

# Irradiation and combination immunotherapy

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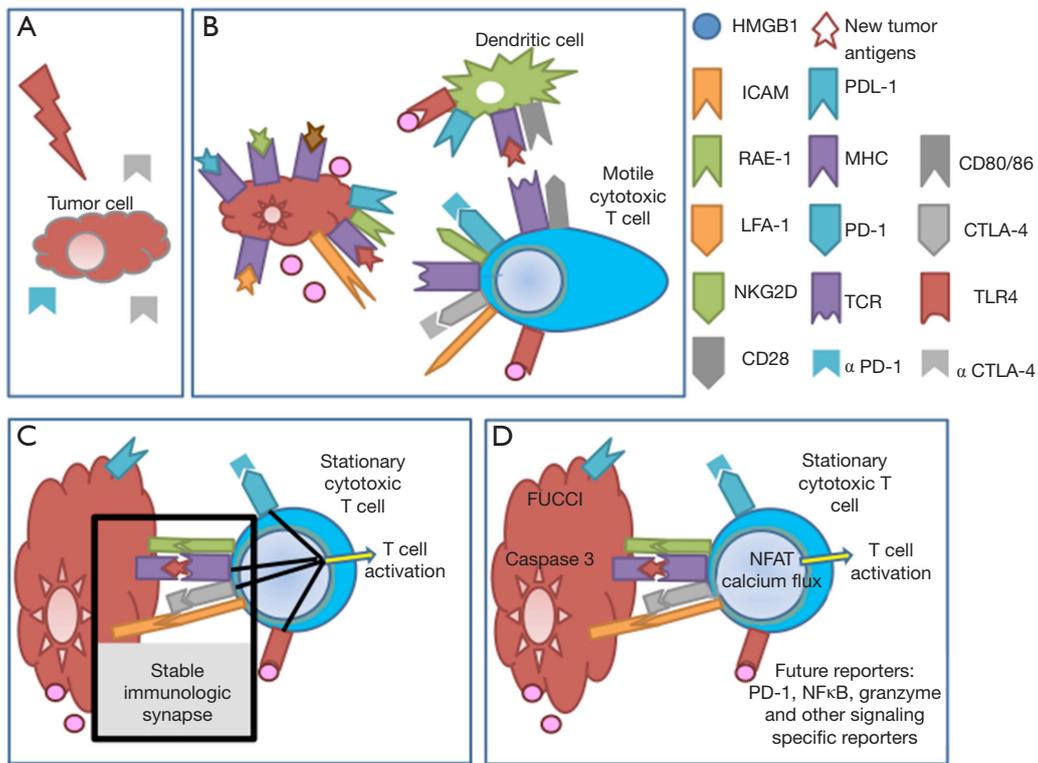
Our ability to control and eradicate cancer has advanced with new combinations of surgery, chemotherapy, irradiation therapy and recently, immunotherapy. The concept of a single cancer therapy has passed with each new discovery revealing the complexity of the genetics and immunology of the tumor microenvironment. Understanding which combination of standard and emerging therapies that can provide long lasting remission of each particular cancer is paramount. Here we review the capacity of radiation therapy (RT) combined with immune checkpoint inhibitors to induce a complete response in mammary carcinoma and melanoma (1,2) and the evaluation of therapy mechanism of action using intravital microscopy. Current tools and approaches to evaluate mechanism of action of therapies have been limited, but real time intravital imaging offers prospects of enhancing our knowledge.

One standard of care cancer therapy of immunological interest is RT. Local RT is effective at killing tumor cells directly, but the effect of RT can extend beyond the treated primary tumor. The abscopal effect of RT is an anti-tumor immune response generated at sites distant (systemic) from the irradiated volume (3). Immunogenic tumor cell death caused by RT represents in effect an *in situ* vaccine specific for that patient (4). It provides neighboring antigen presenting cells with tumor antigens, neoantigens (non-self peptides that are generated by the mutated cancer genome), and activating danger signals, such as HMGB1 which binds to TLR4, and calreticulin which leads to priming and activation of tumor-specific T cells capable of attacking the tumor at primary or distant sites (5). RT

also increases the T cell receptor (TCR) repertoire, which allows the expansion of T cells clones against the tumor with diverse TCR traits (2). Studying this process has been limited to *in vitro* studies and “snap shot” analysis of mouse tumors and perturbed immune systems exposed to RT. The application of imaging technologies to study pathology is one of the most transformative advancements in medicine and as technology advances, it will continue to have broader applications in the future (*Figure 1*).

Previously, it has been shown via single cell microscopy that stable immunological synapse formation between DCs and T cells is required for signalling and that productive interactions can be determined by the prolonged time of contact of the membranes of each cell (6). The outcome of that interaction, whether activating or suppressing, requires the use of specific reporters or other assays. The engagement of CTLA-4 on T cells by CD80/CD86 on DC is a negative regulatory signal for T cells. Immune checkpoint antibodies, like anti-CTLA-4 and anti-PD-1, work by blocking the mechanisms that hinder the activation and function of anti-tumor T cells. Through intravital imaging, anti-CTLA-4 has been shown to increase T cell motility and reduce contact periods between T cells and antigen-presenting cells (7).

Two photon intravital imaging represents a more contemporary way to study the effects of RT and anti-CTLA-4 combination therapy. Ruocco *et al.* studied RT in combination with anti-CTLA-4 (9H10) treatment in a non-immunogenic mouse mammary cancer model (4T1) (1). They showed that standard of care ionizing RT, is



**Figure 1** Radiation in combination with antibody immunotherapy. (A) Local radiation therapy (RT) and anti-CTLA-4 and/or anti-PD-1 antibody therapy leads to an immunogenic response to the tumor driven by DNA damage, release of HMGB1, and generation of tumor antigens and neo-antigens; (B) MHC class I, RAE-1 and ICAM-1 is upregulated on the tumor and new tumor antigens are presented to cytotoxic T cells. Dendritic cells adjacent to the tumor likewise are able to present new tumor antigens to activate the T cells against the tumor. HMGB1 can bind to TLR4 on T cells and dendritic cells to activate the NFκB pathway to initiate activation and proliferation. CTLA-4 and PD-1 targeted antibody therapy primes the T cell to be receptive to activating signals; (C) upon direct T cell contact with the tumor, RAE-1 and ICAM-1 stabilize the immunologic synapse around the T cell receptor (TCR) in contact with MHC class 1 which can lead to T cell signalling even in the absence of co-stimulation. Inhibiting the suppressive signalling of CTLA-4 and PD-1 via antibody therapy, coupled with stable TCR and TLR4 signalling leads to T cell activation and proliferation; (D) imaging reporters useful for interrogating signalling directly in the tumor or in immune cells to determine the effect of therapy within the tumor microenvironment.

able to induce immunogenic tumor antigens and other microenvironment changes required for a robust anti-tumor response. Intravital microscopy was used to determine the efficacy of immunotherapy on tumor growth, describing the direct interaction of CXCR6<sup>+</sup> CD8<sup>+</sup> T cells with the CFP<sup>+</sup> tumor and the behaviour of the CD8<sup>+</sup> T cells within the tumor microenvironment after RT and/or anti-CTLA-4 therapy. RT and anti-CTLA-4 therapy in combination was shown to control the growth of established tumor and this was attributed to the enhanced infiltration of activated CD8<sup>+</sup> T cells (1).

Ruocco *et al.* (1) also showed that anti-CTLA-4 antibody treatment increased T cell motility in the tumor microenvironment, whereas anti-CTLA-4 treatment with

RT promoted T cell arrest in contact with tumor cells. This T cell interaction with tumor cells was an MHC class I-dependent antigen-specific event. Anti-CTLA-4 treatment increased T cell motility on ICAM-1-coated surfaces. After RT, 4T1 cells upregulated expression of MHC class I, ICAM-1, and the NKG2D ligand, RAE-1 $\gamma$ . By using an NKG2D blocking antibody, DX5, with RT and anti-CTLA-4 treatment they showed that the T cell-tumor interactions were decreased and T cell velocity increased suggesting that NKG2D plays a role in stable interactions between CD8<sup>+</sup> effector T cells and tumor cells. Although NKG2D does not play a role in RT-reduced primary tumor growth, the upregulation of RAE-1 does play a role in primary tumor growth in the context of RT and anti-

CTLA-4 therapy, which was also shown to hold true in the experimental metastatic model. Taken together this suggests that tumor antigen recognition by the TCR of CD8<sup>+</sup> effector T cells after RT is stabilized by NKG2D-RAE-1 interactions and activation is enhanced by anti-CTLA-4 treatment resulting in tumor control.

In concert with these combination benefits of RT and immune checkpoint blockade immunotherapy, a subsequent study showed that RT in combination with anti-CTLA-4 had an 18% partial as best response in humans and 17% response in mice with melanoma (2). Additionally, PD-1 is a negative regulatory signal for T cells, where blocking its interaction with PDL-1 on antigen presenting cells or tumor cells has had profound therapeutic effects especially in melanoma patients resulting in an increase in activated T cells (8). Remarkably when RT and anti-CTLA-4 were combined with anti-PD-1 therapy, complete response rates in mice increased to 80% (2). Indeed, the appeal of immune checkpoint blockade therapy is that it induces long lasting anti-tumor responses in patients with advanced-stage cancers.

Intravital microscopy has been used to study the dynamic *in vivo* immune cell responses to infection, autoimmunity and cancer (9,10). Many of the initial tumor intravital studies focused on the development of angiogenesis and the efficacy of anti-angiogenic therapies using intravascular injection of fluorescent dyes (11). Vessel response to RT in a dorsal skin fold chamber showed that there was capillary constriction and thrombus formation from day 4 up to 20 days after treatment (12). Recently this technology has been used to determine the efficacy of a therapy, from chemotherapy penetrance to tumor apoptosis (13,14). There has been an increasing interest in immune cell interactions with tumors and other cells within the tumor microenvironment following the burgeoning field of immunotherapy. Together with development of fluorescent reporter mice to distinguish immune cell subsets and fluorescent reporter tumor cell lines, the migration, invasion and metastasis of tumors have described unexpected interactions with vessels, ECM and the bone marrow niche (9,10,15). The greatest benefit of intravital imaging is the ability to assess the early development of the tumor and interactions between small numbers of transformed cells and immune cells. A consideration of this technique is the depth of penetration where in some cases the first 150  $\mu\text{m}$  of the 400  $\mu\text{m}$  from the outside of the tumor is encapsulation, therefore in a heterogeneous tumor population with potentially a hypoxic or necrotic core, it is important to confirm findings using other methods such as immunofluorescent imaging of tissue sections.

The next important advancement in intravital imaging is real time signalling reporters of immune cell interactions that can be used to predict efficacy of therapies, either by reporting the signalling in immune cells, the metabolic state of cells within the microenvironment, or the apoptosis of tumor cells (14). The NFAT reporter was developed to allow the visualization of activated T cells and can be used to determine the percentage of activated cytotoxic cells within the tumor microenvironment (16). FRET reporters of calcium flux used in neuroscience have also been used to show TCR signalling and recognition of cognate antigen (17). This calcium reporter can be useful in the context of RT and immunotherapy to quantify the number of antigen-specific cytotoxic CD8<sup>+</sup> T cells at the tumor site after therapy. The use of a FRET caspase-3 reporter allows the visualization of apoptotic tumor cells (18), but if multiplexed with additional information about other cells in the tumor microenvironment it could prove to be a powerful tool in dissecting the mechanism of action [reviewed in (14)]. Inhibiting cancer stem cells is a therapeutic approach of interest (15). The Confetti fluorescent construct which randomly assigns different colours to individual cells (19) is useful for lineage tracing of cancer stem cells and has shown that certain clones outcompete adjacent tumor cells. The FUCCI reporter construct allows the visualization of the different stages of cell cycle and is useful in determining whether a therapy is able to stop cancer cell proliferation and the point of cell cycle can be determined (20). Although classical immunological assays allow us to determine the global efficacy of therapies, intravital imaging and the new reporter constructs are unique in resolution of space and time in providing insights into complex interactions within the tumor microenvironment (*Figure 1D*). Ruocco *et al.* effectively used intravital imaging to show that antigen specific recognition of tumor cells after RT is stabilized by NKG2D-RAE-1 interactions resulting in tumor control. The future of intravital imaging is in the development of new functional fluorescent reporters, specific to critical signalling pathways for the direct analysis of therapy mechanisms.

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*Ethical Statement:* The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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