subgroup illustrating the pooled OR under the fixed-effects model in cancer *vs.* controls. Asians: OR =5.90, $P=0.018$; Caucasians: OR =5.28, $P=0.001$.

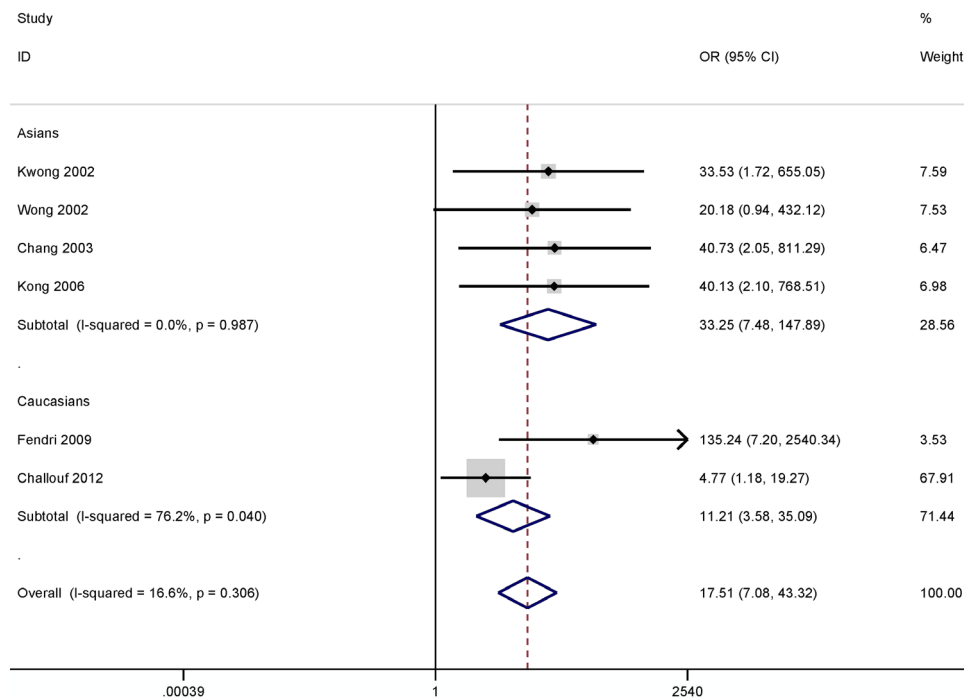


Figure 5 Forest plot for the correlation of *DAPK* promoter methylation with the risk of NPC by ethnic subgroup showing the pooled OR under the fixed-effects model in cancer *vs.* controls. Asians: OR =33.25, $P<0.001$; Caucasians: OR =11.21, $P<0.001$.

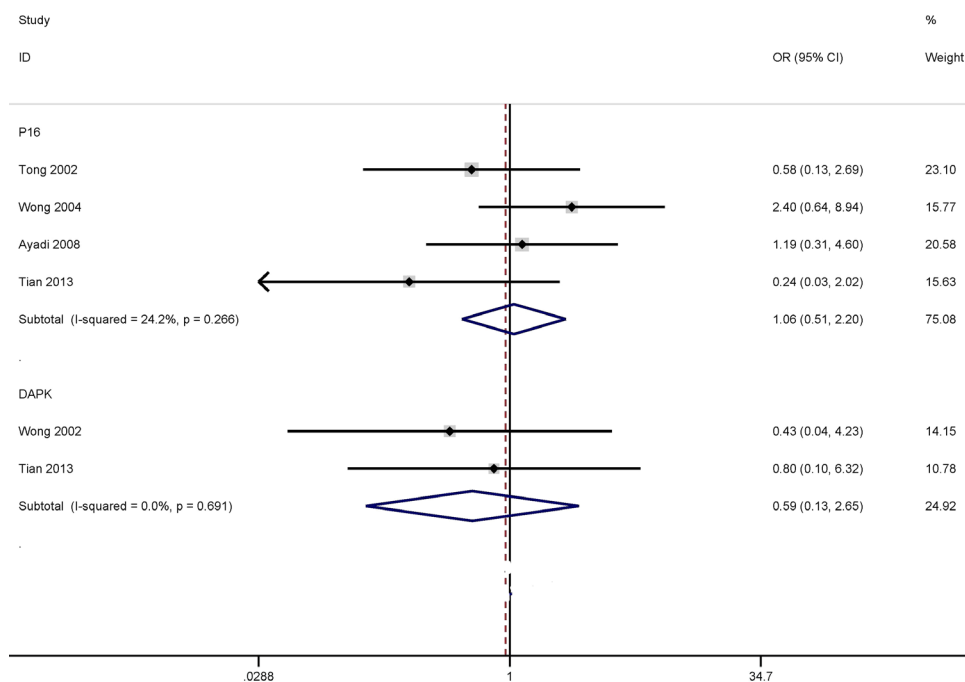


Figure 6 Forest plot for the association of *p16* and *DAPK* promoter methylation with tumor stage showing the pooled OR under the fixed-effects model in cancer. *p16*: OR =1.06, P=0.876, including four studies with 109 advanced NPC patients and 44 early NPC patients; *DAPK*: OR =0.59, P=0.491, including two studies with 61 advanced NPC patients and 11 early NPC patients.

reported in NPC (21,23,34). As a key cell cycle regulator, the *p16* gene is involved in the inhibition of cell cycle progression, and the restoration of its expression may induce G0/G1 arrest and suppress the tumorigenic growth of NPC cells (42). A correlation was found between *p16* promoter methylation and its expression in NPC, with the absence of *p16* expression (26). Earlier studies showed that the loss of *DAPK* expression was associated with its promoter methylation in NPC (35,36). The inactivation of *p16* and *DAPK* through promoter methylation may play a key in NPC tumorigenesis (43).

However, there are still inconsistent and controversial results regarding the methylation rate of *p16* and *DAPK* promoter in NPC specimens. For example, different promoter methylation rates of the *p16* gene, ranging from 22.2% (26) to 61.4% (31), were established in NPC tissues. Inconsistent findings on the frequency of *DAPK* promoter methylation in NPC tissues, which ranged from 47.2% (21) to 88.2% (35) were also reported previously. Therefore, we systematically investigated studies of *p16* and *DAPK* promoter methylation in NPC samples to estimate the association between *p16* and *DAPK* promoter methylation and NPC.

p16 and *DAPK* promoter methylation rates were shown to be significantly higher in NPC than in nontumorous tissue samples from the nasopharynx, suggesting that *p16* or *DAPK* promoter methylation may play a pivotal role in the tumorigenesis of NPC. No significant heterogeneity was found in our study, indicating the stability of our results.

The subgroup analysis by ethnic population comparing *p16* and *DAPK* promoter methylation in NPC and nontumorous tissues from the nasopharynx revealed that the promoter methylation of *p16* or *DAPK* was significantly correlated with the risk of NPC in both the Asian and Caucasian populations, indicating that *p16* and *DAPK* promoter methylation may be susceptible genes for Asian and Caucasian populations. In addition, the Asian population had a higher OR of *DAPK* promoter methylation than the Caucasian population subgroup (33.25 vs. 11.21), which suggested that the Asian population may be more susceptible to *DAPK* promoter methylation. However, due to the limitation of the small sample size, the analysis of the Caucasian population subgroup in the current study should be cautiously interpreted.

Chang *et al.* reported that the promoter methylation rate of *p16* was 0% in NPC blood samples and 16.7% in NPC

brushing samples, respectively (23). On the other hand, Wong *et al.* reported that a *p16* promoter methylation rate of 41.5% detected in NPC blood samples (32), and Hutajulu *et al.* discovered that *p16* promoter had a methylation frequency of 66% in NPC brushing samples (30). Detection of DNA methylation in brushing or blood samples, shows a great potential to be applied as an invasive biomarker for early detection of NPC (44). Previous studies suggest that *p16* and *DAPK* promoter methylation identified in brushing or blood samples may become a useful biomarker in NPC (24,33). Our findings demonstrate that *p16* promoter methylation established in blood and brushing samples is significantly associated with the risk of NPC and its rate is significantly higher in blood or brushing samples of NPC patients than in those of healthy subjects, suggesting that *p16* promoter methylation may be used as a noninvasive biomarker for NPC detection in blood and brushing samples.

Chang *et al.* discovered that *DAPK* promoter methylation frequency was 3.3% in NPC blood samples (23), but a much higher value (51.4%) was reported by Tian *et al.* (24). On the other hand, a frequency of *DAPK* promoter methylation in NPC brushing samples that reaches 50% has been reported only in the study of Chang *et al.* (23). Our results indicated that *DAPK* promoter methylation in blood and brushing samples was significantly correlated with the risk of NPC and its level was significantly higher in the blood or brushing samples of NPC patients than in those of healthy subjects, indicating that *DAPK* promoter methylation is a potential noninvasive biomarker for the detection of NPC in blood or brushing samples. Additionally, Tian *et al.* evidenced that the combined *P16* and *DAPK* promoter methylation did not significantly increase the potential capacity for NPC detection, but the combination of four-gene marker can be applied as a promising tool for the diagnosis of NPC (24). However, more clinical research studies are required to further validate these findings in the future. Only the OR of one study with brushing samples was included in this study. Thus, we should carefully consider the results from the analysis of blood and brushing samples conducted for the detection of *p16* and *DAPK* promoter methylation. Additional studies with larger sample sizes are needed to confirm our results.

Finally, we also investigated whether *p16* or *DAPK* promoter methylation was correlated with the clinical stage of NCR and found that *p16* and *DAPK* promoter methylation was not associated with tumor stage. More studies are exceedingly essential to validate that finding in

the future.

Some limitations of this meta-analysis should be acknowledged. First, only articles published in English were included in this research. Articles in other languages and publications of other types, such as conferences abstracts, were excluded due to their unreadable contents or insufficient information, which might have led to a selection bias. Second, due to the limitation of insufficient data, we did not assess the relationship between *p16* and *DAPK* promoter methylation and other clinicopathological features, such as tumor grade, sex status, and lymph node status. Third, further studies with larger sample sizes should be done to validate our results, especially those concerning *DAPK* promoter methylation in blood and brushing specimens.

In conclusion, our findings suggest that *p16* and *DAPK* promoter methylation may play an important role in NPC development. The promoter methylation levels of *p16* or *DAPK* are potential useful biomarkers for NPC detection in blood and brushing samples in clinical settings. No significant association was found between *p16* or *DAPK* promoter methylation and tumor stage. Due to the limitations of the sample size in the present analysis, further large-scale studies with larger sample sizes of subjects are necessary to investigate more comprehensively the clinical effects of *p16* and *DAPK* promoter methylation in NPC patients.

Acknowledgments

Funding: None.

Footnote

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/tcr.2016.12.08>). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-

commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

References

1. Wu JM, Yu TJ, Yeh SA, et al. Use dose bricks concept to implement nasopharyngeal carcinoma treatment planning. *Biomed Res Int* 2014;2014:720876.
2. Jemal A, Bray F, Center MM, et al. Global cancer statistics. *CA Cancer J Clin* 2011;61:69-90.
3. Yi JL, Gao L, Huang XD, et al. Nasopharyngeal carcinoma treated by radical radiotherapy alone: Ten-year experience of a single institution. *Int J Radiat Oncol Biol Phys* 2006;65:161-8.
4. Mao YP, Xie FY, Liu LZ, et al. Re-evaluation of 6th edition of AJCC staging system for nasopharyngeal carcinoma and proposed improvement based on magnetic resonance imaging. *Int J Radiat Oncol Biol Phys* 2009;73:1326-34.
5. Chen L, Hu CS, Chen XZ, et al. Concurrent chemoradiotherapy plus adjuvant chemotherapy versus concurrent chemoradiotherapy alone in patients with locoregionally advanced nasopharyngeal carcinoma: a phase 3 multicentre randomised controlled trial. *Lancet Oncol* 2012;13:163-71.
6. Ma BB, Chan AT. Systemic treatment strategies and therapeutic monitoring for advanced nasopharyngeal carcinoma. *Expert Rev Anticancer Ther* 2006;6:383-94.
7. Liu MT, Hsieh CY, Chang TH, et al. Prognostic factors affecting the outcome of nasopharyngeal carcinoma. *Jpn J Clin Oncol* 2003;33:501-8.
8. Zhang Y, Zhou GQ, Liu X, et al. Exploration and Validation of C-Reactive Protein/Albumin Ratio as a Novel Inflammation-Based Prognostic Marker in Nasopharyngeal Carcinoma. *J Cancer* 2016;7:1406-12.
9. Tsao SW, Yip YL, Tsang CM, et al. Etiological factors of nasopharyngeal carcinoma. *Oral Oncol* 2014;50:330-8.
10. Xue WQ, Qin HD, Ruan HL, et al. Quantitative association of tobacco smoking with the risk of nasopharyngeal carcinoma: a comprehensive meta-analysis of studies conducted between 1979 and 2011. *Am J Epidemiol* 2013;178:325-38.
11. Lo KW, To KF, Huang DP. Focus on nasopharyngeal carcinoma. *Cancer Cell* 2004;5:423-8.
12. Bruce JP, Yip K, Bratman SV, et al. Nasopharyngeal Cancer: Molecular Landscape. *J Clin Oncol* 2015;33:3346-55.
13. Feng X, Ren C, Zhou W, et al. Promoter hypermethylation along with LOH, but not mutation, contributes to inactivation of DLC-1 in nasopharyngeal carcinoma. *Mol Carcinog* 2014;53:858-70.
14. Piepkorn M. Melanoma genetics: an update with focus on the CDKN2A(p16)/ARF tumor suppressors. *J Am Acad Dermatol* 2000;42:705-22; quiz 23-6.
15. Lukas J, Parry D, Aagaard L, et al. Retinoblastoma-protein-dependent cell-cycle inhibition by the tumour suppressor p16. *Nature* 1995;375:503-6.
16. Huang Y, Chen L, Guo L, et al. Evaluating DAPK as a therapeutic target. *Apoptosis* 2014;19:371-86.
17. Cohen O, Kimchi A. DAP-kinase: from functional gene cloning to establishment of its role in apoptosis and cancer. *Cell Death Differ* 2001;8:6-15.
18. Misawa K, Mochizuki D, Imai A, et al. Prognostic value of aberrant promoter hypermethylation of tumor-related genes in early-stage head and neck cancer. *Oncotarget* 2016;7:26087-98.
19. Laskar RS, Ghosh SK, Talukdar FR. Rectal cancer profiling identifies distinct subtypes in India based on age at onset, genetic, epigenetic and clinicopathological characteristics. *Mol Carcinog* 2015;54:1786-95.
20. Brait M, Loyo M, Rosenbaum E, et al. Correlation between BRAF mutation and promoter methylation of TIMP3, RARBeta2 and RASSF1A in thyroid cancer. *Epigenetics* 2012;7:710-9.
21. Challouf S, Ziadi S, Zaghoudi R, et al. Patterns of aberrant DNA hypermethylation in nasopharyngeal carcinoma in Tunisian patients. *Clin Chim Acta* 2012;413:795-802.
22. Nawaz I, Moumad K, Martorelli D, et al. Detection of nasopharyngeal carcinoma in Morocco (North Africa) using a multiplex methylation-specific PCR biomarker assay. *Clin Epigenetics* 2015;7:89.
23. Chang HW, Chan A, Kwong DL, et al. Evaluation of hypermethylated tumor suppressor genes as tumor markers in mouth and throat rinsing fluid, nasopharyngeal swab and peripheral blood of nasopharyngeal carcinoma patient. *Int J Cancer* 2003;105:851-5.
24. Tian F, Yip SP, Kwong DL, et al. Promoter hypermethylation of tumor suppressor genes in serum as potential biomarker for the diagnosis of nasopharyngeal carcinoma. *Cancer Epidemiol* 2013;37:708-13.
25. Wong TS, Tang KC, Kwong DL, et al. Differential gene methylation in undifferentiated nasopharyngeal carcinoma. *Int J Oncol* 2003;22:869-74.

26. Lo KW, Cheung ST, Leung SF, et al. Hypermethylation of the p16 gene in nasopharyngeal carcinoma. *Cancer Res* 1996;56:2721-5.
27. Coory MD. Comment on: Heterogeneity in meta-analysis should be expected and appropriately quantified. *Int J Epidemiol* 2010;39:932; author reply 933.28.
28. Higgins JP, Thompson SG, Deeks JJ, et al. Measuring inconsistency in meta-analyses. *BMJ* 2003;327:557-60.
29. DerSimonian R. Meta-analysis in the design and monitoring of clinical trials. *Stat Med* 1996;15:1237-48; discussion 49-52.
30. Hutajulu SH, Indrasari SR, Indrawati LP, et al. Epigenetic markers for early detection of nasopharyngeal carcinoma in a high risk population. *Mol Cancer* 2011;10:48.
31. Ayadi W, Karray-Hakim H, Khabir A, et al. Aberrant methylation of p16, DLEC1, BLU and E-cadherin gene promoters in nasopharyngeal carcinoma biopsies from Tunisian patients. *Anticancer Res* 2008;28:2161-7.
32. Wong TS, Kwong DL, Sham JS, et al. Quantitative plasma hypermethylated DNA markers of undifferentiated nasopharyngeal carcinoma. *Clin Cancer Res* 2004;10:2401-6.
33. Tong JH, Tsang RK, Lo KW, et al. Quantitative Epstein-Barr virus DNA analysis and detection of gene promoter hypermethylation in nasopharyngeal (NP) brushing samples from patients with NP carcinoma. *Clin Cancer Res* 2002;8:2612-9.
34. Kwong J, Lo KW, To KF, et al. Promoter hypermethylation of multiple genes in nasopharyngeal carcinoma. *Clin Cancer Res* 2002;8:131-7.
35. Fendri A, Masmoudi A, Khabir A, et al. Inactivation of RASSF1A, RARbeta2 and DAP-kinase by promoter methylation correlates with lymph node metastasis in nasopharyngeal carcinoma. *Cancer Biol Ther* 2009;8:444-51.
36. Kong WJ, Zhang S, Guo CK, et al. Effect of methylation-associated silencing of the death-associated protein kinase gene on nasopharyngeal carcinoma. *Anticancer Drugs* 2006;17:251-9.
37. Wong TS, Chang HW, Tang KC, et al. High frequency of promoter hypermethylation of the death-associated protein-kinase gene in nasopharyngeal carcinoma and its detection in the peripheral blood of patients. *Clin Cancer Res* 2002;8:433-7.
38. Franco R, Schoneveld O, Georgakilas AG, et al. Oxidative stress, DNA methylation and carcinogenesis. *Cancer Lett* 2008;266:6-11.
39. Corson TW, Gallie BL. One hit, two hits, three hits, more? Genomic changes in the development of retinoblastoma. *Genes Chromosomes Cancer* 2007;46:617-34.
40. Bodmer WF. 1998 Runme Shaw Memorial Lecture: somatic evolution of cancer. *Ann Acad Med Singapore* 1999;28:323-9.
41. Maziveyi M, Alahari SK. Breast Cancer Tumor Suppressors: A Special Emphasis on Novel Protein Nischarin. *Cancer Res* 2015;75:4252-9.
42. Wang GL, Lo KW, Tsang KS, et al. Inhibiting tumorigenic potential by restoration of p16 in nasopharyngeal carcinoma. *Br J Cancer* 1999;81:1122-6.
43. Lo KW, Chung GT, To KF. Deciphering the molecular genetic basis of NPC through molecular, cytogenetic, and epigenetic approaches. *Semin Cancer Biol* 2012;22:79-86.
44. Yang X, Dai W, Kwong DL, et al. Epigenetic markers for noninvasive early detection of nasopharyngeal carcinoma by methylation-sensitive high resolution melting. *Int J Cancer* 2015;136:E127-35.

Cite this article as: Xiao L, Jiang L, Hu Q, Li Y. Promoter methylation of p16 and DAPK genes in brushing, blood, and tissue samples from patients with nasopharyngeal carcinoma: a systematic meta-analysis. *Transl Cancer Res* 2016;5(6):827-837. doi: 10.21037/tcr.2016.12.08