



# Clinical translation of predictive markers for anti-EGFR monoclonal antibody therapy in metastatic colorectal cancer

Andrew Rowland<sup>1,2</sup>, Christos S. Karapetis<sup>2</sup>, Arduino A. Mangoni<sup>1</sup>, Michael D. Wiese<sup>3</sup>, Ganessan Kichenadasse<sup>2</sup>, Ross A. McKinnon<sup>2</sup>, Michael J. Sorich<sup>1,2</sup>

<sup>1</sup>Department of Clinical Pharmacology, <sup>2</sup>Flinders Centre for Innovation in Cancer, Flinders University, Adelaide, Australia; <sup>3</sup>School of Pharmacy and Medical Sciences, University of South Australia, Adelaide, Australia

*Correspondence to:* Andrew Rowland, PhD. Department of Clinical Pharmacology and Flinders Centre for Innovation in Cancer, School of Medicine, Flinders University, Finders Drive, Bedford Park, Adelaide, South Australia, 5042, Australia. Email: andrew.rowland@flinders.edu.au.

*Comment on:* Seligmann JF, Elliott F, Richman SD, *et al.* Combined Epiregulin and Amphiregulin Expression Levels as a Predictive Biomarker for Panitumumab Therapy Benefit or Lack of Benefit in Patients With RAS Wild-Type Advanced Colorectal Cancer. JAMA Oncol 2016. [Epub ahead of print].

Submitted Apr 29, 2016. Accepted for publication May 09, 2016.

doi: 10.21037/tcr.2016.05.19

**View this article at:** <http://dx.doi.org/10.21037/tcr.2016.05.19>

Altered function of the epidermal growth factor receptor (EGFR) and associated mitogen-activated protein (MAP) kinase pathway plays a key role in disease progression in metastatic colorectal cancer (mCRC). Identification of EGFR as a therapeutic target has led to the development of anti-EGFR monoclonal antibodies (mAbs) cetuximab and panitumumab. However, the use of anti-EGFR mAbs in mCRC is associated with considerable treatment-related toxicity and a lack of response in a significant proportion of patients. Given the existence of alternate therapeutic options of comparable efficacy such as the anti-vascular endothelial growth factor receptor mAb bevacizumab (1), there is potential for improved outcomes through the identification of strategies to guide selection of the most appropriate first-line treatment.

Predictive markers can be used to identify patient sub-groups that are most likely to derive benefit from an intervention, facilitating selection of therapies with the greatest likelihood of success for an individual patient. By virtue of their capacity to influence treatment decisions, predictive markers are typically of greater clinical interest compared to prognostic markers (2,3). Assessment of a predictive marker requires the determination of a treatment effect (intervention versus control) in marker-positive and marker-negative cohorts (4). A formal statistical test for interaction between the marker and the treatment group is undertaken. In the oncology setting, the standard approach is to use a Cox proportional hazards model containing

the treatment group, marker, and treatment-by-marker interaction term to model a time-to-outcome endpoint such as progression free survival (PFS) or overall survival (OS). As this analysis involves two or more comparison groups and evaluates the treatment effect of an intervention, data from randomised controlled trials (RCTs) are required. Due to the comparatively limited number of RCTs, particularly for targeted anticancer drugs, this requirement can limit the capacity to definitively validate these markers, compared to prognostic markers, which require only observational studies.

When considering the clinical translation of predictive markers for anti-EGFR mAb therapy in mCRC, sub-group analyses have been performed across a series of RCTs reporting outcomes for both cetuximab and panitumumab (Table 1). On the basis of these analyses, it has been well established that mutation of the downstream RAS oncogenes (collectively present in approximately 50% of mCRC tumors) is associated with a lack of treatment benefit (12). Accordingly, the use of cetuximab and panitumumab is limited to individuals with RAS wild-type (WT) tumors in treatment guidelines (13). While necessary to facilitate response to anti-EGFR mAb therapy, RAS WT status does not in itself ensure benefit, and there remains significant scope to identify additional predictive markers of treatment benefit. Highlighting the importance of consistency in the approach taken to assess predictive markers, due to differences in statistical interpretation, recent meta-analyses

**Table 1** Randomised controlled trials reporting outcomes for anti-EGFR mAbs

| Anti-EGFR agent vs. comparator | Trial (References)   | Background therapy, lines of treatment                              |
|--------------------------------|----------------------|---|
| Cetuximab vs. no cetuximab     | CO.17 (5)            | BSC, $\geq 2^{\text{nd}}$ line                                      |
|                                | CRYSTAL and OPUS (6) | FOLFIRI (CRYSTAL) or FOLFOX-4 (OPUS), 1 <sup>st</sup> line          |
|                                | COIN (7)             | Oxaliplatin and fluoropyrimidine chemotherapy, 1 <sup>st</sup> line |
| Panitumumab vs. no panitumumab | 20020408 (8)         | BSC, $\geq 3^{\text{rd}}$ line                                      |
|                                | 20050181 (9)         | FOLFIRI, 2 <sup>nd</sup> line                                       |
|                                | PICCOLO (10)         | Irinotecan, $\geq 2^{\text{nd}}$ line                               |
|                                | PRIME (11)           | FOLFOX-4, 1 <sup>st</sup> line                                      |

considering the predictive value of BRAF mutation status in this setting have reported conflicting findings (14,15). This led to conjecture regarding the appropriate clinical translation of this marker (16). Similarly, despite pre-clinical and observational evidence for differential effects of individual RAS mutations on response to anti-EGFR mAb therapy, specifically that mCRC patients with KRAS G13D MT tumors may derive a benefit from treatment with anti-EGFR mAbs, a recent meta-analysis demonstrated no significant difference between KRAS G13D and other KRAS MT tumors in terms of benefit from anti-EGFR mAb therapy for mCRC (17).

Epiregulin (EREG) and amphiregulin (AREG) are ligands for EGFR that are overexpressed in mCRC (18), and as such are considered biologically plausible markers of EGFR pathway activity and inhibition (19). Consistent with this mechanistic insight, multiple observational studies (18,20-22) have reported positive correlations between AREG/EREG expression and anti-EGFR mAb efficacy in mCRC, whereby higher ligand expression is associated with improved survival (prognostic effect). More recently, sub-group analyses of two major RCTs (23,24) have reported AREG and EREG expression as a predictive marker of benefit for anti-EGFR mAb therapy in mCRC. It is important to note that assessment of EREG and AREG expression as a predictive marker of treatment benefit from anti-EGFR mAb therapy in mCRC is complicated as expression of these ligands is measured as continuous rather than discrete variables, and thus their analysis requires the determination of a threshold to discriminate marker-positive and marker-negative groups. In order to facilitate translation to clinical practice, both the predictivity of the marker and the robustness of the threshold determination

require validation.

A sub-group analysis of the PICCOLO study reported in JAMA Oncology by Seligmann *et al.* (24) presents a novel dichotomous classification model to synthesize the combined effect of AREG/EREG expression as a predictive marker of treatment effect for panitumumab in mCRC. Using this model, the authors demonstrate that 'high' expression of AREG, EREG or both is predictive of benefit for panitumumab; hazard ratio for PFS in RAS WT patients of 0.38 (95% CI, 0.24 to 0.61) for 'high' expressors, compared to 0.93 (95% CI, 0.64 to 1.37) for 'low' expressors (test of interaction,  $P < 0.001$ ). In this analysis a 'pragmatically chosen' threshold was selected to define 'high' and 'low' ligand expression in order to give 'high' expressor ( $n=140/99$  RAS WT) and 'low' expressor ( $n=183/120$  RAS WT) groups of comparable size. While justified on the basis of maximising power within this analysis, the general applicability of this threshold to a broader patient cohort is unclear. Additionally, while partially inferable data are presented within the manuscript, no absolute criteria defining expression thresholds for either ligand are specified. Notably, in a secondary analysis considering AREG and EREG as independent, continuous markers, AREG (test for interaction,  $P=0.008$ ) but not EREG (test for interaction,  $P=0.08$ ) was demonstrated to predict PFS benefit from panitumumab. Trends toward superior OS were reported for 'high' expressors but were not statistically significant, although this may be anticipated given that the results of the PICCOLO trial were similarly negative for the primary outcome of OS (10). In a prior sub-group analysis of the CO.17 study (23), a threshold defining 'high' EREG expression based on a normalised delta cycle threshold ( $\Delta Ct$ ) for EREG expression relative

to GAPDH expression (6.27) was reported to discriminate benefit from cetuximab in mCRC. Given the importance of reproducibility between cohorts and analyses required to facilitate clinical translation for predictive markers (25), the lack of absolute description of the criteria for assessment of AREG and EREG expression threshold presented by Seligmann *et al.* represents an important barrier to the clinical translation of AREG/EREG expression as a predictive marker of anti-EGFR mAb therapy in mCRC.

Given the emergence of multiple reports supporting the use of EREG/AREG as markers, both prognostic and predictive, for anti-EGFR mAb therapy in mCRC and the clinical imperative of selecting the most appropriate first-line intervention for these patients, clarification of robust and reproducible thresholds to facilitate clinical interpretation of an individual patient's level of expression independent of a study cohort is urgently required.

## Acknowledgments

*Funding:* None.

## Footnote

*Provenance and Peer Review:* This article was commissioned and reviewed by the Section Editor Zhen-Yu Lin, MD (Cancer centre, Union hospital, Huazhong University of Science and Technology, Wuhan, China).

*Conflicts of Interest:* All authors have completed the ICMJE uniform disclosure form (available at <http://dx.doi.org/10.21037/tcr.2016.05.19>). The authors have no conflicts of interest to declare.

*Ethical Statement:* The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

*Open Access Statement:* This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: <https://creativecommons.org/licenses/by-nc-nd/4.0/>.

## References

1. Heinemann V, von Weikersthal LF, Decker T, et al. FOLFIRI plus cetuximab versus FOLFIRI plus bevacizumab as first-line treatment for patients with metastatic colorectal cancer (FIRE-3): a randomised, open-label, phase 3 trial. *Lancet Oncol* 2014;15:1065-75.
2. Italiano A. Prognostic or predictive? It's time to get back to definitions! *J Clin Oncol* 2011;29:4718; author reply 4718-9.
3. Ballman KV. Biomarker: Predictive or Prognostic? *J Clin Oncol* 2015;33:3968-71.
4. Sorich MJ, Coory M. Interpreting the clinical utility of a pharmacogenomic marker based on observational association studies. *Pharmacogenomics J* 2014;14:1-5.
5. Karapetis CS, Jonker D, Daneshmand M, et al. PIK3CA, BRAF, and PTEN status and benefit from cetuximab in the treatment of advanced colorectal cancer--results from NCIC CTG/AGITG CO.17. *Clin Cancer Res* 2014;20:744-53.
6. Bokemeyer C, Van Cutsem E, Rougier P, et al. Addition of cetuximab to chemotherapy as first-line treatment for KRAS wild-type metastatic colorectal cancer: pooled analysis of the CRYSTAL and OPUS randomised clinical trials. *Eur J Cancer* 2012;48:1466-75.
7. Smith CG, Fisher D, Claes B, et al. Somatic profiling of the epidermal growth factor receptor pathway in tumors from patients with advanced colorectal cancer treated with chemotherapy ± cetuximab. *Clin Cancer Res* 2013;19:4104-13.
8. Peeters M, Oliner KS, Parker A, et al. Massively parallel tumor multigene sequencing to evaluate response to panitumumab in a randomized phase III study of metastatic colorectal cancer. *Clin Cancer Res* 2013;19:1902-12.
9. Peeters M, Oliner KS, Price TJ, et al. Updated analysis of KRAS/NRAS and BRAF mutations in study 20050181 of panitumumab (pmab) plus FOLFIRI for second-line treatment (tx) of metastatic colorectal cancer (mCRC). *J Clin Oncol* 2014;32:abstr 3568.
10. Seymour MT, Brown SR, Middleton G, et al. Panitumumab and irinotecan versus irinotecan alone for patients with KRAS wild-type, fluorouracil-resistant advanced colorectal cancer (PICCOLO): a prospectively stratified randomised trial. *Lancet Oncol* 2013;14:749-59.
11. Douillard JY, Oliner KS, Siena S, et al. Panitumumab-FOLFOX4 treatment and RAS mutations in colorectal cancer. *N Engl J Med* 2013;369:1023-34.
12. Sorich MJ, Wiese MD, Rowland A, et al. Extended RAS

- mutations and anti-EGFR monoclonal antibody survival benefit in metastatic colorectal cancer: a meta-analysis of randomized, controlled trials. *Ann Oncol* 2015;26:13-21.
13. National Comprehensive Cancer Network. Clinical Practice Guidelines in Oncology (NCCN Guidelines): colon cancer. Version 2.2016. Available online: [www.nccn.org/professionals/physician\\_gls/pdf/colon.pdf](http://www.nccn.org/professionals/physician_gls/pdf/colon.pdf).
  14. Rowland A, Dias MM, Wiese MD, et al. Meta-analysis of BRAF mutation as a predictive biomarker of benefit from anti-EGFR monoclonal antibody therapy for RAS wild-type metastatic colorectal cancer. *Br J Cancer* 2015;112:1888-94.
  15. Pietrantonio F, Petrelli F, Coinu A, et al. Predictive role of BRAF mutations in patients with advanced colorectal cancer receiving cetuximab and panitumumab: a meta-analysis. *Eur J Cancer* 2015;51:587-94.
  16. Rowland A, Dias MM, Wiese MD, et al. Reply: Comment on 'Meta-analysis of BRAF mutation as a predictive biomarker of benefit from anti-EGFR monoclonal-antibody therapy for RAS wild-type metastatic colorectal cancer'. *Br J Cancer* 2015;113:1635.
  17. Rowland A, Dias MM, Wiese MD, et al. Meta-analysis comparing the efficacy of anti-EGFR monoclonal antibody therapy between KRAS G13D and other KRAS mutant metastatic colorectal cancer tumours. *Eur J Cancer* 2016;55:122-30.
  18. Khambata-Ford S, Garrett CR, Meropol NJ, et al. Expression of epiregulin and amphiregulin and K-ras mutation status predict disease control in metastatic colorectal cancer patients treated with cetuximab. *J Clin Oncol*. 2007;25:3230-7.
  19. Smyth EC, Khan K, Cunningham D. AREG and EREG as Predictive Biomarkers for RAS Wild-Type Colorectal Cancer Treated With Panitumumab: A Fresh Approach to an Old Puzzle. *JAMA Oncol* 2016. [Epub ahead of print].
  20. Jacobs B, De Roock W, Piessevaux H, et al. Amphiregulin and epiregulin mRNA expression in primary tumors predicts outcome in metastatic colorectal cancer treated with cetuximab. *J Clin Oncol* 2009;27:5068-74.
  21. Pentheroudakis G, Kotoula V, De Roock W, et al. Biomarkers of benefit from cetuximab-based therapy in metastatic colorectal cancer: interaction of EGFR ligand expression with RAS/RAF, PIK3CA genotypes. *BMC Cancer* 2013;13:49.
  22. Baker JB, Dutta D, Watson D, et al. Tumour gene expression predicts response to cetuximab in patients with KRAS wild-type metastatic colorectal cancer. *Br J Cancer* 2011;104:488-95.
  23. Jonker DJ, Karapetis CS, Harbison C, et al. Epiregulin gene expression as a biomarker of benefit from cetuximab in the treatment of advanced colorectal cancer. *Br J Cancer* 2014;110:648-55.
  24. Seligmann JF, Elliott F, Richman SD, et al. Combined Epiregulin and Amphiregulin Expression Levels as a Predictive Biomarker for Panitumumab Therapy Benefit or Lack of Benefit in Patients With RAS Wild-Type Advanced Colorectal Cancer. *JAMA Oncol* 2016. [Epub ahead of print].
  25. Simon RM, Paik S, Hayes DF. Use of archived specimens in evaluation of prognostic and predictive biomarkers. *J Natl Cancer Inst* 2009;101:1446-52.

**Cite this article as:** Rowland A, Karapetis CS, Mangoni AA, Wiese MD, Kichenadasse G, McKinnon RA, Sorich MJ. Clinical translation of predictive markers for anti-EGFR monoclonal antibody therapy in metastatic colorectal cancer. *Transl Cancer Res* 2016;5(S1):S31-S34. doi: 10.21037/tcr.2016.05.19