

KDM1 links nuclear GSK3β to epigenetic alterations

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Submitted Jan 21, 2017. Accepted for publication Feb 24, 2017. doi: 10.21037/tcr.2017.03.26 **View this article at:** http://dx.doi.org/10.21037/tcr.2017.03.26

Accumulating evidence indicates that cancer stem cells (CSCs) play an important role during the processes of tumor development, progression, and recurrence and are primarily responsible for therapeutic resistance and poor clinical outcome of patients (1). Epigenetic alterations, particularly histone methylation, have been increasingly recognized as a global transcriptional regulator that contributes to stem cell self-renewal and differentiation under physiological and pathological conditions (2). For example, methylations of Lys9 and Lys27 residues of histone H3 (H3K9me2/3 and H3K27me3), associate with heterochromatin and transcriptional repression; whereas H3K4me2/3, often found in active gene promoters, is associated with transcriptional activation. These epigenetic modifications create unique promoter architectures that control gene expression. Glycogen synthase kinase 3 beta $(GSK3\beta)$ has been shown to take part in the regulation of histone methylation including H3K4 methylation in the promoter regions of multiple genes (3,4). However, the molecular mechanisms for GSK3^β in mediating alterations of histone methylation remain to be defined. In September 2016 issue of Nature Cell Biology, Zhou et al. (5) addressed this challenge and singled out a connection between nuclear GSK3β and epigenetic aberration via regulation of lysinespecific histone demethylase 1A (KDM1A) stability.

Histone lysine methylation is a dynamic and reversible process. KDM1A (also known as LSD1) is the first identified lysine-specific histone demethylase that selectively remove the methyl group from H3K4me1/2 through flavin-adeninedinucleotide (FAD)-dependent oxidative reaction (6). KDM1A exerts its gene repression functions by forming as a key catalytic component in several corepressor complexes, including Co-REST, NuRD, CtBP, HDAC, or SIRT1 (7,8). KDM1A is often overexpressed in breast (9), bladder, lung, colon (10) cancers and glioblastoma (11), and is required for the maintenance and differentiation of stem cells (12) and CSCs (13). Despite its determinant roles in stem cell pluripotency and differentiation and tumorigenesis, the mechanisms that lead to KDM1A dysregulation in these tumors remain to be better delineated.

Consistent with the notion that KDM1A is a labile protein that under constant protein ubiquitination and degradation (6,9), Zhou *et al.* found that nuclear GSK3 β stabilizes KDM1A by decreasing its ubiquitylation. GSK3β prefers substrates that are primed phosphorylated by another kinase at a Serine or Threonine residue located 4 or 5 amino acids downstream or upstream of GSK3β phosphorylation site (14). The authors found that KDM1A contains several potential GSK3^β phosphorylation sites harboring a canonical -S/TxxxS/T-motif. They further identified that Ser687 of KDM1A is phosphorylated by casein kinase 1 alpha (CK1 α) and this priming phosphorylation facilitates GSK3ß phosphorylation at Ser683. As GSK3β does not directly deubiquitylate KDM1A, the authors proposed that there should be deubiquitylase (DUB) to cooperate the deubiquitylation of KDM1A. Thus, they screened a panel of DUBs and identified ubiquitin-specific protease 22 (USP22) as a direct DUB for KDM1A. Subsequently, the authors explored the roles of GSK3β-USP22-KDM1A axis in glioma. They showed that GSK3β-USP22-KDM1A axis is required for tumorigenesis of CSCs and associated with the grade of

Translational Cancer Research, Vol 6, Suppl 2 March 2017

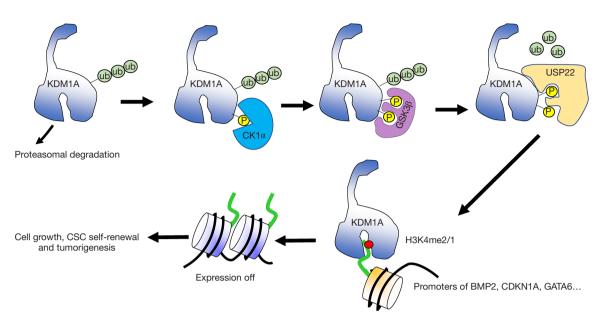


Figure 1 Schematic illustration of GSK3β-USP22-KDM1A axis in the regulation of tumorigenesis. Ubiqutylated KDM1A either enters the proteasomal degradation. Alternatively, nuclear GSK3β phosphorylates KDM1A after the priming phosphorylation by CK1α. Phosphorylation of KDM1A by GSK3β enhances its binding with and deubiquitylation by USP22, leading to its stabilization. Stabilized KDM1A promotes demethylation of histone H3K4 and suppresses the expression of *BMP2*, *CDKN1A* and *GATA6*, which in turn favors tumorigenesis.

human glioma malignance.

A picture appears in which KDM1A connects nuclear GSK3 β with epigenetics (*Figure 1*). First, KDM1A is phosphorylated by CK1a at Ser687 which facilitates its binding with and phosphorylation by GSK3^β. Subsequently, USP22, a direct KDM1A DUB, deubiquitylates KDM1A. Consequently, stabilized KDM1A removes the activation marks on H3K4 in promoter regions of stem cell pluripotency inhibition genes, including BMP2, CDKN1A and GATA6. Finally, as reported here, silencing of these tumor suppressors promotes tumorigenesis of CSCs. More importantly, the authors also showed therapeutic potential of tideglusib, a selective non-ATP-competitive GSK3 inhibitor with good blood-brain barrier penetration and now in a phase II trial for the treatment of Alzheimer's disease, along or in combination with temozolomide, a FDA approved drug for glioma. Although accumulating numbers of oncogenes have been extensively characterized, many of these oncogenes are not good drug targets. Thus, there are urgent needs for alternative therapeutic approaches to interfere these oncogenes. As KDM1A is structurally related monoamine oxidases A & B (MAO-A and MAO-B), a KDM1A inhibitor may exhibit side-effect by suppressing

MAO-A and MAO-B. In addition, blood-brain barrier makes it harder for KDM1A as a therapeutic target for treating glioma. Thus, targeting the proposed GSK3β-USP22-KDM1A signaling axis represents an excellent approach in suppressing the function of KDM1A in glioma. Recent works suggest that onco-DUBs, which contribute substantially to the stability of onco-proteins and pathways and are amenable to pharmacologic inhibition by small molecules, possess great potential for cancer treatment (15). Though not explored in their work, USP22, may also serve as a promising target for treating glioma and deserve further investigation. Nevertheless, the therapeutic potential established in this outstanding study will warrant further translational investigation and inspire many areas of basic inquiry that will further our understanding of the concerted connections between aberrant cell signaling and epigenetics during malignant tumor progression.

Acknowledgments

Funding: This work was supported by the Shared Resources of the University of Kentucky Markey Cancer Center (P30CA177558), grants from NIH (CA125454 and

CA188118), DoD (BC140733P1), and Mary Kay Ash Foundation.

Footnote

Provenance and Peer Review: This article was commissioned and reviewed by the Section Editor Chen Qian (Center for Inflammation & Epigenetics, Houston Methodist Hospital Research Institute, Houston, TX, USA).

Conflicts of Interest: Both authors have completed the ICMJE uniform disclosure form (available at http://dx.doi. org/10.21037/tcr.2017.03.26). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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Cite this article as: Guo W, Zhou BP. KDM1 links nuclear GSK3β to epigenetic alterations. Transl Cancer Res 2017;6(Suppl 2):S366-S368. doi: 10.21037/tcr.2017.03.26 Inhibition of glycogen synthase kinase-3 activity leads to epigenetic silencing of nuclear factor kappaB target genes and induction of apoptosis in chronic lymphocytic leukemia B cells. Blood 2007;110:735-42.

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