

# EV-based EGFR genotyping in bronchoalveolar lavage fluid—a step forward from blood?

# Wonjun Ji<sup>1</sup>, Chang Min Choi<sup>1,2</sup>, Jae Cheol Lee<sup>2</sup>

<sup>1</sup>Department of Pulmonary and Critical Care Medicine, <sup>2</sup>Department of Oncology, University of Ulsan College of Medicine, Asan Medical Center, Seoul, Korea

*Correspondence to:* Jae Cheol Lee, MD, PhD. Department of Oncology, University of Ulsan College of Medicine, Asan Medical Center, 88 Olympic-ro, 43-gil, Songpa-gu, Seoul 05505, Korea. Email: jclee@amc.seoul.kr.

Provenance and Peer Review: This article was commissioned by the Editorial Office, Translational Lung Cancer Research. This article did not undergo external peer review.

Comment on: Hur JY, Lee JS, Kim IA, et al. Extracellular vesicle-based EGFR genotyping in bronchoalveolar lavage fluid from treatment-naive nonsmall cell lung cancer patients. Transl Lung Cancer Res 2019;8:1051-60.

Submitted Feb 04, 2020. Accepted for publication Mar 02, 2020. doi: 10.21037/tlcr.2020.03.24 View this article at: http://dx.doi.org/10.21037/tlcr.2020.03.24

The efficacy of osimertinib aiming at T790M-mediated EGFR-TKI resistance was proven in a large, randomized, phase III trial (AURA 3 trial) leading to becoming the standard treatment in T790M-positive cases following the 1<sup>st</sup>-line EGFR-TKI failure (1,2). However, the demonstration of T790M is mandatory to use this drug, which poses a clinical problem related with re-biopsy. Sometimes, the procurement of adequate tissue through rebiopsy is very difficult or even unable to do according to the location and size of tumor (3,4). The poor performance status of several patients is another obstacle for repeated tissue biopsy. In these cases, the less invasive liquid biopsy with cell-free DNA (cfDNA) in blood which had been already approved for the clinical application can be a reasonable alternative (5). Nonetheless, the analysis of tissue sample is preferentially recommended because of the high false negative rate of the plasma cfDNA T790M testing at present despite its definite advantages and future promising perspectives (1).

The composition of EVs which are released by exocytosis from tumor cells may reflect that of parent cells. They contain various kind of components such as proteins, lipids, RNAs and even double-stranded DNA (6). In addition, because of their stability which is secured by protecting envelope, EVs can be more reliable sources than plasma cfDNA for analysis of genetic materials. In fact, the combined use of RNA in EVs and cfDNA increased the sensitivity for EGFR mutation detection in plasma, especially in the subgroup of M0/M1a disease with low levels of cfDNA (7). As for T790M of which allele frequency is lower than that of sensitizing EGFR mutations such as 19 deletion and L858R resulting in the low detection rate, an analysis of pooled AURA extension and AURA2 studies showed 61% of sensitivity by the cobas EGFR Mutation Test v2 using cfDNA (8) while a test using RNA/DNA in EVs and cfDNA resulted in 92% of sensitivity (9).

Hur et al. ventured to use EV DNA in bronchoalveolar lavage fluid (BALF) rather than in plasma for the detection of EGFR mutation (10). It seems to be a clever attempt because BALF would have much more tumor-derived EVs than plasma considering the vicinity to tumor itself although the bronchoscopic approach is required to retrieve the BALF. The average sensitivity and specificity of BALF EVbased EGFR genotyping were 76% and 87%, respectively, while the sensitivity improved according to the increased stage. Surprisingly, BALF EV-based EGFR typing found all tissue-proven EGFR mutant cases (n=31) and detected 6 additional mutant cases in stage IV. The concordance rate was 79% in stage I, 100% in stage II, 74% in stage III, and 92% in stage IV. As TNM stage advanced, especially in the presence of metastasis, concordance rate significantly increased (P<0.05).

These are very impressive results which might be contributed by the superiority of EV DNA to cfDNA and BALF to plasma. Authors exhibited that BALF EV-based EGFR typing could find 11 more EGFR mutation-positive patients compared to the conventional tissue/cytologybased test. However, we don't know whether these patients truly have EGFR mutation or not because the results about drug response are not provided in this study. Nevertheless, as we recognize that false negative results can be obtained in cases of the low tumor cell proportion in tissue/cytology samples, this BALF EV-based test has the potential to find some of those missing patients by conventional approach. For the clinical application in near future, it should be proven by more confirmatory studies as well as expanding studies to demonstrate the performance for detection of T790M along with the clinical response to osimertinib.

Furthermore, it is noteworthy that BALF EV EGFR testing in stage I lung cancer showed the relatively high positive predictive value of 85.7% despite the low sensitivity of 35.3%. Physicians often face the difficulty in making a decision how to manage the slowly growing ground-glass nodule (GGN) with low to intermediate risk of cancer because the tissue diagnosis of GGN is very difficult in most cases. According to the study by Kobayashi et al., three fourths of resected GGNs were positive for EGFR, KRAS, ALK or HER2 mutations (11). Therefore, considering the high positive predictive value of BALF EV genotyping, the detection of mutation by this method may be helpful to direct treatment to the surgical resection rather than the watchful follow-up. The low sensitivity could be improved by the use of techniques with higher sensitivity such as nextgeneration sequencing and droplet digital PCR.

## **Acknowledgments**

Funding: None.

## Footnote

*Conflicts of Interest:* All authors have completed the ICMJE uniform disclosure form (available at http://dx.doi. org/10.21037/tlcr.2020.03.24). The authors have no conflicts of interest to declare.

*Ethical Statement:* The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Open Access Statement: This is an Open Access article

distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the noncommercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: https://creativecommons.org/licenses/by-nc-nd/4.0/.

#### References

- Mok TS, Wu YL, Ahn MJ, et al. osimertinib or Platinum-Pemetrexed in EGFR T790M-Positive Lung Cancer. N Engl J Med 2017;376:629-40.
- National Comprehensive Cancer Network. NCCN Clinical Practice Guidelines in Oncology: Non-Small Cell Lung Cancer. V.1.2020. Available online: www. nccn.org/professionals/physician\_gls/PDF/nscl.pdf. November 6, 2019.
- Chouaid C, Dujon C, Do P, et al. Feasibility and clinical impact of re-biopsy in advanced non smallcell lung cancer: a prospective multicenter study in a real-world setting (GFPC study 12-01). Lung Cancer 2014;86:170-3.
- 4. Nosaki K, Satouchi M, Kurata T, et al. Re-biopsy status among non-small cell lung cancer patients in Japan: A retrospective study. Lung Cancer 2016;101:1-8.
- Jenkins S, Yang JC, Ramalingam SS, et al. Plasma ctDNA Analysis for Detection of the EGFR T790M Mutation in Patients with Advanced Non-Small Cell Lung Cancer. J Thorac Oncol 2017;12:1061-70.
- Reclusa P, Sirera R, Araujo A, et al. Exosomes genetic cargo in lung cancer: a truly Pandora's box. Transl Lung Cancer Res 2016;5:483-91.
- Krug AK, Enderle D, Karlovich C, et al. Improved EGFR mutation detection using combined exosomal RNA and circulating tumor DNA in NSCLC patient plasma. Ann Oncol 2018;29:700-6.
- Jenkins S, Yang JC, Ramalingam SS, et al. Plasma ctDNA Analysis for Detection of the EGFR T790M Mutation in Patients with Advanced Non-Small Cell Lung Cancer. J Thorac Oncol 2017;12:1061-70.
- Castellanos-Rizaldos E, Grimm DG, Tadigotla V, et al. Exosome-Based Detection of EGFR T790M in Plasma from Non-Small Cell Lung Cancer Patients. Clin Cancer Res 2018;24:2944-50.
- 10. Hur JY, Lee JS, Kim IA, et al. Extracellular vesicle-based

#### Translational Lung Cancer Research, Vol 9, No 3 June 2020

EGFR genotyping in bronchoalveolar lavage fluid from treatment-naive non-small cell lung cancer patients. Transl Lung Cancer Res 2019;8:1051-60.

11. Kobayashi Y, Mitsudomi T, Sakao Y, et al. Genetic features

**Cite this article as:** Ji W, Choi CM, Lee JC. EV-based EGFR genotyping in bronchoalveolar lavage fluid—a step forward from blood? Transl Lung Cancer Res 2020;9(3):427-429. doi: 10.21037/tlcr.2020.03.24

of pulmonary adenocarcinoma presenting with groundglass nodules: the differences between nodules with and without growth. Ann Oncol 2015;26:156-61.