

Identification of novel candidate genes and small molecule drugs in ovarian cancer by bioinformatics strategy

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Background: Ovarian cancer (OC) is the most lethal type of malignancies in the female reproductive system. This study aimed to identify novel biomarkers and potential small molecule drugs in OC by integrating two expression profile datasets.

Methods: GSE18520 and GSE14407 from the Gene Expression Omnibus (GEO) database were selected and the overlapped differentially expressed genes (DEGs) were detected. The Gene Ontology (GO) analysis and Kyoto Encyclopedia of Genes and Genome (KEGG) pathway enrichment analysis were performed to establish the protein-protein interaction (PPI) network of DEGs and identified the hub genes. Gene Expression Profiling Interactive Analysis (GEPIA), Oncomine database and The Human Protein Atlas (HPA) were used to validate the expression of the identified hub genes. The prognostic value of these hub genes were evaluated by the Kaplan Meier plotter online tool. The expression of NCAPG was further explored by immunohistochemistry in our OC tissues. Moreover, CMap database was used to look for prospective small compounds with therapeutic efficacy based on OC RNA-seq.

Results: A total of 433 DEGs were identified. The DEGs were mainly enriched in negative regulation of transcription and pathways in cancer. A PPI network was constructed with 344 nodes and 1,596 interactions. The top ten module genes were chosen as hub genes. Among which, survival analysis showed that patients with high expression of *CCNB1*, *TOP2A*, *NUSAP1*, *NCAPG*, *KIF20A* and *DLGAP5* had poorer survival results than those with low expression. These six genes were all overexpressed in OC tissue by means of bioinformatics analysis. In our clinical patients, the expression rate of NCAPG in OC tissues was significantly higher than that in benign serous ovarian cystadenoma and borderline serous ovarian cystadenoma tissues. Meanwhile, several small molecules with potential therapeutic efficacy against OC were identified in our study.

Conclusions: By means of bioinformatics analysis, we identified six real hub genes and indicated a group of candidate small molecule drugs as adjunctive agents for OC. They could be the potential novel biomarkers for the diagnosis and promising therapeutic targets of OC.

Keywords: Ovarian neoplasms; bioinformatics strategy; novel candidate genes; small molecule drugs; nonstructural maintenance of chromosomes condensin I complex subunit G (*NCAPG*)

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Introduction

Ovarian cancer (OC) is the second most common gynecologic cancer and most deadly gynecologic malignancy around the world, with 1.6% incidence and 2.1% mortality in 2020 and a 5-year survival rate as 47.6% (1,2). As of 2021, OC is still the fifth cause of cancer related death in females in the United States (3). There are approximately 225,500 new cases and an estimated 140,200 ovarian cancerrelated deaths each year (4). The high annual mortality was attributed to late diagnosis and recurrence associated with chemotherapy-resistance. Most patients diagnosed with metastatic disease will have disease recurrence within 5 years, and the 5-year survival rate is reduced to <20% (5). At present, the most common treatment is cytoreductive surgery combined with platinum-based chemotherapy. However, 60% of patients relapsed following first-line treatment and 50% were resistant to chemotherapy. Therefore, it is crucial to discover valid biomarkers for early diagnosis and find drugs that affect the mechanism of drug resistance.

In recent years, a significant number of genes and targets related to the disease were found because of the Human Genome Project and other genomic studies. With the rapid development of gene microarray technologies and high throughput sequencing (6), more and more ovarian cancer related differentially expressed genes (DEGs) and functional pathways having exactly been identified, leading to breakthroughs in the ovarian cancer research. Thanks to the advent of bioinformatics, direct drug development for biomolecules against disease-related targets has gradually become the primary strategy for drug development (7).

In this research, GSE18520 and GSE14407 from the Gene Expression Omnibus (GEO) database were selected and the overlapped DEGs were detected. The Gene Ontology (GO) analysis and Kyoto Encyclopedia of Genes and Genome (KEGG) pathway enrichment analysis were performed to establish the PPI network of DEGs and identified the hub genes. Furthermore, we validated the expression of the identified hub genes by using Gene Expression Profiling Interactive Analysis (GEPIA), Oncomine database and Human Protein Atlas (HPA) databases. The prognostic value of them were evaluated by the Kaplan Meier plotter online tool. NCAPG is one of the identified hub gene. The expression of NCAPG was further explored by immunohistochemistry in our OC tissues. Moreover, we used the CMap database to look for prospective small compounds with therapeutic efficacy based on OC RNA-seq, indicating the way for future drug development. We present the following article in accordance with the REMARK reporting checklist (available at https://tcr.amegroups.com/article/view/10.21037/tcr-21-2890/rc).

Methods

Data resources

Two original microarray datasets were downloaded from the GEO database (http://www.ncbi.nlm.nih.gov/geo/). GSE18520, which was based on the GPL570 (HG-U133_ Plus_2; Affymetrix Human Genome U133 Plus 2.0 Array), contained 53 high-grade serous papillary carcinoma samples and 10 normal ovarian surface epithelium samples. The platform for GSE14407 is also GPL570, contained 12 serous papillary ovarian cancer samples and 12 normal ovarian epithelium samples.

Data preprocessing and DEGs screening

The GEO2R online tools were utilized to identify the DEGs between OC samples and normal tissue samples with the cutoff threshold of adjust P<0.05 and llog2(fold change) | >2. The online Venn 2.1.0 software (https://bioinfogp.cnb.csic.es/tools/venny/) was applied to get the overlap genes between the two datasets.

Functional annotation and pathway analysis

GO enrichment analysis is a commonly used approach to investigate the biological process (BP), molecular function (MF) and cell component (CC) of genes or gene products. KEGG is a widely used database for identifying functional and metabolic pathways. The DAVID (Annotation, Visualization and Integrated Discovery) database (version 6.8; http://david.abcc.ncifcrf.gov/) was used to carry out GO function and KEGG pathway enrichment analysis. P<0.05 was set as the cut-off criterion for the significant enrichment.

Protein-protein interaction (PPI) network construction and module analysis

The online database Search Tool for the Retrieval of Interacting Genes (STRING, version 11.0, https:// string-db.org/) was used to illustrate the potential interrelationships among the overlapping DEGs. Then the Cytoscape software (https://cytoscape.org) was utilized

| Characteristics | Malignant (n=69) | Borderline (n=15) | Benign (n=35) |
|-------------------------|---------------------|----------------------|------------------|
| Age, years, n (%) | | | |
| ≤60 | 48 (69.6) | 12 (80.0) | 27 (77.1) |
| >60 | 21 (30.4) | 3 (20.0) | 8 (22.9) |
| FIGO stage, n (%) | | | |
| I | 8 (11.6) | | |
| II | 10 (14.5) | | |
| III | 45 (65.2) | | |
| IV | 6 (8.7) | | |
| Histologic grade, n (%) | | | |
| Low grade | 31 (44.9) | | |
| High grade | 38 (55.1) | | |
| CA125, n (%) | | | |
| ≤500 | 27 (39.1) | | |
| >500 | 42 (60.9) | | |
| HE4, n (%) | | | |
| ≤500 | 39 (56.5) | | |
| >500 | 30 (43.5) | | |
| Ascites | | | |
| Yes | 45 (65.2) | | |
| No | 24 (34.8) | | |
| Lymph node metastasis | | | |
| Yes | 37 (53.6) | | |
| No | 29 (42.0) | | |
| Ki 67, n (%) | | | |
| ≤50% | 19 (27.5) | | |
| >50% | 50 (72.5) | | |

FIGO, International Federation of Gynecology and Obstetrics.

to construct the network and investigate the interaction relationship of the DEGs encoding proteins in OC. A combined score above 0.4 was set as the cut-off value. Finally, the MCODE (Molecular Complex Detection) plugin of Cytoscape software was used to visualize and identify significant modules from the PPI network for the selection of hub genes (degree =5, node score =0.2, k-core =2, and maximum depth =100).

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Validation of key genes

The intersecting genes from the enrichment pathways and top 10 nodes in PPI network were identified as hub genes. "Expression analysis-Box Plots" module of The Gene Expression Profiling Interactive Analysis (GEPIA, http:// gepia.cancer-pku.cn) was adopted for analyzing hub genes mRNA levels in OC, considering that the default setting cut-off value are the |Log₂FC| Cutoff:1, P value Cutoff: 0.01, and "Match TCGA normal and GTEx data". The Oncomine database (https://www.oncomine.org/) were also used to analyze their expression in OC compared with levels in normal specimens. The threshold value was set as follows: fold change: 1, P value: 0.05, and gene ranking: 10%.

The immunohistochemical results of the real hub genes were obtained from the Human Protein Atlas (version 18, https://www.proteinatlas.org/). The Kaplan-Meier-plotter (http://kmplot.com/analysis) online database was used to evaluate survival analysis of these 10 hub genes. A total of 1,656 OC patients for overall survival (OS) analysis. The genes and Affy ID are as follows: *CCNB1* (214710_s_at), *TOP2A* (201291_s_at), *NUSAP1* (218039_at), *NCAPG* (218662_s_at), *KIF20A* (218755_at), *DLGAP5* (203764_at), *CDK1* (203468_at), *FOXM1* (202580_x_at), *RRM2* (202580_x_at), *AURKB* (209464_at).

Median expression of hub genes of all samples was chosen as a cut-off to divide samples into the high expression group and low expression group. The Kaplan-Meier analysis was performed to investigate the prognosis of patients in the two groups.

Identification of candidate small molecules

The Connectivity Map (CMap) database (http://www. broadinstitute.org/cmap/), which is based on the gene signature of OC was used to find potential drugs for patients. The common dysregulated probe sets were used to query the CMap database.

Using the perl language, convert the up- regulated genes and down-regulated genes in *Table 1* to GPL571 probe IDs and enter them into the CMap official online "rapid query" tool. Tag files can be found in the Supplementary Material. As a result, the enrichment scores that represent similarity were calculated (ranging from -1 to 1). A positive connectivity score (closer to 1) shows that a drug is capable to induce the input signature of OC cells, whereas a negative connectivity score (closer to -1) shows that a drug is capable to reverse the input signature. The

negative connectivity scores were confirmed as candidate molecules with potential therapeutic value. Tomographs of these candidate small molecular drugs were inquired in the Pubchem database (https://pubchem.ncbi.nlm.nih.gov/).

Human tissue samples

Totally 69 serous ovarian cancer, 35 benign serous ovarian cystadenoma tissues and 15 borderline serous ovarian cystadenoma tissues were obtained, all of them underwent first surgery at the Department of Obstetrics and Gynecology of the First Hospital of Lanzhou University from January 2015 to December 2020. None of them had received any chemoradiation, radiation, or other treatments before the surgery. The diagnoses of the pathological sections were confirmed by pathology department consultation (*Table 1*). This study was approved by the Ethics Committee of the First Hospital of Lanzhou University. All the patients were informed and signed informed consent.

Immunohistochemical analysis

Each tissue was embedded in paraffin blocks and continues processed into 4-µm thick paraffin sections. Specimens were deparaffinized and hydrated, 3% H₂O₂ were used for 15 min to eliminate endogenous peroxidase and then citrate buffer solution (pH=6) were used to repair antigens. The sections were then blocked with 10% serum for 60 min and incubated with rabbit anti-NCAPG antibody (1:100; Abcam, ab251864, Cambridge, UK) at 4 °C overnight followed by incubation with secondary antibody goat antirabbit IgG (Zhongshan Golden bridge Biotechnology, SAP-9100, Beijing, China) for 30 mins at 37 °C. Then diaminobenzidine and hematoxylin were employed to stain these sections, and they were sealed with neutral resins. The results were evaluated by two pathologists blindly and randomly. The positive staining of NCAPG was located in the cytoplasm and membrane. The intensity of NCAPG expression was scored as follows: 0, negative staining; 1, mild staining; 2, moderate staining; 3, strong staining. The percentage of NCAPG expression was scored as follows: 0, <10% positive cancer cells; 1, 11–25% positive cancer cells; 2, 26-50% positive cancer cells; 3, 51-75% positive cancer cells; 4, >75% positive cancer cells. The final scores of each staining were calculated by multiplying the two scores. The section with a score of 0-4 was defined as a low expression,

whereas the section with a score of 5-12 was defined as high expression.

Statistical analysis

Statistical analysis was performed using SPSS software (version 26.0). In our tissue samples, Chi-square test was used to analyze the difference between three groups. The P<0.05 was considered to indicate a statistically significant difference.

Ethical statement

The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by the Ethics Committee of the First Hospital of Lanzhou University (No. LDYYLL2019-275) and individual consent for this retrospective analysis was waived.

Results

Identification of DEGs in OC

A total of 433 overlapping genes (*Table 2*) were extracted to be differentially expressed. 159 up-regulated genes (*Figure 1A*) and 274 down-regulated genes (*Figure 1B*) as shown respectively in the Venn diagrams.

Enrichment analyses

The bubble map was revealed via the ggplot2.R package for the top 10 significant GO and pathway based on P value. For BP of GO enrichment, GO analysis results showed that DEGs were markedly enriched in negative regulation of transcription (*Figure 2A*). As for CC of GO enrichment, DEGs were particularly involved in extracellular region (*Figure 2B*). In addition to MF, the overlapping DEGs were particularly enriched in calcium ion binding (*Figure 2C*). Furthermore, KEGG pathway enrichment analysis demonstrated that these DEGs were mainly enriched in pathways in cancer and proteoglycans in cancer (*Figure 2D*).

PPI network construction and module analysis

There were 344 nodes and 1,596 edges found in the network (*Figure 3A*). The module genes CDK1, FOXM1, CCNB1, RRM2, AURKB, TOP2A, NUSAP1, NCAPG, KIF20A and DLGAP5 were the most significant 10 node degree genes

Table 2 Screening DEGs in ovarian cancer by integrated microarray

| DEGs | Genes name |
|------------------------|---|
| Upregulated (159) | HMGA1 LZTS3 MECOM SUSD2 CLUH CD24 KLHL14 DUXAP10 SOX17 MCM10 LOC101060391 C1orf186 CLDN3 GRHL2 ELF3 LYPD6B WFDC2 NEK2 DLX6-AS1 CP E2F1 MPZL2 FOXM1 NES SMIM22 ESM1 ARHGAP11B SLC52A2 KLF12 KIF14 CLDN4 C1orf106 TIMELESS CYYR1 FOLR1 SUSD4 PSAT1 DLGAP5 EHF NRXN1 PAX8 KIF20A SCGB2A1 FILIP1 BUB1 GPM6B LPAR3 FAM83D TTK AIF1L TRIP13 CENPF CEP55 NCAPG SLC4A11 RNF157-AS1 DTL IGF2BP3 CATIP-AS1 CDCA5 KIF4A S100A1 SLC2A1 EPCAM KIF11 EPHX4 STON2 BUB1B MELK UBE2C LIMS3 CRABP2 MUC1 SULT1C2 CENPA SOX9 PROM2 CBS SORT1 ESRP1 MXD3 CKS2 CDK1 PRC1 FZD10 CDC20 CXXC5 TOP2A CENPK CDH6 MMP7 LOC101928554 FOXQ1 ECT2 LINC00511 ESCO2 C8orf4 CCNB2 NUSAP1 S100A2 KLK6 LYNX1 PRR11 KIAA0101 C12orf56 LYPD1 EPB41L5 RGS1 PRSS2 LRP8 CENPU NR2F6 SLC26A7 HMMR CCNB1 RAD51AP1 S100A13 RRM2 PRAME ST6GALNAC1 MTHFD2 MTFR2 MAL TTC39A INHBB KLK8 AURKB KIAA1217 LOC150051 DCDC2 DIS3L2 HEY2 GLDC TMTC1 LCN2 DUXAP10 DEPDC1 RAPGEF3 DIRAS2 MAGEA11 HIST1H1C PTX3 KCCAT333 PCDH7 NRTN TFAP2A SCGB1D2 HMGA2 MEOX1 FAM107A DEFB1 PTH2R LOC613266 COL12A1 WDR72 LIX1 SST PDCL2 |
| Downregulated (274) | LINC01105 BNC1 ITLN1 LHX9 MAF PDE8B GADL1 PEX5L SVEP1 ABCA8 REEP1 NKX3-1 METTL7A HAND2-AS1 HBB SNX29P2 SNCAIP OGN GFPT2 SYT4 SNX13 ANTXR2 SLC4A4 MNDA AADACL2 DIRAS3 CHGB WNT2B CLDN15 HBA1 NPY1R AOX1 GPRASP1 BCHE PRG4 PPM1K TBX3 LGALS2 CLEC4M DOCK5 HELQ TCEAL2 AGTR1 SMPD3 TCEAL7 S100A10 SFRP1 PCOLCE2 SNCA PKD2 ADH1B SHISA3 DDR2 PTPRZ1 DFNA5 B3GALT2 MTUS1 LGALS8 ADH1C PRSS35 EIF1 ALDH1A2 DSC3 AKAP12 TCF21 PCDH9 PROCR NBR1 CMAHP CALB2 CPB1 CSGALNACT1 ARHGAP18 GATA4 FLRT2 SEMA5A THBD OGFOD1 DST BTD FAM153B MGARP COL8A1 LHX2 WNT16 LSAMP HBG1 ABHD12B ANKRD29 CHRDL1 SORBS2 HLF DAB2 ADGRD1 PTGDR OMD GAS1RR RYR2 SERTM1 KLF2 RTN1 MGC24103 LOC100996760 MCTP2 KCNT2 TMEM37 PDGFD SPOCK1 PLEKHH2 VGLL3 TMEM255A ILDR2 ARX ANXA8 LINC01133 ANXA1 NELL2 DMD PTGIS NR2F1-AS1 HHIP LOC286191 CXorf57 SLC30A4 PAPSS2 FGF1 PLCE1 BDH2 TFPI BCAR3 RBMS3 NEFH PEG3 PMP22 IL18 FAM153B C7 RERG MEOX2 HSD17B2 MARCO PRDM5 RAB27B GHR IRAK3 AKT3 FGF13 ABI3BP CALCRL MEDAG CPED1 NT5E HPSE EZR OLFML1 SLC41A2 ALDH1A1 NEGR1 PTGER3 BAMBI PCDH17 CFI GIPC2 DOCK11 CELF2 MUM1L1 SBSPON NLGN4X CFC1 DPP10 RNASE4 AQP9 CYP39A1 LY75 DTNA RGS4 SCD5 EFEMP1 MMP28 LIMA1 GPM6A SDPR PRRX1 NAP1L2 CLMP GABRB2 RUNX1T1 RNF128 NXPH2 AKR1C2 CYP2U1 CNTN1 PTHLH NAP1L3 SIGLEC11 FABP4 PITPNC1 CAV1 DSE CNTN4 KLF4 PDPN FLRT3 C10rf168 FAM134B ARHGAP44 CBLN4 GATM HSD17B6 ECM2 PGR PKHD1L1 CFH SFRP2 HPGD ADAMTS3 TMEM150C DPYD MGP MCOLN3 LAMA4 CYS1 NKAIN2 COL14A1 S100A8 TRPC1 FAM13C MCC RARRES1 TBX18 COL3A1 BNC2 LRP2 DCN LINC01279 NBEA SLITRK5 ITM2A FRY GNG11 FGF9 FAM155A CCDC80 LINC01116 ZFPM2 GAS1 FRAS1 TMEM98 WDR17 TSPAN8 RASSF3 SLIT2 PRKAR2B NDN TFPI2 DPP6 MDFIC TLE4 PROS1 PHLDB2 CFH NRXN3 MSRB3 BEX1 MEIS2 ARRDC4 GATA6 STK26 MAOA CRNDE WNT5A RSP01 MICU3 PCDH20 |

DEGs, differentially expressed genes.



Figure 1 Venn diagram of overlapping 433 DEGs in the two datasets. Different color represents different datasets. (A) 159 Up-regulated DEGs (logFC >0). (B) 274 Down-regulated DEGs (logFC <0). DEGs, differentially expressed genes.



Figure 2 Functional and signaling pathway analysis of the overlapped DEGs in OC. (A) Biological processes. (B) Cellular components. (C) Molecular function. (D) KEGG pathway. DEGs, differentially expressed genes; OC, ovarian cancer; KEGG, Kyoto Encyclopedia of Genes and Genome.

and identified as the hub genes (*Figure 3B*). Additionally, the most important PPI network module was detected using MCODE (*Figure 3C*). Pathway enrichment analysis showed that the most top module genes were primarily enriched in cell cycle, p53 signaling pathway, progesterone-mediated oocyte maturation and oocyte meiosis (*Table 3*).

Validation of bub genes

GEPIA and Oncomine tool verified that the expressions of CDK1, FOXM1, CCNB1, RRM2, AURKB, TOP2A, NUSAP1, NCAPG, KIF20A and DLGAP5 were significant elevated in OC compared with normal ovarian tissues (Figure 4A,4B). It was found that higher expressions of CCNB1, TOP2A, NUSAP1, NCAPG, KIF20A and DLGAP5 were significantly correlated with worse overall survival for OC patients (Figure 5). Via the Human Protein Atlas database, these six genes' expression were higher in OC compared than in normal tissues (Figure 6).

Validation of NCAPG expression in patient tissues

As shown in *Table 4* and *Figure 7*, NCAPG was mainly located in the cell cytoplasm and membrane. The scoring

system results demonstrated that NCAPG high expression rate was 58/69 (84.1%) in serous ovarian cancer (SOC) samples, which was significantly higher than that in benign serous ovarian cystadenoma (5/35, 14.3%) and borderline serous ovarian cystadenoma tissues (2/15, 13.3%) (P<0.05).

Identification of related small molecule drugs

According to the P value, the top 10 related small molecules which matched to the OC gene expression changes are listed in (*Table 5*). Six of these small molecules indicated a negative correlation and showed that latent capacity of tumor inhibition in clinical applications, four of them indicated a positively correlation. Among them, trichostatin A, vorinostat, phenoxybenzamine, 8-azaguanine showed a significantly negative correlation (P<0.05) and the tomography of them shown in (*Figure 8*).

Discussion

Ovarian cancer remains the most lethal gynecologic cancer due to delayed diagnosis, which could explain the dismal prognosis of OC patients. Early diagnosis of OC will improve the overall survival. Therefore, to discover

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Figure 3 The PPI network analysis of DEGs and module analysis. (A) The PPI network consisted of 344 nodes and 1596 edges were constructed by overlapping DEGs. Bigger nodes and dark colors represent genes with higher degree values. (B) Top 10 hub genes in the PPI network. (C) The most top module of PPI network identified in MOCDE. DEGs, differentially expressed genes; PPI, protein-protein interaction; MCODE, Molecular Complex Detection.

| There is a manual analysis of module genes | | | | |
|--|---|-------------|----------|--|
| Pathway ID | Pathway description | Gene counts | P value | |
| 4110 | Cell cycle | 7 | 1.39E-08 | |
| 4115 | p53 signaling pathway | 4 | 1.33E-04 | |
| 4914 | Progesterone-mediated oocyte maturation | 4 | 3.06E-04 | |
| 4114 | Oocyte meiosis | 4 | 5.99E-04 | |
| | | | | |

 Table 3 KEGG pathway analysis of module genes

KEGG, Kyoto Encyclopedia of Genes and Genome.

reliable biomarkers and to reveal the possible molecular mechanisms of OC is especially important for diagnosis and prognosis. In the present study, we utilized bioinformatics methods to identify novel biomarkers and potential small molecule drugs in OC by integrating two expression profile datasets GSE18520 and GSE14407. 433 common DEGs were identified, among which 159 genes were upregulated and 274 genes were downregulated. Applied GO function and KEGG pathway enrichment analysis, we found that the biological processes involved in DEGs were mainly acted in negative regulation of transcription and DNA-templated. The DEGs were mainly enriched in Pathways in cancer and Proteoglycans in cancer. These DEGs might involve in the carcinogenesis of OC by means of affect the biological behavior of OC cells via these pathways.

Based on the identified DEGs, we constructed a PPI network with 344 nodes and 1596 interactions. We extracted the most important module with the most significant degree from the PPI network. The top ten module genes CDK1, FOXM1, CCNB1, RRM2, AURKB, TOP2A, NUSAP1, NCAPG, KIF20A, and DLGAP5 were chosen as hub genes based on their degree rankings. Survival analysis for 1,656 OC patients from Kaplan Meier plotter database showed that patients with higher expression of CCNB1, TOP2A, NUSAP1, NCAPG, KIF20A and DLGAP5 experienced worse survival outcomes than those with low expression. These results provide signs for us to create an ovarian cancer prognosis model, allow us to more correctly identify high-risk patients. The GEPIA, Oncomine and the Human Protein Atlas database showed that they were all overexpressed in OC tissue.

NCAPG (non-structural maintenance of chromosomes condensin I complex subunit G), a crucial component of the chromosomes condensin complex, which plays an important role in the condensation and stabilization of chromosomes during meiosis and mitosis (8). In hepatocellular carcinoma (HCC), Wang *et al.* found that elevated NCAPG

expression was remarkably associated with worse diseasefree survival and overall survival in patients suffering from HCC. NCAPG could be regard as a novel prognostic biomarker to predict HCC early recurrence after surgical resection (9). Arai et al. reported that high NCAPG expression was closely related to prognosis and clinical stage in patients with prostate cancer. Knockdown of NCAPG significantly inhibited the ability of proliferation, migrationand invasion in prostate cancer cells (10). In our studies, we verified the expression of NCAPG by the immunohistochemistry analysis in ovarian cancer tissues. The results showed that the expression rate of NCAPG in OC tissues was significantly higher than that in benign serous ovarian cystadenoma and borderline serous ovarian cystadenoma tissues that obtained from clinical patients, and the tendency was consistent with the results discussed above. Taken all these studies together, we assumed that NCAPG may be a valuable biomarker for the diagnosis and prognosis of various cancer patients.

CCNB1 (*Cyclin B1*), one member of the cyclin family, an important regulator in normal cell cycle progresses at the G2/M transitions by binding to CDK1 (11), plays a key role in various human malignancies. Han *et al.* found that knockdown long non-coding RNA SOX2OT could inhibit ovarian cancer cell migration and cell invasion by downregulated CCNB1 (12).

TOP2A (topoisomerase II alpha), is a key enzyme in DNA transcription and replication, repair, chromosome condensation and separation (13), which has been identified overexpression in several cancer types. Researchers found that TOP2A is the prime therapeutic target of many anticancer therapy regimens. It was proved that inhibitors targeting human topoisomerase II alpha were useful chemotherapy option for the treatment of many patients suffering from all kinds of cancers (14).

NUSAP1 (nucleolar and spindle associated protein 1) played a role in mitotic progression, spindle formation, and



Figure 4 Validation the expression of hub genes on transcription level by (A) GEPIA database and (B) Oncomine database. GEPIA, Gene Expression Profiling Interactive Analysis.



Figure 5 The prognostic value of the 6 hub genes in ovarian cancer patients. Prognostic value of CCNB1. (B) Prognostic value of TOP2A. (C) Prognostic value of NUSAP1. (D) Prognostic value of NCAPG. (E) Prognostic value of KIF20A. (F) Prognostic value of DLGAP5. HR, hazard ratio.



Figure 6 Validation the expression of 6 hub genes on translational level by The Human Protein Atlas database (immunohistochemistry).

| Table T Expression of TOPAL O in uniferent ovarian ussues | | | | | | |
|---|---------|-----------------------|----------------------|----------|---------|--|
| Group | Casas | NCAPG expression | | | | |
| | Cases — | High expression cases | Low expression cases | χ^2 | P value | |
| Malignant | 69 | 58 (84.1%) | 11 (15.9%) | 57.412 | 0.000 | |
| Benign | 35 | 5 (14.3%) | 30 (85.7%) | | | |
| Borderline | 15 | 2 (13.3%) | 13 (86.7%) | | | |

Table 4 Expression of NCAPG in different ovarian tissues

NCAPG, non-structural maintenance of chromosomes condensin I complex subunit G.



Figure 7 The immunohistochemical analysis of NCAPG in serous ovarian cancer. (A) A benign ovarian tissue showing low NCAPG expression. (B) A borderline ovarian tissue showing low NCAPG expression. (C) A serous ovarian cancer showing high NCAPG expression. NCAPG, non-structural maintenance of chromosomes condensin I complex subunit G.

stability overexpression of NUSAP1 has been found in a variety of human malignancies, and it is commonly linked to tumor invasiveness. Zhang *et al.* reported that the expression of NUSAP1 in OC tissues and cells was upregulated, which could promote the proliferation and invasiveness of OC cells (15).

KIF20A (Kinesin family member 20A), as a mitotic kinesin-like protein, belongs to the kinesin perfamily-6. Depending on microtubule-dependent plus-end motility, KIF20A is contributed to different cellular processes including formation of the mitotic spindle and chromosome partitioning. In the field of oncology, increasing studies illustrated its carcinogenic properties in various types of cancers. DLGAP5 (Discs large-associated protein 5), also known as HURP, is a mitotic spindle protein that plays a crucial role in the formation of novel tubulin sheet, the formation of facilitating spindle, and forming a connection between the centrosome and kinetochore (16). A large number of studies has shown DLGAP5 serve as oncogenic role in many types of human cancers. Chen *et al.* reported that DLGAP5 silencing could suppress tumorigenicity and cellular proliferation by arresting the cell cycle at the G2/M phase in ovarian cancer cells (17). All results above suggested that these 6 hub genes may relate to the occurrence and development of OC.

In this study, we also identified several small molecules with potential therapeutic efficacy against OC. Trichostatin

Table 5 List of the top ten OC-related small molecules with highly significant correlations in results of CMap analysis

| Rank | CMap name | Mean | Ν | Enrichment | P value |
|------|-----------------------|--------|-----|------------|---------|
| 1 | Trichostatin A | -0.469 | 182 | -0.428 | 0.00000 |
| 2 | Vorinostat | -0.598 | 12 | -0.621 | 0.00000 |
| 3 | Phenoxybenzamine | -0.835 | 4 | -0.937 | 0.00000 |
| 4 | 8-azaguanine | -0.768 | 4 | -0.917 | 0.00010 |
| 5 | Resveratrol | -0.694 | 9 | -0.669 | 0.00016 |
| 6 | Podophyllotoxin | 0.672 | 4 | 0.849 | 0.00074 |
| 7 | Chenodeoxycholic acid | 0.595 | 4 | 0.846 | 0.00080 |
| 8 | Thioperamide | 0.563 | 5 | 0.798 | 0.00082 |
| 9 | Vinburnine | 0.576 | 4 | 0.83 | 0.00127 |
| 10 | Luteolin | -0.734 | 4 | -0.835 | 0.00131 |

Cmap, Connectivity Map; OC, ovarian cancer.



Figure 8 The tomographs of the four candidate small molecule drugs for OC. OC, ovarian cancer.

A (TSA), as a potent inhibitor of histone deacetylase (HDAC), was considered to be the most significant small molecule with strong dose-dependent antitumor activity. Drug resistance is the main cause of the high mortality in patients with OC. Combination therapy is a possible method of overcoming drug resistance. More and more studies confirmed that TSA could enhance the antitumor effects when combinated with other anticancer drugs in multifarious cancer cells (18-20). Lambert (21) found that TSA could reduce cell viability instigates apoptosis, autophagy, inhibition of cell cycle progression in human ovarian cancer cells. Zhang (22) confirmed that TSA caused significant cytotoxicity and induced apoptosis in SKOV3 cells. Vorinostat is a small molecule inhibitor of both class

I and class II HDAC enzymes. It can directly activate $p21^{WAF1/CIP1}$ gene transcription, which is one of the most important cyclin-dependent kinase (CDK) inhibitors in ovarian cancer cells (23). Vorinostat has been reported to produce specific modifications in the pattern of acetylation and methylation of lysines in H3 and H4 histones that are associated with the $p21^{WAF1/CIP1}$ gene promoter (24). It played an important role in antiproliferative and proapoptotic effects by inducing the expression of IL-8/CXCL8 in OC cells (25). In ovarian cancer cell lines, the combination of vorinostat and paclitaxel was superior to either drug individually (26). Phenoxybenzamine, is approved for the treatment of hypertensive emergencies associated with the adrenal medulla tumors and pheochromocytomas by the US

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Food and Drug Administration. At present, more and more studies have reported that phenoxybenzamine can inhibit the growth, invasion, and migration of human tumor cells as a histone deacetylase inhibitor. Inchiosa (27) reported that Phenoxybenzamine could inhibitory the activity of OVCAR-8 cells. 8-Azaguanin (8-AG), originally developed as an antineoplastic agent, but recent study has found it to be a novel immunomodulatory drug (28). 8-AG as an immunomodulatory drug, increased the functionality of NK cells via NK-target cell conjugate formation and cytolytic granule polarization. But it has not yet been demonstrated to have effectiveness against OC, and therefore further research is required.

In summary, by means of bioinformatics analysis, we identified six real hub genes of OC, including *CCNB1*, *TOP2A*, *NUSAP1*, *NCAPG*, *KIF20A* and *DLGAP5*. Survival analysis showed these genes were significantly associated with negative survival of patients with OC. They might play crucial roles and as novel biomarkers in OC diagnosis, prognosis and treatment. In this study, we also indicated several important signaling pathways associated with the OC onset and progression, and identified a group of candidate small molecule drugs as adjunctive agents for OC. However, further studies are needed to determine the relationship between OC and these pathways and these small molecule drugs.

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Footnote

Reporting Checklist: The authors have completed the REMARK reporting checklist. Available at https://tcr. amegroups.com/article/view/10.21037/tcr-21-2890/rc

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Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at https://tcr.amegroups.com/article/view/10.21037/tcr-21-2890/coif). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by the Ethics Committee of the First Hospital of Lanzhou University (No. LDYYLL2019-275) and individual consent for this retrospective analysis was waived.

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