

Overexpression of KLHL22 correlates with poor prognosis in patients with triple-negative breast cancer

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Background: Kelch-like family member 22 (*KLHL22*) is a protein-coding gene that is responsible for several Mendelian diseases and has been reported to promote tumorigenesis and aging. The purpose of this study was to investigate its expression in triple-negative breast cancer (TNBC) and its prognostic significance.

Methods: Immunohistochemistry (IHC) was performed to examine the expression levels of KLHL22 in 146 patients with TNBC. The Chi-squared test was used to analyze the correlations between KLHL22 expression level and clinicopathological features, and the Kaplan-Meier survival analysis and Cox multivariate regression model were used to analyze the prognostic significance of KLHL22 in patients with TNBC.

Results: The results of immunohistochemical analysis showed that the high expression rate of KLHL22 protein in TNBC was 56.85% (83/146). Further analysis revealed a significantly positive correlation (P<0.05) between KLHL22 expression and primary tumor and regional lymph node status, clinical stage, and relapse. Kaplan-Meier survival analysis revealed that patients with low KLHL22 expression had a longer mean survival time than those with high KLHL22 expression (147.93 *vs.* 90.1 months; P<0.05). In the multivariate analysis, KLHL22 level, P53 expression, and clinical stage were found to be independent prognostic factors for overall survival (P<0.05), while clinical stage and KLHL22 level were independent prognostic factors for progression-free survival (P<0.05).

Conclusions: The present study concludes that KLHL22 may serve as a biomarker for poor prognosis in patients with TNBC.

Keywords: Triple-negative breast cancer (TNBC); Kelch-like family member 22 (KLHL22); immunohistochemistry (IHC); prognosis

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Introduction

According to the 2020 Global Cancer Statistics Report, an estimated 19.3 million new cancer cases and nearly 10.0 million cancer deaths had occurred worldwide. Female breast cancer has surpassed lung cancer as the most commonly diagnosed cancer and is now ranked as the fifth leading cause of cancer mortality worldwide (1). Triple-negative breast cancer (TNBC) lacks human epidermal growth factor receptor 2 gene amplification and hormone receptor expression and is a subtype of breast tumor that accounts for approximately a quarter of newly diagnosed breast tumors (2). TNBC generally has a poor prognosis, high systemic recurrence rate, and refractoriness to conventional therapy, regardless of the choice of adjuvant treatment (3). Therefore, more effective treatment regimens and prognostic indicators are urgently needed (4).

Kelch-like family member 22 (KLHL22) is a BTB (Broad-complex, Tramtrack, and Bric-à-brac) adaptor protein that usually forms a functional cullin-RING E3 ubiquitin ligase complex with scaffold protein cullin 3 (CUL3) and ring-finger RING box protein 1 (5-7). Studies have shown that inactivation of KLHL22 homologs in nematodes can prolong their lifespan as the protein promotes tumorigenesis and aging by activating mechanistic target of rapamycin in complex 1 (mTORC1) signaling (8). In multiple breast cancer cell lines, the protein level of KLHL22 was elevated, and its deletion prevented mTORC1 activation and tumorigenesis (8).

However, studies on KLHL22 in the context of TNBC are rare, and the clinical prognostic factors of KLHL22 in patients with breast cancer remain unclear. In this study, we explored KLHL22 protein expression in TNBC via immunohistochemistry (IHC) and assessed its relationship

Highlight box

Key findings

• We report that the Kelch-like family member 22 (KLHL22) protein can serve as a prognostic marker for triple-negative breast cancer (TNBC).

What is known and what is new?

- TNBC has biological characteristics that make it prone to metastasis and recurrence.
- KLHL22 can predict the prognosis of TNBC.

What is the implication, and what should change now?

• Further research is needed on the role of KLHL22 in TNBC.

with TNBC prognosis using statistical analysis methods. We present this article in accordance with the REMARK reporting checklist (available at https://tcr.amegroups.com/article/view/10.21037/tcr-23-654/rc).

Methods

Patients and tissue specimens

In this retrospective study, 146 patients with TNBC who underwent surgical treatment between 2005 and 2013 at Sun Yat-sen University Cancer Center (Guangzhou, China) were included. The criteria for case selection were based on the pathological diagnosis of TNBC and complete follow-up data were included. Among the patients with TNBC, 75 (51.4%) were aged \leq 49 years and 71 (48.6%) were aged >49 years. In these cases, 112 patients (76.7%) were diagnosed at the early stages (I and II), and the remaining 34 (23.3%) were diagnosed at the late stages (III and IV). Table 1 details the clinicopathological features (age, tumor size, Ki67 level, P53 expression, clinical stage, and relapse) of the patients. Tumor staging was defined according to the tumor node metastasis classification system proposed by the American Joint Committee on Cancer/International Union Against Cancer. Tumor differentiation was based on the Edmondson and Steiner criteria.

IHC

Immunohistochemical staining of KLHL22 was performed using the standard EnVision procedure. First, we sequentially cut the paraffin-embedded tissue blocks into 3 µm thick sections and then dried and deparaffinized the slides with the tissue flakes in xylene. Next, the slides were rehydrated through graded alcohol and immersed in 3% hydrogen peroxide for 10 min to block endogenous peroxidase activity. After that, the antigen was retrieved by pressure cooking for 3 min in an ethylenediaminetetraacetic acid pH 9.0 antigen retrieval solution. To reduce nonspecific reactions, the slides were incubated with 5% bovine serum albumin for 15 min, after which they were incubated with the mouse anti-KLHL22 monoclonal antibody (1:500 dilution; abs132394; Absin Biotechnology Company, Shanghai, China) for 50 min at 37 °C. The slides were then incubated with the secondary antibody (Envision, k5007; Dako, Denmark) for 30 min at 37 °C, followed by staining with 3,3-diaminobenzidine. Finally, the sections were counterstained with Mayer's hematoxylin, dehydrated, and

Variable	All cases	Expression k	Expression KLHL22, n (%)		
		Low expression	High expression	P value	
Age (year)				0.261	
≤49	75	46 (61.3)	29 (38.7)		
>49	71	37 (52.1)	34 (47.9)		
Tumor size (cm)				0.826	
≤2.5	78	45 (57.7)	33 (42.3)		
>2.5	68	38 (55.9)	30 (44.1)		
рТ				0.017	
T1+2	127	77 (60.6)	50 (39.4)		
T3+4	19	6 (31.6)	13 (68.4)		
pN				0.016	
N0	77	51 (66.2)	26 (33.8)		
N1	69	32 (46.4)	37 (53.6)		
рМ				0.192	
M0	142	82 (57.7)	60 (42.3)		
M1	4	1 (25.0)	3 (75.0)		
Clinical stage				0.004	
1+11	112	71 (63.4)	41 (36.6)		
III+IV	34	12 (35.3)	22 (64.7)		
Ki67				0.654	
≤15	61	36 (59.0)	25 (41.0)		
>15	85	47 (55.3)	38 (44.7)		
P53				0.485	
Yes	95	56 (58.9)	39 (41.1)		
No	51	27 (52.9)	24 (47.1)		
Relapse				0.001	
Yes	16	3 (18.8)	13 (81.3)		
No	130	80 (61.5)	50 (38.5)		

Table 1 Correlation between the clinicopathologic variables and expression of KLHL22 in TNBC

KLHL22, Kelch-like family member 22; TNBC, triple-negative breast cancer.

mounted. For the negative control, the primary antibody was replaced with normal mouse immunoglobulin G (IgG).

IHC evaluation

Positive KLHL22 staining was mostly observed in the cytoplasm and were brown or yellow in color. The

immunoreactivity score was evaluated by positive cell number and positive intensity score. We first evaluated the percentage of positive tumor cells by taking five fields per slide to count the percentage in 5% increments from 0 to 100%, with 0 indicating negative staining. A positive intensity score was then defined in four degrees as follows: negative [0], weak [1], moderate [2], and strong [3]. Total scores were obtained from the intensity and proportion (0-300 scores). Then, we used the receiver operating characteristic (ROC) curve to determine the cut-off value for KLHL22 expression level in TNBC. Three independent senior pathologists who were blinded to the clinicopathological data of the patients scored KLHL22 expression. All analyses were performed by at least two pathologists in a single laboratory. The value was selected when at least two were in agreement with the results of the diagnosis. If they had different opinions, the diagnosis would be repeated and discussed to reach a consensus. The diagnostic coincidence rate was approximately 75%, which proves that this method has high repeatability.

Statistical analysis

All statistical analyses were performed using SPSS software (version 21.0; SPSS Inc., Chicago, IL, USA). ROC analysis was used to determine the cut-off value for KLHL22. The Chi-squared test was used to evaluate the relationship between the clinicopathological characteristics of patients with TNBC and KLHL22 protein levels. We used the Kaplan-Meier method with the log-rank test to evaluate the survival of patients with TNBC for univariate analysis. The independent prognostic factors were identified using Cox regression analysis. Statistical significance for two-tailed hypothesis was set at P value <0.05.

Ethics approval and consent to participate

The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by the Institute Research Medical Ethics Committee of Sun Yat-sen University Cancer Center (No. SZR2021-053). Written informed consent for participation was not required due to the retrospective nature of this study. All samples were anonymized.

Results

Cut-off value for KLHL22 expression

Immunohistochemical analysis showed that in TNBC, KLHL22 was mainly expressed in the cytoplasm (*Figure 1*). The cut-off score for positive KLHL22 expression was determined using ROC curves. According to previously reported methods, the point with maximum specificity and sensitivity was selected as the cut-off point (9). The sensitivity and specificity for each outcome were plotted as follows: survival outcome (*Figure 2A*), primary tumor (pT) status (*Figure 2B*), regional lymph node (pN) status (*Figure 2C*), clinical stage (*Figure 2D*), relapse (*Figure 2E*), and tumor size (*Figure 2F*). Based on the ROC curve analysis, the cut-off value for the expression of KLHL22 protein was 190 (P<0.001). Using this cut-off, a high level of KLHL22 was detected in 56.85% (83/146) of the TNBC samples.

Association between KLHL22 expression and clinicopathologic features of patients with TNBC

Chi-squared analysis demonstrated that KLHL22 expression was significantly correlated with pT and pN status, clinical stage, and relapse (*Table 1*, P<0.05). There was no significant correlation between KLHL22 expression and other clinicopathological features, such as patient age, tumor size, metastasis (pM) status, and P53 and Ki67 expression (*Table 1*, P>0.05).

Correlation between KLHL22 expression status, clinicopathological characteristics, and patients survival

Univariate survival analysis revealed a significant effect of clinicopathological prognostic parameters, such as pT (P=0.001), pN (P<0.001), clinical stage (P<0.001), and relapse (P<0.001), on prognosis. The results also demonstrated that KLHL22 expression (P<0.001) could also be used as an important factor in evaluating the prognosis of TNBC (*Table 2*). Kaplan-Meier survival analysis (*Figure 3*) showed that the mean survival time of the low KLHL22 group was significantly longer than that of the high KLHL22 group (147.93 vs. 90.1 months, P<0.001; *Figure 3A*).

We further evaluated the relationship between KLHL22 protein expression and prognosis of patients with TNBC in different clinicopathological subgroups. The results showed that high KLHL22 expression was associated with poor prognosis in the no relapse (P<0.001; *Figure 3B*), pT1+pT2 (P<0.001; *Figure 3C*), pN0 (P=0.002; *Figure 3E*), pN1+pN2+pN3 (P<0.001; *Figure 3F*), stage I+II (P<0.001; *Figure 3G*), stage III+IV (P<0.001; *Figure 3H*), Ki67 \leq 15% (P=0.002; *Figure 3I*), Ki67 >15% (P<0.001; *Figure 3J*), p53(-) (P=0.001; *Figure 3K*), and p53(+) (P<0.001; *Figure 3L*) groups. Regardless of the Ki67 expression rate (i.e., >15% or \leq 15%), high KLHL22 expression was associated with shorter overall survival. Similarly, regardless of whether p53 was negatively or positively expressed, the group with

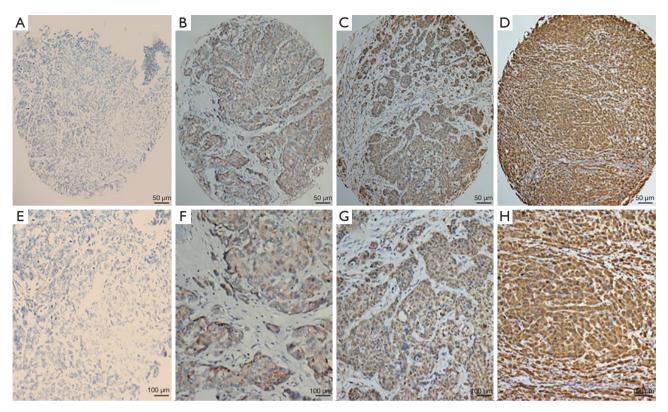


Figure 1 Expression of KLHL22 in TNBC tissues. (A,E) Negative expression; (B,F) low expression; (C,G) moderate expression; (D,H) strong expression. (A-D) magnification ×4; (E-H) magnification ×20. IHC was performed using standard EnVision method. KLHL22, Kelch-like family member 22; TNBC, triple-negative breast cancer; IHC, immunohistochemistry.

lower KLHL22 protein expression levels had longer overall survival. However, there was no significant relationship between KLHL22 expression and prognosis in the pT3+pT4 group (P=0.089; *Figure 3D*). In general, patients with low KLHL22 levels had a longer mean survival than those with high KLHL22 levels. It was associated with no relapse, pT1+pT2, pN0, pN1+pN2+pN3, stage I+II, stage III+IV, and Ki67 and p53 expression (*Figure 3*).

Independent prognostic factors for patents with TNBC

Multivariate analysis showed that P53 expression, clinical stage, and KLHL22 level were independent prognostic factors for overall survival (P<0.05), whereas clinical stage and KLHL22 level were independent prognostic factors for progression-free survival (P<0.05; *Table 3*). We performed Cox regression analysis for several factors that showed significant differences in the univariate analysis. As shown in *Table 3*, high KLHL22 level was an independent prognostic factor ratio

(HR): 10.41; 95% confidence interval (CI): 4.313–25.126; P<0.001] and progression-free survival (HR: 8.493; 95% CI: 2.210–32.642; P=0.002). At the same time, it was also found that P53 expression (P=0.024) and clinical stage (P<0.001) were independent prognostic factors for overall patient survival, while clinical stage (P<0.001) was an independent prognostic factors for progression-free survival (*Table 3*).

Discussion

As a member of the KLHL family, KLHL22 encodes a group of proteins that typically contain a BTB/POZ domain and is most visible as a diffuse cytoplasmic signal (7,10). As an adaptor of the CUL3-based E3 ligase, KLHL22 activates amino acid-dependent mTORC1 signaling to promote aging and tumorigenesis (8). KLHL22 can also affect translational modification through the role of CUL3-based ubiquitin ligase, thereby affecting cell function (6,11,12). *KLHL22* mRNA levels are elevated in prostate cancer, breast cancer, and melanoma, according to the Oncomine

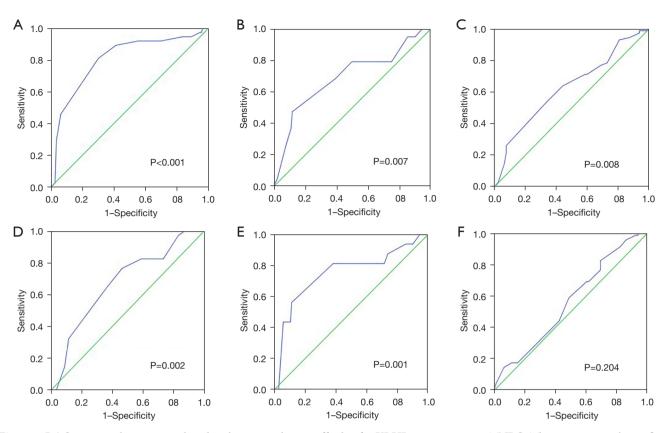


Figure 2 ROC curve analysis was employed to determine the cut-off value for KLHL22 expression in TNBC. The sensitivity and specificity for each outcome were plotted: survival outcome (A), pT status (B), pN status (C), clinical stage (D), relapse (E), tumor size (F). ROC, receiver operating characteristic curve.

database. KLHL22 protein levels are elevated in multiple breast cancer cell lines and breast tumor tissues of patients, and *KLHL22* deletion prevents mTORC1 activation and tumorigenesis in breast cancer cells (8). However, the upstream molecular mechanism of KLHL22 is not thoroughly studied, and its role in tumor development remains unclear.

In this study, a high level of KLHL22 was detected in the TNBC samples. We also found a significant relationship between KLHL22 expression and clinicopathological factors, such as pT and pN status, clinical stage, and relapse. Among patients with TNBC, those with low KLHL22 levels had longer mean survival than those with high KLHL22 levels.

Ki67 expression is related to the proliferation and growth of many tumor cells and is used as a proliferation marker in routine pathological examinations (13). Although the value of Ki67 in breast cancer has been controversial, recent guidelines have included it in pathologic tests as a biological marker (14). Ki67 has also been proposed as a biological tumor and prognostic marker in breast cancer (15). In this study, we detected KLHL22 and Ki67 expression in TNBC using IHC and found that high KLHL22 expression was significantly associated with poor prognosis in the Ki67 \leq 15% and Ki67 >15% groups. This provides evidence for its potential role in TNBC.

In addition, we found that p53 expression was an independent prognostic factor for the overall survival of patients with TNBC. Moreover, a high level of KLHL22 was a prognostic factor in patients with p53(+) and p53(-) expression. However, the relationship between KLHL22 and p53 remains unclear. Jeong found that Daxx-mediated repression of ETS1- and p53-dependent transcription was reversed after expression of SPOP with CUL3 (11), which might explain the relationship between p53 expression and KLHL22. Li *et al.* (16) showed that p53 is a candidate gene for poor prognosis in patients with TNBC. This is consistent with the results of our study, which showed that

Table 2 Univariate anal	lysis of clinicopatholog	gic variables in 146	patients with TNBC	(log-rank test)

Variable	All cases	Mean survival (months)	χ^2	P value
Age (year)			0.1	0.752
≤49	75	126.01		
>49	71	123.31		
Tumor size (cm)			1.22	0.269
≤2.5	78	129.64		
>2.5	68	119.03		
рТ			10.103	0.001
T1+2	127	129.75		
T3+4	19	85.36		
pN			21.471	<0.001
N0	77	145.06		
N1	69	102.95		
рМ			1.909	0.167
M0	142	125.67		
M1	4	68.5		
Clinical stage			34.255	<0.001
1+11	112	138.43		
III+IV	34	77.37		
Ki67			0.464	0.496
≤15	61	128.52		
>15	85	108.96		
P53			2.677	0.102
Yes	95	11.75		
No	51	135.82		
Relapse			45.469	<0.001
Yes	16	49.19		
No	130	133.91		
KLHL22			41.476	<0.001
High	63	90.1		
Low	83	147.93		

TNBC, triple-negative breast cancer.

p53 expression was an independent prognostic factor for the overall survival of patients with TNBC.

Chen *et al.* found that the CUL3-KLHL22 E3 ubiquitin ligase promotes the degradation of DEP domain containing 5 (DEPDC5), an essential subunit of GTPase-activating

protein (GAP) Activity TOward Rags 1 (GATOR1), and K48-linked polyubiquitination in response to amino acids. The GATOR1 complex, which is composed of DEPDC5, NPRL3, and NPRL2, exhibits GAP activity to inactivate RAG GTPases under amino acid-deficient conditions (8).

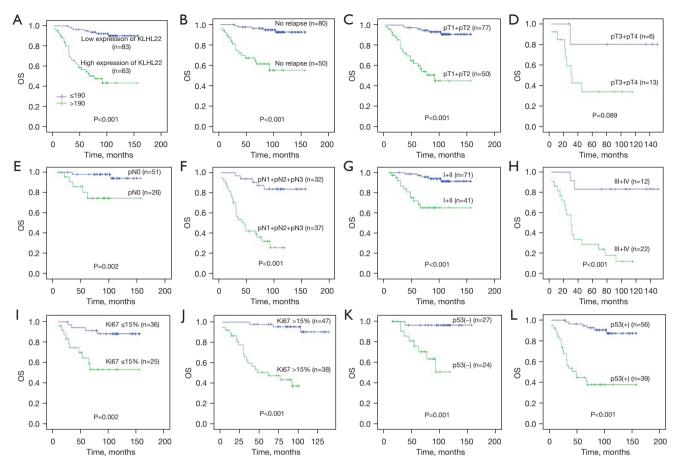


Figure 3 Kaplan-Meier survival analysis of KLHL22 expression in each different patient with TNBC. Total cohort (A), no relapse (B), pT1+pT2 (C), pT3+pT4 (D), pN0 status (E), pN1+pN2+pN3 (F), stage I+II (G), stage III+IV (H), Ki67 ≤15% (I), Ki67 >15% (J), p53(-) (K) and p53(+) (L). OS, overall survival rate; TNBC, triple-negative breast cancer.

Table 3 Multivariate survival analyses of clinicopathologic variables in TNBC patients

Veriele		OS		PFS		
Variable	HR	95% CI	P value	HR	95% CI	P value
Age (>49 <i>vs.</i> ≤49) (year)	0.602	0.300-1.207	0.152	0.52	0.185–1.466	0.216
P53 (yes <i>vs.</i> no)	2.441	1.125–5.299	0.024	0.633	0.234-1.714	0.368
Tumor size (>2.5 vs. ≤2.5) (cm)	0.847	0.424-1.691	0.637	0.74	0.254–2.154	0.581
Clinical stage (III+IV vs. I+II)	4.596	2.260-9.345	<0.001	3.093	1.062-9.008	0.038
KLHL22 (high vs. low)	10.41	4.313–25.126	<0.001	8.493	2.210-32.642	0.002

TNBC, triple-negative breast cancer; OS, overall survival; PFS, progression-free survival; HR, hazard ratio; CI, confidence interval.

RAG GTPases can recruit mTORC1 to lysosomes in response to amino acids (17). mTORC1 is a primary regulator of cell growth (18) that is unregulated and is associated with cancer. Deregulation of mTORC1 has been associated with carcinogenesis (19,20), and this explains how KLHL22 activates amino acid-dependent mTORC1 signaling to promote tumors. This is consistent with our finding that KLHL22 is an oncoprotein in TNBC. In addition, it has been reported that KLHL22 promoted breast tumorigenesis by activating mTORC1 signaling (8). Liu et al. (21) found that KLHL22 expression was upregulated in human malignant melanoma tissues and demonstrated that its overexpression promoted malignant melanoma cell growth in vitro. These studies concluded that KLHL22 functions as an oncogene, and these results are consistent with ours. Overexpression of KLHL22 by tumor cells resulted in a poor prognosis for these patients. The overall and progression-free survival rates were significantly longer in the low KLHL22 group than in the high KLHL22 group. Overall, KLHL22 could be used as an independent prognostic factor to predict the overall and progression-free survival of patients with TNBC, as evidenced by our clinical data and expression studies. However, a study by Song et al. (22) found that KLHL22 protein levels in clinicopathological tissue samples of patients were lower than those in normal tissues, and they further explained that KLHL22 acts as a tumor suppressor and regulates proliferation and epithelial-mesenchymal transition through the Wnt/β-catenin signaling pathway in colorectal cancer cells. In addition, Zhou et al. (23) revealed that KLHL22 plays a crucial role in maintaining the programmed cell death protein 1 (PD-1) expression balance and preventing excessive suppression of T cells. KLHL22 deficiency inhibits the antitumor response of T cells and promotes tumor progression through an excessive accumulation of PD-1. KLHL22 may function as a tumor suppressor gene; however, its role in tumor immunity and tumorigenesis warrants further study. Furthermore, this study is limited to single-center data and needs to be expanded to a multi-center validation analysis. Additionally, the molecular mechanism of KLHL22 in breast cancer needs to be further explored.

Conclusions

In summary, detection of KLHL22 expression via IHC is an effective tool to identify the prognosis of patients with TNBC. Our study also provides evidence that KLHL22 may be a potential therapeutic target for TNBC.

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Footnote

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Data Sharing Statement: Available at https://tcr.amegroups. com/article/view/10.21037/tcr-23-654/dss

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Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at https://tcr.amegroups.com/article/view/10.21037/tcr-23-654/coif). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). The study was approved by the Institute Research Medical Ethics Committee of Sun Yatsen University Cancer Center (No. SZR2021-053). Written informed consent for participation was not required due to the retrospective nature of this study.

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Translational Cancer Research, Vol 13, No 2 February 2024

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