Original Article

A cross-trait study of lung cancer and its related respiratory diseases based on large-scale exome sequencing population

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Background: Genome-wide association studies (GWASs) explain the genetic susceptibility between diseases and common variants. Nevertheless, with the appearance of large-scale sequencing profiles, we could explore the rare coding variants in disease pathogenesis.

Methods: We estimated the genetic correlation of nine respiratory diseases and lung cancer in the UK Biobank (UKB) by linkage disequilibrium score regression (LDSC). Then, we performed exome-wide association studies at single-variant level and gene-level for lung cancer and lung cancer-related respiratory diseases using the whole-exome sequencing (WES) data of 427,934 European participants. Cross-trait meta-analysis was conducted by association analysis based on subsets (ASSET) to identify the pleiotropic variants, while in-silico functional analysis was performed to explore their function. Causal mediation analysis was used to explore whether these pleiotropic variants lead to lung cancer is mediated by affecting the chronic respiratory diseases.

Results: Five respiratory diseases [emphysema, pneumonia, asthma, chronic obstructive pulmonary disease (COPD), and fibrosis] were genetically correlated with lung cancer. We identified 102 significant independent variants at single-variant levels for lung cancer and five lung cancer-related diseases. 15:78590583:G>A (missense variant in *CHRNA5*) was shared in lung cancer, emphysema, and COPD. Meanwhile, 14 significant genes and 87 suggestive genes were identified in gene-based association tests, including *HSD3B7* (lung cancer), *SRSF2* (pneumonia), *TNXB* (asthma), *TERT* (fibrosis), *MOSPD3* (emphysema). Based on the cross-trait meta-analysis, we detected 145 independent pleiotropic variants. We further identified abundant pathways with significant enrichment effects, demonstrating that these pleiotropic genes were functional. Meanwhile, the proportion of mediation effects of these variants ranged from 6 to 23 (emphysema: 23%; COPD: 20%; pneumonia: 20%; fibrosis: 7%; asthma: 6%) through these five respiratory diseases to the incidence of lung cancer.

Conclusions: The identified shared genetic variants, genes, biological pathways, and potential intermediate causal pathways provide a basis for further exploration of the relationship between lung cancer and respiratory diseases.

Keywords: Exome-wide association study; lung cancer; respiratory diseases; rare variants; cross-trait

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Introduction

Lung cancer is one of the most common and fatal cancer. A few respiratory diseases have been described as possible risk factors for lung cancer (1,2), such as chronic obstructive pulmonary disease (COPD) and emphysema. Existing studies have confirmed that the close relationship between COPD and lung cancer is not just about shared smoking exposure, but is likely to reflect in part, a shared genetic susceptibility to chronic smoking-induced inflammation (3). By parity of reasoning, these respiratory diseases might share a common mechanism with the development of lung cancer. Studying them can identify shared genetic factors and provide an important foundation for lung cancer prevention and early warning.

In the past decade, genome-wide association study (GWAS) has thoroughly changed the perception of complex diseases and provided us with a number of significant and compelling risk variants (4,5). However, GWAS tends to concentrate on common variants, which usually have weak effect sizes and are difficult to map to causal genes (6). From the view of natural evolution, common variants appear early and have withstood natural selection pressure. Nevertheless, low-frequency variants have emerged late and have not been eliminated during human evolution and are more likely to be functional (7). Thus, rare variants may play an essential role in the development of disease.

In recent years, next-generation sequencing technologies have been iteratively upgraded, and large cohort studies have been scaled up. The UK Biobank (UKB) provides us with an unprecedented chance to explore the effect of both common and rare variants in human diseases (8-10). Compared with previous sequencing studies with limited sample size, large-scale exome sequencing population enables sufficient statistical power that could be used to identify rare coding variants associated with diseases (11,12).

In this study, we analyzed lung cancer and five respiratory diseases (asthma, COPD, emphysema, fibrosis, and pneumonia) with significant genetic correlations with lung cancer. We performed a comprehensive association study using exome sequencing data from 427,934 UKB participants of European ancestry at both variant-level and gene-level. Subsequent cross-trait meta-analysis, functional analyses, and causal mediation inference comprehensively depicted the genetic relationship between these five respiratory diseases and lung cancer.

Highlight box

Key findings

- We identified rare coding variants and genes associated with respiratory diseases, revealing genetic pleiotropy with lung cancer. It quantified mediation effects of these variants on lung cancer development via other respiratory diseases. Pathway and protein interaction analyses elucidated functional significance, highlighting potential therapeutic targets.

What is known and what is new?

- Genetic variants were associated with respiratory diseases and genetic pleiotropy was observed in complex diseases like lung cancer.
- Rare coding variants associated with respiratory diseases were identified, broadening understanding beyond common variants. Mediation effects of these variants on lung cancer via other respiratory diseases were quantified, revealing complex disease relationships.

What is the implication, and what should change now?

- This research underscores the complex genetic underpinnings of respiratory diseases and lung cancer, highlighting the need for a comprehensive understanding of genetic factors in disease development. Healthcare practices should integrate genetic screening for respiratory diseases, considering both common and rare variants.

Methods

Study population and phenotypes

The UKB is a large population-based prospective cohort

Table 1 Demographic characteristics and respiratory disease information in the UKB

Characteristics	N (%)
Sex	
Female	232,409 (54.31)
Male	195,525 (45.69)
Age (years)	
38–49	95,773 (22.38)
50–59	141,946 (33.17)
60–73	190,215 (44.45)
Smoke	
Ever	259,671 (60.68)
Never	166,895 (39.00)
Diseases	
Lung cancer	5,003 (1.17)
Asthma	38,627 (9.03)
COPD	17,561 (4.10)
Emphysema	4,002 (0.94)
Fibrosis	2,340 (0.55)
Pneumonia	21,424 (5.01)
Bronchiectasis	4,660 (1.09)
Acute bronchitis	14,364 (3.36)
Chronic bronchitis	1,382 (0.32)
Tuberculosis	654 (0.15)

UKB, UK Biobank; COPD, chronic obstructive pulmonary disease.

study from the UK with deep phenotypic and genetic data on approximately 500,000 individuals aged 40–69 years at enrollment (13). The work described herein was approved by the UKB under application No. 57471. All the phenotype data were accessed in March 2022. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013). Health-related outcomes were ascertained via individual record linkage to national cancer and mortality registries and hospital in-patient encounters. Cancer diagnoses were coded by the International Classification of Diseases version 10 (ICD-10) codes. Individuals with at least one recorded incident diagnosis of a borderline, in situ, or primary malignant cancer were defined as cases collected from data fields 41270 (diagnoses: ICD-10), 41202 (diagnoses: main ICD10), 40006 (type of cancer: ICD-10), and 40001 (primary cause of death: ICD-10). ICD-10 codes for other diseases included in the study is shown in Table S1.

The UKB provides detailed diseases follow-up information linked to whole-exome sequencing (WES) for approximately 450,000 participants (data field: 23148). We included 427,934 white European participants in this research, and detailed inclusion information is presented in Table S2. Ten respiratory diseases were analyzed (*Table 1*), including lung cancer (n=5,003), asthma (n=38,627), COPD (n=17,561), emphysema (n=4,002), idiopathic pulmonary fibrosis (n=2,340), pneumonia (n=21,424), bronchiectasis (n=4,660), acute bronchitis (n=14,364), chronic bronchitis (n=1,382), and tuberculosis (n=654).

Genetic correlation estimation

To find the respiratory diseases with significant genetic relationships with lung cancer, we used the linkage disequilibrium score regression (LDSC) (14) to assess the genetic correlation between each disease pair using the imputed genetic variants from the UKB (data field: 22828) (15). We conducted a genetic correlation analysis on ten respiratory diseases using the imputed genotype data from the Haplotype Reference Consortium (HRC) and UK10K haplotype resource (data field: 22828), and utilized the resulting summary data to estimate genetic correlations, which was not biased by sample overlap (15). Diseases significantly genetically correlated with lung cancer (nominal $P<0.05$) were included in further analyses.

Quality control for the genetic variants

WES data for UKB participants were generated using the IDT xGen v1 capture kit on the NovaSeq6000 platform. The UKB 450k release was performed with a Functional Equivalence specification that retained the original quality scores (OQFE protocol) in the CRAM files (16). The OQFE protocol mapped to a full GRCh38 reference version including all alternative contigs in an alt-aware manner. The OQFE CRAMs were then called for small variants with DeepVariant 0.0.10 to generate per-sample genome variant call formats (gVCFs), which were aggregated and joint-genotyped with GLnexus 1.2.6 to create a single multi-sample VCF [project VCF (pVCF)] for all UKB 450k samples. Genotype depth (DP) filters [single nucleotide variant (SNV) DP ≥7, indel DP ≥10] were applied prior to variant site filters requiring at least one variant genotype

passing an allele balance (AB) filter (heterozygous SNV AB >0.15, heterozygous indel <0.20). The detailed parameters were described in Category 170 of the UKB showcase. In addition, all the variants with call rate <90% and minor allele count (MAC) ≤1 were filtered out.

Single-variant association tests

Single-variant association tests were performed on 427,934 European participants. All the variants with MAC ≥10 were incorporated in the following association tests. We used SAIGE v1.1.4 (11) to conduct association tests based on logistic mixed models adjusting for age, gender, smoking status, and top five ancestry principal components to assess the association between the respiratory diseases and genetic variants (17,18). A genetic relationship matrix (GRM) was created to fit the model to eliminate the effect of kinship. We also included five principal components in mixed model to adjust for both population structures and non-genetic confounders (19). We included all the variants that passed quality control in the WES dataset, including loss-of-function (LOF), missense, synonymous, and a small proportion of non-coding variants. Variants passed the genome-wide significance threshold ($P \leq 5\times10^{-8}$) were defined as significant and independent variants were pruned out using the PLINK v1.9 clump function (-clump-r2 0.50, -clump-kb 500). We calculated the adjusted genomic inflation factor λ_{adj}. Because the genomic inflation factor increases with sample size, we rescaled the genomic inflation factor λ_{obs} to adjusted the genomic inflation factor λ_{adj} reflecting a standardized sample size of 1,000 cases and 1,000 controls based on the following formula:

$$\lambda_{adj} = 1 + (\lambda_{obs} - 1) \times \frac{\frac{1}{N_{cases}} + \frac{1}{N_{controls}}}{\frac{1}{1,000} + \frac{1}{1,000}} \quad [1]$$

If genomic inflation arose, SAIGE integrated linear mixed models to control for population structure and familial relationships, effectively mitigating genomic inflation.

Gene-based association tests for rare variants and ultra-rare variants

Afterward, we performed gene-level association tests using the SKAT-O method (20), which was implemented by SAIGE-GENE+ (21). Variants with minor allele frequency (MAF) ≤1% were considered rare, while variants with MAC ≤10 were considered ultra-rare. To improve power to detect the association signals, we performed tests for rare variants with different MAF cutoffs (MAF ≤1%, MAF ≤0.1%, and MAF ≤0.01%). According to the latest research that synonymous mutations may be strongly non-neutral (22), we considered all the functional annotations. Therefore, multiple variant sets with different MAF cutoffs and functional annotations (LOF, missense, and synonymous) were analyzed. We reported the association results with the lowest P value for one gene to collectively capture a wide range of genetic architectures (23). We used a P value threshold of $P \leq 1\times10^{-5}$ to report genes associated with these diseases.

Cross-trait meta-analysis for the respiratory diseases

We conducted cross-trait meta-analysis via the R package association analysis based on subsets (ASSET) (24). Briefly, ASSET explored all possible subsets of all six diseases (five lung cancer-related diseases and lung cancer) for the presence of association signals, resulting in the best combination of diseases to maximize the test statistic. According to the result of single-variant association tests, the variants with P value $\leq 1\times10^{-4}$ in any one of the six diseases association tests were included. Because the method explores all possible subsets of studies and evaluates fixed-effect meta-analysis-type test-statistics for each subset, to avoid excessive computational effort, we used a relatively lenient P value to comprehensively consider all suggestive association variants across the six respiratory diseases.

By ASSET, we achieved the P values of significance for the overall evidence of association of a variant across these diseases as well as the "best subset" that contributed to the overall association signal (24). Finally, all the independent variants with $P < 5\times10^{-8}$ were reported.

Intermediate pathways analysis

We were interested in whether there was a mediating effect of variants causing other respiratory diseases and consequently leading to lung cancer. Based on the cross-trait meta-analysis, we searched for variants shared between lung cancer and five other respiratory diseases. Then, we constructed polygenic scores (PGSs) for the shared variants found for each respiratory disease (25). PGS is not applied here for the purpose of disease risk prediction, but for the purpose of using the idea of PGS to comprehensively measure the impact of all shared variants and calculate the mediation effect using a unified indicator. The beta

coefficients of each variant was used as the weight in PGSs: $PRS = \sum \beta_i SNV_i$ (26). We calculated the area under the receiver operator characteristic curves (AUCs) of all PGSs used for mediation analyses using Bootstrap. Finally, we carried out mediation analyses by these polygenic risk scores and identified mediating effects for these five respiratory diseases. All the mediation analyses were performed by R package "mediation".

Genetic functional analysis

To explore further biological explanations and assess the biological functions of the pleiotropic genes, we conducted pathway enrichment analysis via Metascape (27). During this analysis, the gene list we detected were compared to thousands of gene sets defined by their involvement in specific biological processes, protein localization, pathway member, or other features (27). Pathway and process enrichment analysis had been carried out with the following ontology sources: Kyoto Encyclopedia of Genes and Genomes (KEGG), Gene Ontology (GO) biological processes, reactome gene sets, canonical pathways, and WikiPathways.

Metascape reported the terms with a P value <0.01, a minimum count of 3, and an enrichment factor >1.5 (the enrichment factor was the ratio between the observed counts and the counts expected by chance). Then the terms were grouped into clusters based on their membership similarities. Kappa scores were used as the similarity metric when clustering on the enriched terms (28), and sub-trees with similarity >0.3 were considered a cluster.

To further understand the protein-protein interactions, we used the Search Tool for the Retrieval of Interacting Genes-Proteins (STRING) database, which considered both physical interactions as well as functional associations (29). The protein-protein interaction network was clustered into different colors using k-means clustering.

We used the R software (version 4.2.0) for statistical analysis and graphing. All P values were two-sided.

Results

Genetic correlation of the respiratory diseases

Figure 1 depicts the study design and workflow. The cross-trait genetic correlation calculated by LDSC showed intricate relationships among respiratory diseases. Lung cancer was significantly genetically correlated with five respiratory diseases, including emphysema (r_g=0.61, P=0.0001), pneumonia (r_g=0.64, P=0.0018), asthma (r_g=0.24, P=0.0056), COPD (r_g=0.69, P=9×10^{-7}), and idiopathic pulmonary fibrosis (r_g=0.60, P=0.0285) (Figure S1). Therefore, we focused on the shared genetic basis for them and lung cancer in the subsequent exome sequencing analyses.

Single-variant association tests

In the single-variant association analysis, 102 independent loci mapped to 53 genes passed the genome-wide significance level (P<5×10^{-8}). Of them, six were associated with lung cancer, six were associated with COPD, six were associated with emphysema, three were associated with idiopathic pulmonary fibrosis, three were associated with pneumonia, and 88 were associated with asthma (*Figure 2*). The adjusted genomic inflation factor λ_{adj} did not suggest population stratification (Figure S2). Noteworthy, only one genomic region 15q25.1 had shared signals with lung cancer. The sentinel variant 15:78590583:G>A (missense, HGVSp: p.Asp398Asn) in *CHRNA5* were significant in lung cancer {odds ratio (OR) [95% confidence interval (CI)]: 1.22 (1.18, 1.26), P=8.41×10^{-20}}, emphysema [OR (95% CI): 1.22 (1.16, 1.28), P=1.50×10^{-14}], and COPD [OR (95% CI): 1.13 (1.10, 1.15), P=7.79×10^{-25}]. *CHRNA5* was related to the mechanism of nicotine addiction (30) in smoking that could lead to respiratory diseases (31,32).

In addition, the missense variant 11:1167980:C>T [MAF =4.0%, OR (95% CI): 2.21 (1.87, 2.61), P=1.98×10^{-20}] in *MUC5AC* was significant in fibrosis. The synonymous variant 6:32584335:A>G [MAF =9.3%, OR (95% CI): 1.18 (1.15, 1.21), P=2.20×10^{-37}] in *HLA-DRB1* was associated with asthma. Moreover, we identified 15 additional rare variants with MAF ≤1% in asthma, emphysema, and pneumonia. For example, the missense variant 9:5073770:G>T in *JAK2* [MAF =0.03%, OR (95% CI): 6.23 (3.69, 10.51), P=7.14×10^{-12}] was associated with pneumonia. These rare variants with large effects may play an essential role in the onset of respiratory diseases (33). All the association results for independent single variants with P<5×10^{-8} are shown in Table S3.

Gene-based association tests

We analyzed 18,184 protein-coding genes in the gene-based association tests and identified 14 significant genes (P≤1×10^{-5}) (*Figure 3*, Table S4). Among them, *HSD3B7*

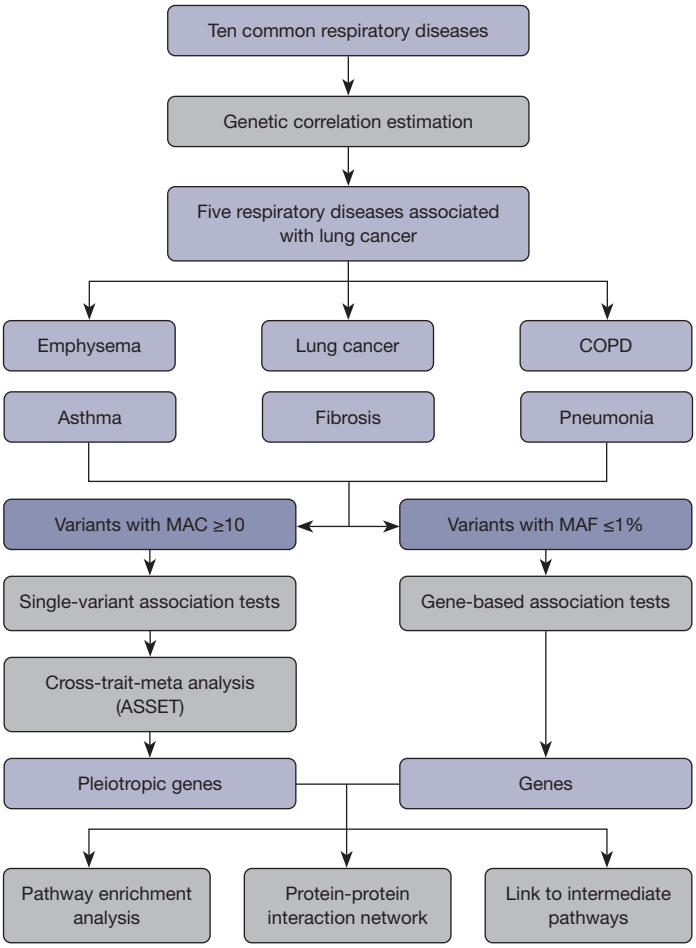

Figure 1 Workflow of this study. COPD, chronic obstructive pulmonary disease; MAC, minor allele count; MAF, minor allele frequency; ASSET, association analysis based on subsets.

($P=7.7\times10^{-7}$) and *TARM1* ($P=9.2\times10^{-6}$) were associated with lung cancer; *LACRT* ($P=6.8\times10^{-6}$) was associated with COPD; *MOSPD3* ($P=2.7\times10^{-6}$) was associated with emphysema; *TERT* ($P=3.8\times10^{-7}$) and *LMNA* ($P=1.1\times10^{-6}$) were associated with fibrosis; *SRSF2* ($P=7.4\times10^{-15}$) and *JAK2* ($P=2.5\times10^{-6}$) were associated with pneumonia; *TNXB* ($P=2.4\times10^{-7}$), *C6orf10* ($P=9.4\times10^{-7}$), *NOTCH4* ($P=8.2\times10^{-6}$), *HLA-DQA2* ($P=8.3\times10^{-6}$), *TTK* ($P=8.9\times10^{-6}$), and *SPINK7* ($P=9.4\times10^{-6}$) were associated with asthma. Moreover, 87 genes showed suggestive significance ($P\leq1\times10^{-4}$) (Table S4).

Shared genetic variants for the six respiratory diseases

A total of 781 independent variants that reached $P<10^{-4}$ in any disease were included in the cross-trait meta-analysis. Strong evidence supported the shared genetic foundation underlying these six diseases that 145 independent variants had $P\leq5\times10^{-8}$ (*Figure 4A*, table available at https://cdn.amegroups.cn/static/public/tlcr-24-4-1.xlsx). The number of variants that overlapped between disease pairs is demonstrated in *Figure 4B*. Best subset of these diseases that contributed to the overall association signal is shown in *Figure 4C*. It was illustrated that lung cancer and its related respiratory diseases were genetically linked.

In summary, the independent pleiotropic genes associated with any two or more of these six diseases are shown in *Figure 4D*. Intuitively, human leukocyte antigen (HLA) family made a remarkable contribution to genetic interactions among these diseases. The pleiotropic genes that overlapped with lung cancer with the largest OR among all the significant genes in cross-trait meta-analysis are displayed in *Table 2*. In the previous analysis,

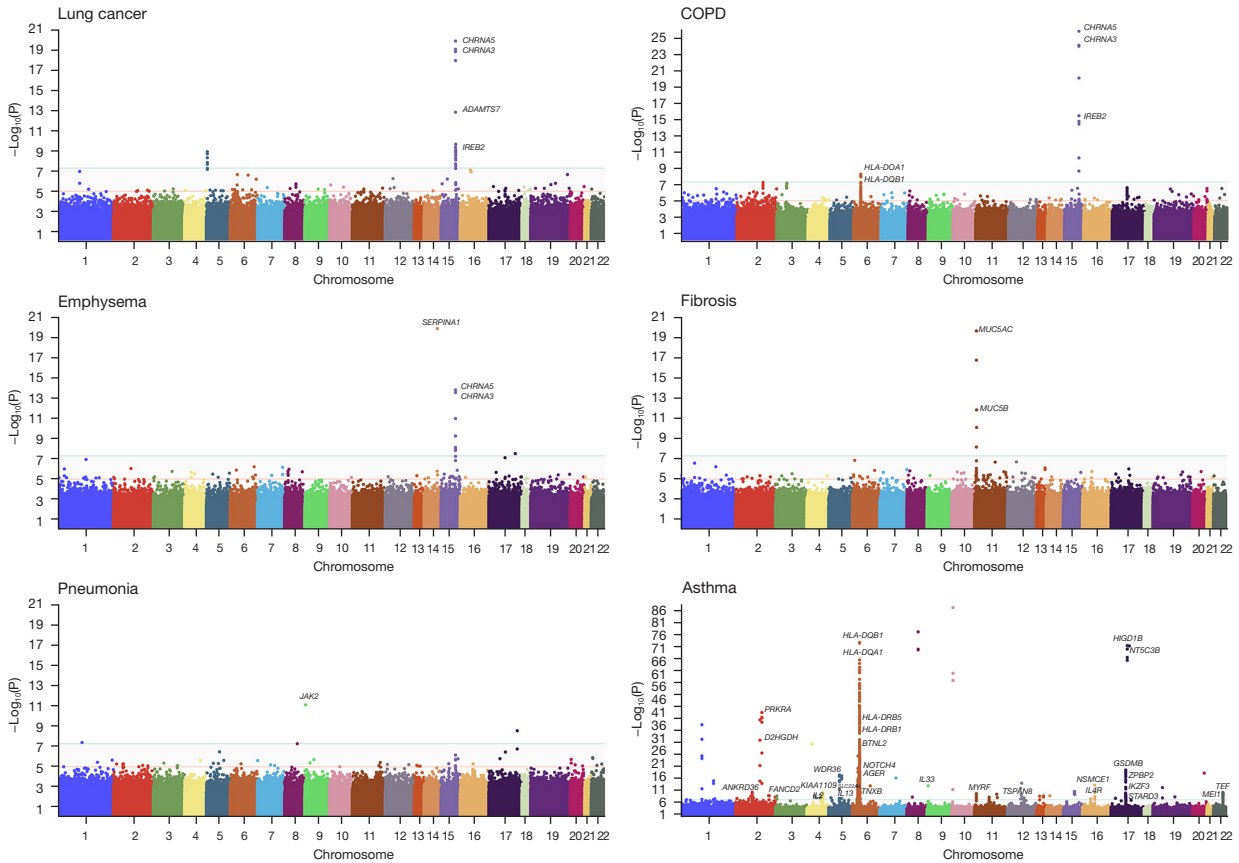

Figure 2 Manhattan plot for the single variant association results of the six lung cancer-related diseases. The green line indicates the genome-wide significance level ($P<5\times10^{-8}$). The red line indicates the suggestive significance level ($P<1\times10^{-5}$). The significant genes for each disease are labeled. COPD, chronic obstructive pulmonary disease.

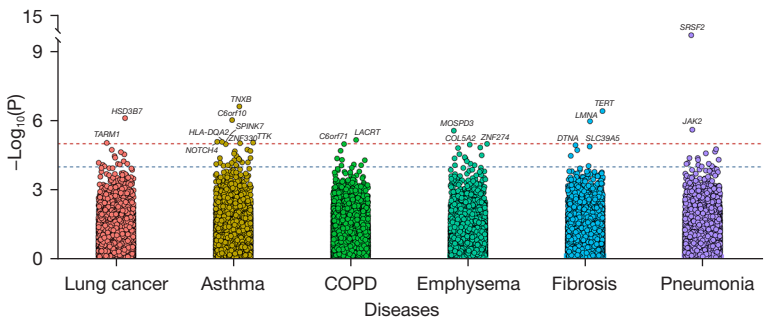

Figure 3 Point plot for the gene-based association results of the six lung cancer-related respiratory diseases. The red dash line indicates the gene-based significance level ($P<1\times10^{-5}$). The blue dash line indicates the suggestive significant level ($P<1\times10^{-4}$). All significant genes are labeled on the figure. COPD, chronic obstructive pulmonary disease.

few genes shared by respiratory diseases and lung cancer were found, but a large number of shared genes associated with lung cancer were found in this step. For example, missense variant 10:132909243:C>T in *CFAP46* (MAF $=1.2\times10^{-5}$), missense variant 1:38017675:C>T in *UTP11* (MAF $=1.8\times10^{-5}$), synonymous variant 19:42079623:C>T

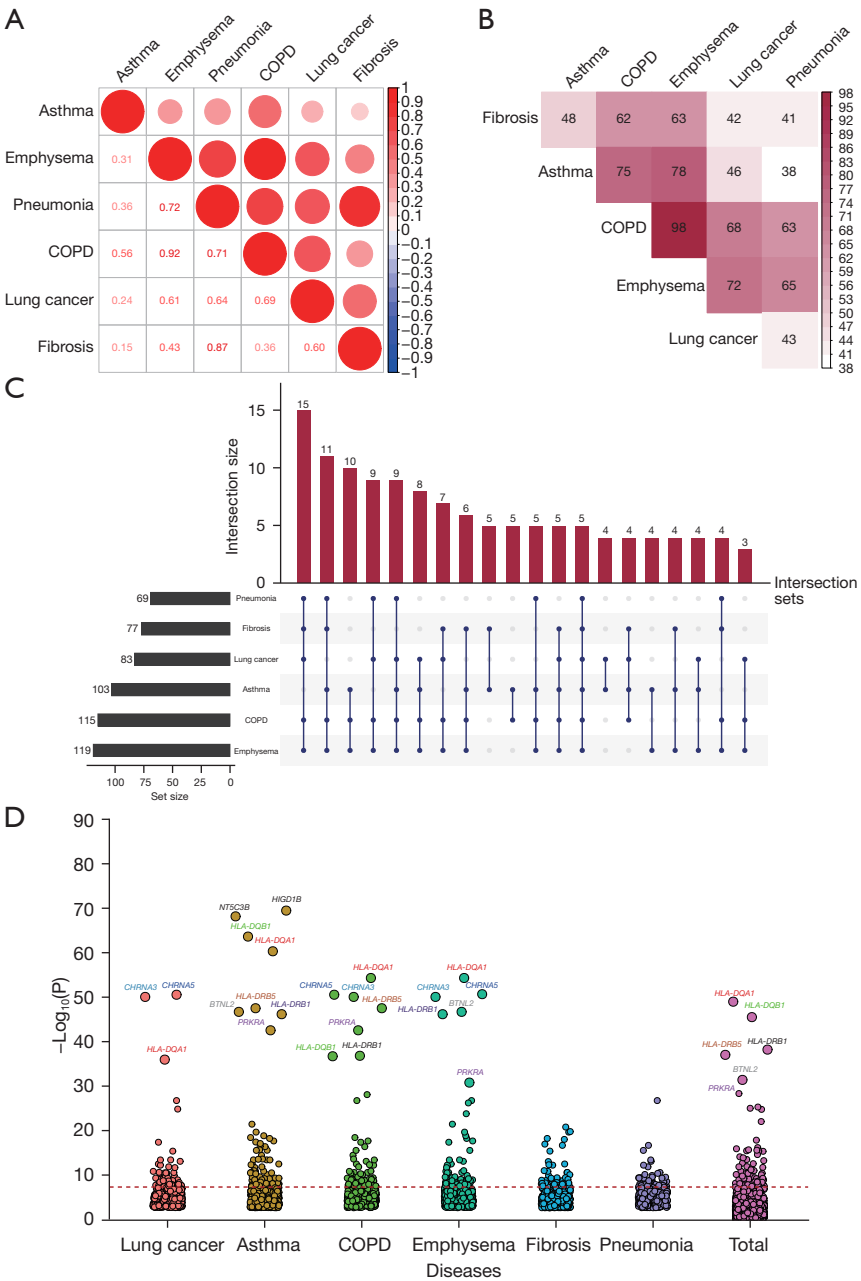

Figure 4 Cross-trait meta-analysis. (A) Estimated genetic correlation of the six lung cancer-related respiratory diseases with the LDSC method. The color and size of circle on the top triangle indicates the magnitude of the genetic correlation; the coefficient of genetic correlation is shown on the bottom triangle. (B) Number of overlap pleiotropic variants discovered by ASSET for the six lung cancer-related respiratory diseases. (C) UpSet plot to illustrate the numbers (N>5) and distribution of pleiotropic variants shared across Lung cancer-related respiratory diseases and the number of pleiotropic variants in each lung cancer-related respiratory diseases. (D) Point plot shows the independent pleiotropic genes associated with any two or more of these six diseases. Identical genes are labeled with the same color. COPD, chronic obstructive pulmonary disease; LDSC, linkage disequilibrium score regression; ASSET, association analysis based on subsets.

Table 2 The pleiotropic genes that overlap with lung cancer with the largest OR

Marker ID	MAF	Gene	Annotation	P	OR (95% CI)	Subset
10:132909243:C>T	1.2×10^{-5}	*CFAP46*	Missense	3.80×10^{-8}	245.01 (39.31, 1,527.15)	LC, COPD, EM, PN, AS
1:38017675:C>T	1.8×10^{-5}	*UTP11*	Missense	1.03×10^{-8}	240.05 (36.84, 1,564.14)	LC, COPD, EM, PN, AS
14:94055016:T>C	1.3×10^{-5}	*DDX24*	Missense	4.64×10^{-9}	174.38 (33.01, 921.28)	LC, COPD, EM, PN, AS
19:32383003:G>A	2.6×10^{-5}	*ZNF507*	Missense	8.74×10^{-10}	155.66 (32.04, 756.23)	LC, COPD, EM, FI, PN
1:53264370:G>A	2.1×10^{-5}	*LRP8*	Missense	1.71×10^{-10}	64.05 (15.98, 256.72)	LC, COPD, EM, FI, PN, AS
19:42079623:C>T	8.2×10^{-4}	*ZNF574*	Synonymous	1.96×10^{-8}	11.47 (4.73, 27.80)	LC, COPD, FI, PN
1:16208697:G>A	1.2×10^{-4}	*ARHGEF19*	LOF	4.05×10^{-9}	11.42 (5.44, 24.00)	LC, COPD, EM, FI, PN
1:43313932:G>A	1.6×10^{-4}	*TIE1*	Synonymous	3.87×10^{-8}	5.00 (2.74, 9.11)	LC, EM, PN, AS
1:171783869:C>T	5.9×10^{-4}	*METTL13*	Missense	1.18×10^{-9}	2.26 (1.70, 3.02)	LC, EM, PN, AS

OR, odds ratio; MAF, minor allele frequency; CI, confidence interval; LC, lung cancer; COPD, chronic obstructive pulmonary disease; EM, emphysema; PN, pneumonia; AS, asthma; FI, fibrosis; LOF, loss-of-function.

in *ZNF574* (MAF =8.2×10^{-4}), and LOF variant 1:16208697:G>A in *ARHGEF19* (MAF =1.2×10^{-4}) were all associated with lung cancer. Moreover, these genes were also strongly associated with other respiratory diseases.

Intermediate causal pathways

Based on the identified pleiotropic variants, we screened for shared genetic variants for the respiratory diseases and lung cancer, and the shared variants and their weights are provided in table available at https://cdn.amegroups.cn/static/public/tlcr-24-4-2.xlsx. Then, the PGS was constructed for these five respiratory diseases. The AUCs (95% CI) of PGS_asthma (PGS_AS), PGS_COPD, PGS_emphysema (PGS_EM), PGS_fibrosis (PGS_FI), and PGS_pneumonia (PGS_PN) are shown in Table S5. Because only shared variants with lung cancer were included in the PGS models, the AUCs performed moderately, but they were all statistically significant.

Applying causal mediation analysis to these polygenic risk scores, we identified the mediating effect of variants causing other respiratory diseases and consequently leading to lung cancer. The direct effect (DE), indirect effect (IE), proportion of mediation, and corresponding significant P value are all shown in *Figure 5*. The mediating effect of COPD was 20%, the mediating effect of emphysema was 23%, and the mediating effect of pneumonia was 20%. This further suggested the existence of some shared variants by causing the development of these three respiratory diseases, which in turn allowed patients to eventually develop lung cancer. However, the mediating effect of asthma and fibrosis was relatively low.

Pathway enrichment analysis and protein-protein interaction network for the pleiotropic genes

We performed gene set enrichment analyses for the 157 unique pleiotropic genes based cross-trait meta-analysis and gene-based association tests using Metascape. We discovered 146 significant pathways [false discovery rate (FDR)-q <0.05] (table available at https://cdn.amegroups.cn/static/public/tlcr-24-4-3.xlsx). The top significant pathways were immune-related, such as phosphorylation of CD3 and TCR zeta chains (P=1.35×10^{-13}), which was associated with T cell receptors (34,35). For the KEGG terms, Th17 cell differentiation (P=1.62×10^{-6}) and T helper (Th)1 and Th2 cell differentiation (P=2.88×10^{-8}) were significant. The biological process of these pleiotropic genes was prominently enriched in positive regulation of immune response (P=5.01×10^{-8}) and regulation of immune effector process (P=3.98×10^{-7}) (Figure S3A).

Furthermore, additional functional pathways were discovered, including regulation of leukocyte proliferation (P=8.13×10^{-6}), response to the bacterium (P=3.09×10^{-5}), regulation of cell-cell adhesion mediated by cadherin (P=1.70×10^{-4}). Meanwhile, the KEGG and Wiki enrichment analysis identified these genes enriched in some respiratory-related pathways, such as asthma (P=2.12×10^{-10}), staphylococcus aureus infection (pulmonary infection, P=7.26×10^{-7}), tuberculosis (P=4.90×10^{-6}), pathogenesis

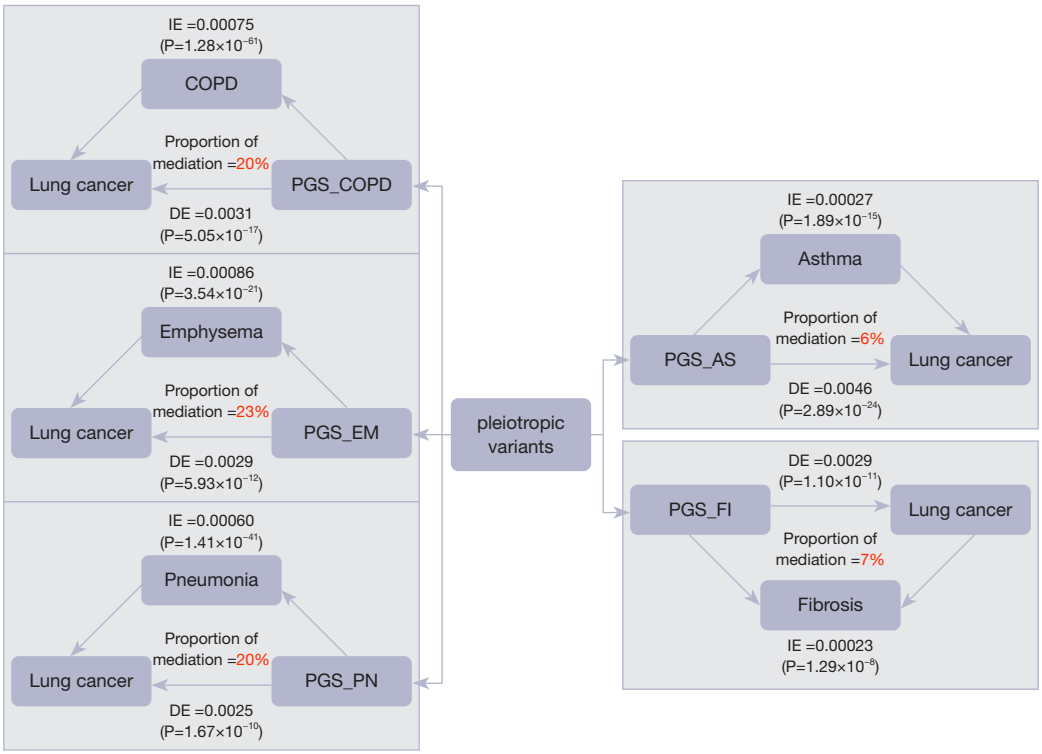

Figure 5 Causal mediation analysis plot shows these pleiotropic variants lead to lung cancer is mediated by affecting the chronic respiratory diseases. The DE, IE, proportion of mediation and corresponding significant P value were all shown in the figure. IE, indirect effect; COPD, chronic obstructive pulmonary disease; PGS, polygenic score; DE, direct effect; EM, emphysema; PN, pneumonia; AS, asthma; FI, fibrosis.

of SARS-CoV-2 mediated by nsp9-nsp10 complex ($P=1.71\times10^{-4}$) and lung fibrosis ($P=3.30\times10^{-4}$).

Meanwhile, we used the STRING database to explore all known and predicted protein-protein interactions among these 157 protein-coding genes (Figure S3B). We identified four large clusters: first was related to the HLA and immune function; second was related to cell adhesion and leukocyte proliferation (including asthma); third was related to pulmonary diseases (lung cancer and COPD) and telomerase; the last was related to inflammatory response. Overall, these results further supported the effect of the identified pleiotropic genes on the respiratory system.

Discussion

In the present work, we comprehensively evaluated the shared genetics of human exome on lung cancer-related respiratory diseases in approximately 420,000 UKB participants of European ancestry, which could improve the statistical power and compensate for the neglect of rare variants in GWASs. To our knowledge, this is the first exome-wide association study for respiratory diseases including almost the whole UKB population. We systematically examined nine common respiratory diseases and identified five that were significantly associated with lung cancer. Based on the single-variant and gene-based association tests, we carried out cross-trait meta-analysis and pathway enrichment analysis, which provided crucial insights into genetic background underlying these diseases and revealed their shared genetic factors with lung cancer. It is worthy to note that there is a small overlap of cases among between these six diseases in which while overlapping subjects can inflate the test statistics of association signals (36,37). Therefore, we adopted ASSET to perform cross-trait meta-analysis to reduce the bias by overlapping subjects (38). Moreover, based on the signal genes and pleiotropic genes, we analyzed protein-protein interaction and identified key modules. Pathway enrichment analysis confirmed that these genes were associated with immune system function and cancer development.

Our first major discovery was that exome-wide signals were associated with the lung cancer-related respiratory diseases. In addition to some known genes (e.g., *CHRNA3*, *CHRNA5*) (39,40), we identified novel genes that have not been reported. *HSD3B7*, which had previously found to be been linked to immune and bile acid function (41,42), was found to be associated with lung cancer. The 3-beta-HSD enzymatic system plays a crucial role in the biosynthesis of all classes of hormonal steroids and HSD VII is active against four 7-alpha-hydroxylated sterols. *SRSF2* and *JAK2* are highly associated with the occurrence of pneumonia, *SRSF2* is necessary for the splicing of pre-messenger RNA (pre-mRNA), and is required for formation of the earliest ATP-dependent splicing complex and interacts with spliceosomal components bound to both the 5'- and 3'-splice sites during spliceosome assembly. While *SRSF2* was found to contribute to myelodysplasia in previous study (43), *JAK2* regulates non-receptor tyrosine kinase involved in various processes such as cell growth, development, differentiation or histone modifications, and mediates essential signaling events in both innate and adaptive immunity.

Our second major contribution was the exploration of the pleiotropic variants shared among lung cancer-related diseases. In single-variant association tests, we did not identify lots of shared variants between respiratory diseases and lung cancer. Nevertheless, we identified 83 shared variants between lung cancer and other five respiratory diseases through cross-trait meta-analysis. Among these shared variants, there were several with incredibly large OR values. This was attributed to the inverse relationship between MAF and OR, where smaller MAFs yield larger ORs. If these variants meet significance thresholds, it suggests their potential significant roles in the occurrence and progression of respiratory diseases, warranting focused investigation. We observed strong functional evidence for the identified genes from the KEGG pathway, GO pathway, Wiki pathway, and protein-protein interaction network. These genes significantly enriched in immune, inflammation, and cell adhesion pathway, which were closely associated with the development of diseases. From protein-protein interaction network, we found these pleiotropic genes could be grouped into four clusters with distinct biological function. The most well-known is HLA family locating on chromosome 6, which is considered as the most polymorphic regions of the human genome (44). Various mutations of HLA are deeply related with immune evasion events and progression of diseases including multiple cancers (45,46). Other clusters included cell adhesion and leukocyte proliferation, pulmonary diseases and telomerase and inflammatory response. These biological processes are closely related to the development of respiratory diseases and even lung cancer, further confirming the important role of these pleiotropic genes in lung cancer and its related respiratory diseases.

Our third major contribution was the exploration of the relation of genetic variants and intermediate causal pathways. We assume that some pleiotropic variants might contribute to the development of lung cancer by causing other respiratory diseases first. Many respiratory diseases have been commonly considered to be risk factors for lung cancer (3,47,48). It is reasonable to assume that there may be some variants that first cause certain respiratory diseases that lead to the development of lung cancer. We calculated the mediating effects of COPD, emphysema, pneumonia, asthma, and fibrosis. The mediating effect was significant for all five lung cancer-related diseases, and the proportions of the mediating effect for COPD, emphysema, and pneumonia all exceeded 20%, which further proved the relationship between these shared pleiotropic variants and the occurrence of lung cancer. Understanding the impact of these common respiratory diseases on lung cancer at the level of shared pleiotropic variants can help us to better assess the risk of lung cancer and to provide effective early warning and prevention of lung cancer.

Our work has several prominent features. First, we comprehensively evaluated the exome-wide genetic variants in six respiratory diseases among 420,000 participants and discovered many coding variants that are difficult to find in traditional GWAS studies. We analyzed the genetic pleiotropy centered on lung cancer and identified the potential shared genes through cross-trait meta-analysis. Second, we identified the proportion of mediation effects of these pleiotropic variants by causing other respiratory diseases to develop, which in turn cause lung cancer. Third, we explored the relationship between identified genes and diseases by exhaustive enrichment pathway analysis and protein-protein interaction analysis.

It is essential to acknowledge the limitations of our study. First, this study was conducted with the UKB population only. The results need further replication in external independent cohorts with large-scale sequencing profiles. Second, we focused on individuals of European ancestry only, and the number of incident lung cancer cases in the UKB is low (n≈4,000) and provides insufficient power to assess the effects of rare variants. It is necessary to include individuals from non-European ancestries in genetic analyses, which is crucial for healthcare equity and genetic

discovery (49). Third, the candidate genes were reported based on statistical evidence and further basic medical experimental studies are still needed to confirm.

Conclusions

Our study provides novel insights into human exomes and rare variants through comprehensive analyses of genetic susceptibility to lung cancer-related diseases and subsequent exploration of shared pleiotropic genes and potential causal pathways.

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Footnote

Peer Review File: Available at https://tlcr.amegroups.com/article/view/10.21037/tlcr-24-4/prf

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at https://tlcr.amegroups.com/article/view/10.21037/tlcr-24-4/coif). The authors have no conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved. The study was conducted in accordance with the Declaration of Helsinki (as revised in 2013).

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Supplementary

Table S1 Lung cancer-related diseases definition and extraction in the UKB

Respiratory diseases	ICD-10 (data fields: 41270)
Asthma	J45
COPD	J44
Emphysema	J43
Fibrosis	J84.1
Pneumonia	J18
Bronchiectasis	J47
Acute bronchitis	J20, J21, J22
Chronic bronchitis	J40, J41, J42, Data-Field 22129
Tuberculosis	A15

UKB, UK Biobank; ICD-10, International Classification of Diseases version 10; COPD, chronic obstructive pulmonary disease.

Table S2 Study population included in the study

Ethnic background	Total number of UKB participants	Number of WES participants included
White British	442,510	401,277
White Irish	13,201	11,916
Any other white background	16,327	14,741
Total	472,038	427,934

UKB, UK Biobank; WES, whole-exome sequencing.

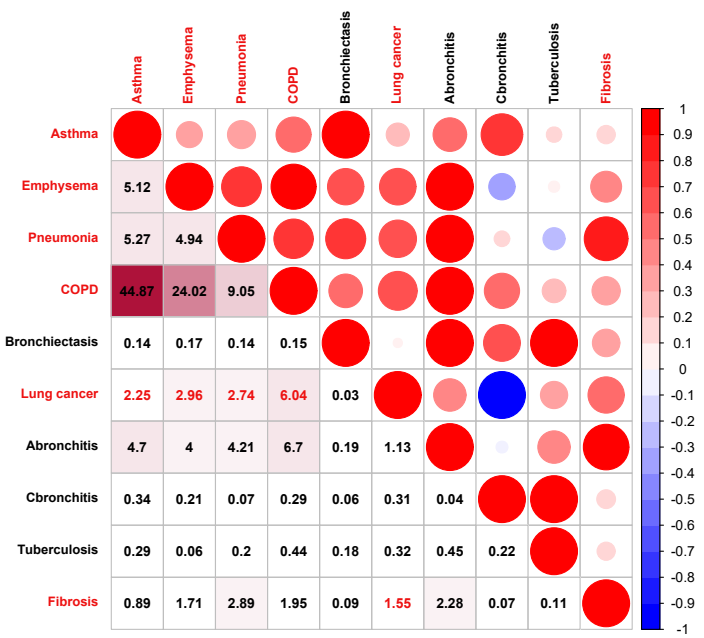

Figure S1 Estimated genetic correlation of ten common respiratory diseases with the LDSC method. The significance of genetic correlation ($-\log_{10}P$) is shown on the bottom triangle and the top triangle indicates the magnitude of the genetic correlation. Diseases related to lung cancer are marked in red. COPD, chronic obstructive pulmonary disease; LDSC, linkage disequilibrium score regression.

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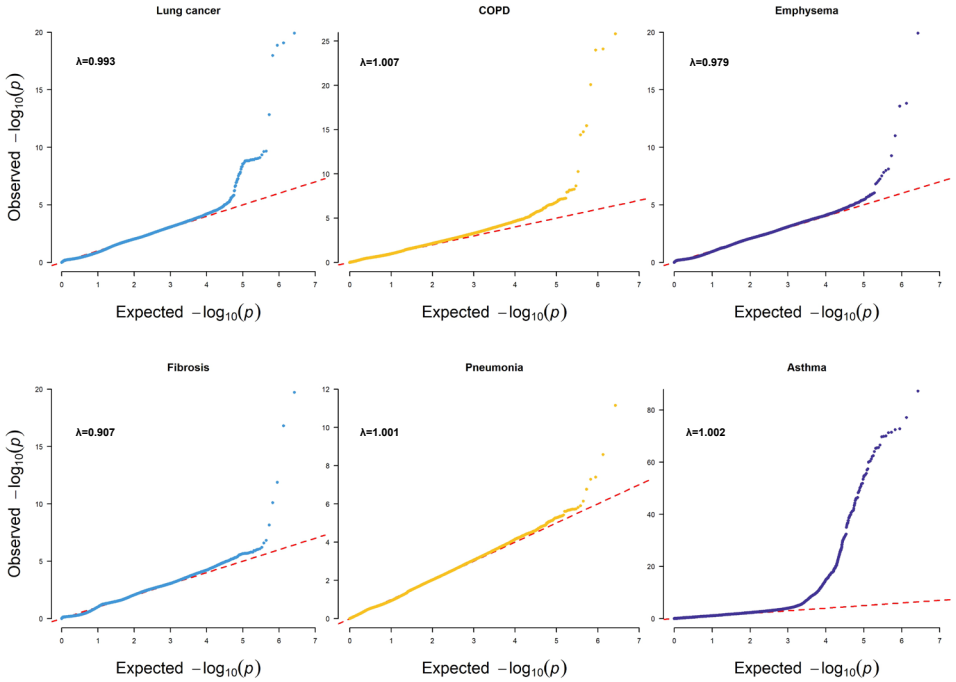

Figure S2 Quantile-quantile plots of association P values for the six lung cancer-related diseases, genomic inflation factor λ are indicated in the figures. COPD, chronic obstructive pulmonary disease.

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Table S3 Association results for independent single variants with $P<5\times10^{-8}$ in the whole UKB-450k population

MarkerID	CHR	Gene	Type	MAC	MAF	BETA	SE	P	Diseases
6:32519337:T>G	6	*HLA-DRB5*	Intron	591,148	0.693	0.05	0.01	1.85E−17	Asthma
6:32519337:T>G	6	*HLA-DRB5*	Intron	591,148	0.693	0.06	0.01	5.61E−09	COPD
15:78596058:G>A	15	*CHRNA3*	Intron	239,508	0.281	−0.08	0.01	5.69E−11	COPD
15:78596058:G>A	15	*CHRNA3*	Intron	239,508	0.281	−0.16	0.03	7.55E−09	Emphysema
15:78596058:G>A	15	*CHRNA3*	Intron	239,508	0.281	−0.13	0.02	1.99E−08	Lung cancer
15:78618839:T>C	15	*CHRNA3*	Missense	334,561	0.392	0.12	0.01	1.49E−26	COPD
15:78618839:T>C	15	*CHRNA3*	Missense	334,561	0.392	0.19	0.02	1.22E−20	Lung cancer
15:78618839:T>C	15	*CHRNA3*	Missense	334,561	0.392	0.17	0.03	1.01E−11	Emphysema
15:78590583:G>A	15	*CHRNA5*	Missense	281,566	0.330	0.20	0.02	8.41E−20	Lung cancer
15:78590583:G>A	15	*CHRNA5*	Missense	281,566	0.330	0.12	0.01	7.97E−25	COPD
1:152305146:G>T	1	*FLG*	Missense	3,282	0.004	0.41	0.08	4.59E−08	Asthma
1:152312600:CACTG>C	1	*FLG*	LOF	20,403	0.024	0.24	0.03	7.04E−16	Asthma
1:152313385:G>A	1	*FLG*	Synonymous	20,451	0.024	0.23	0.03	8.12E−15	Asthma
2:102340888:C>T	2	*IL1RL1*	Intron	117,370	0.138	−0.10	0.01	3.98E−11	Asthma
2:168834219:T>C	2	*NOSTRIN*	Intron	49,448	0.058	0.22	0.02	3.08E−41	Asthma
2:178444419:TA>T	2	*PRKRA*	LOF	11,915	0.014	0.31	0.04	8.17E−15	Asthma
2:241742614:T>C	2	*D2HGDH*	Intron	215,750	0.253	−0.07	0.01	4.82E−09	Asthma
2:97158166:T>G	2	*ANKRD36*	Missense	4,179	0.005	0.41	0.07	2.08E−09	Asthma
4:122456327:C>A	3	*IL2*	Synonymous	281,389	0.330	0.05	0.01	2.33E−10	Asthma
3:10048054:A>G	3	*FANCD2*	Intron	331,784	0.389	−0.08	0.01	1.43E−09	Asthma
4:122207746:T>A	4	*KIAA1109*	Intron	60,319	0.071	0.10	0.01	4.36E−11	Asthma
4:15985886:T>TTTAAG	4	*PROM1*	Intron	764,978	0.897	−0.11	0.01	3.47E−31	Asthma
5:111073450:C>G	5	*TSLP*	Intron	52,130	0.061	−0.11	0.02	2.79E−12	Asthma
5:1280362:G>A	5	*TERT*	Intron	108,034	0.127	0.18	0.03	4.86E−09	Lung cancer
5:1325688:A>G	5	*CLPTM1L*	Intron	374,186	0.439	−0.12	0.02	1.21E−09	Lung cancer
5:132660272:A>G	5	*IL13*	Missense	698,387	0.819	−0.08	0.01	5.64E−18	Asthma
5:14610200:C>G	5	*OTULINL*	Missense	65,800	0.077	0.08	0.01	8.09E−09	Asthma
6:30108978:G>A	6	*TRIM31*	Intron	355,740	0.417	0.04	0.01	2.70E−08	Asthma
6:30720650:G>T	6	*TUBB*	Intron	184,859	0.217	0.05	0.01	8.71E−09	Asthma
6:31355235:G>A	6	*HLA-B*	Intron	201,440	0.236	0.05	0.01	2.36E−09	Asthma
6:31529929:C>T	6	*MCCD1*	Missense	634,093	0.744	0.07	0.01	1.21E−14	Asthma
6:31557961:TG>T	6	*NFKBIL1*	Intron	17,343	0.020	0.15	0.03	4.10E−08	Asthma
6:31795067:G>T	6	*VARS1*	Synonymous	357,625	0.420	0.05	0.01	3.54E−10	Asthma
6:31896897:T>C	6	*EHMT2*	Intron	51,055	0.060	0.13	0.02	7.42E−17	Asthma
6:32058330:C>T	6	*TNXB*	Missense	602,416	0.707	0.08	0.01	7.69E−20	Asthma
6:32402869:A>G	6	*BTNL2*	Intron	55,488	0.065	−0.09	0.02	2.26E−09	Asthma
6:32519576:T>C	6	*HLA-DRB5*	Missense	319,195	0.374	−0.06	0.01	9.30E−16	Asthma
6:32522199:C>T	6	*HLA-DRB5*	Intron	71,181	0.084	0.12	0.01	5.49E−28	Asthma
6:32530151:G>C	6	*HLA-DRB5*	Missense	38,291	0.045	−0.11	0.02	4.17E−09	Asthma

Table S3 *(continued)*

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Table S3 *(continued)*

MarkerID	CHR	Gene	Type	MAC	MAF	BETA	SE	P	Diseases
6:32642197:T>G	6	*HLA-DQA1*	Missense	86,184	0.101	0.07	0.01	2.66E−11	Asthma
6:32642282:T>G	6	*HLA-DQA1*	Intron	434,622	0.510	0.12	0.01	1.81E−73	Asthma
6:32642841:G>GGA	6	*HLA-DQA1*	Intron	60,745	0.071	0.07	0.01	7.73E−09	Asthma
6:32660268:C>G	6	*HLA-DQB1*	Intron	203,106	0.238	−0.05	0.01	2.03E−16	Asthma
6:32661417:G>A	6	*HLA-DQB1*	Missense	49,493	0.058	0.11	0.02	2.57E−11	Asthma
6:32661482:T>C	6	*HLA-DQB1*	Intron	14,797	0.017	0.15	0.02	4.40E−09	Asthma
6:32664780:A>C	6	*HLA-DQB1*	Intron	16,044	0.019	0.13	0.02	1.42E−08	Asthma
6:32664824:G>T	6	*HLA-DQB1*	Missense	53,136	0.062	0.07	0.01	2.62E−10	Asthma
6:32665000:C>T	6	*HLA-DQB1*	Missense	101,067	0.119	−0.10	0.01	6.98E−27	Asthma
6:32665043:C>G	6	*HLA-DQB1*	Missense	190,623	0.224	0.09	0.01	2.69E−32	Asthma
6:32759189:C>T	6	*HLA-DQB2*	Intron	185,349	0.217	0.07	0.01	6.60E−11	Asthma
6:32828569:T>C	6	*TAP2*	Intron	210,056	0.246	−0.08	0.01	1.15E−18	Asthma
6:32830032:C>T	6	*TAP2*	Synonymous	85,105	0.100	0.11	0.01	1.70E−19	Asthma
6:32837693:C>G	6	*TAP2*	Intron	362,951	0.426	0.06	0.01	3.34E−13	Asthma
6:32850253:T>C	6	*TAP1*	Intron	21,290	0.025	0.15	0.02	1.05E−09	Asthma
8:28058313:T>C	8	*NUGGC*	Intron	673,465	0.790	−0.04	0.01	4.65E−09	Asthma
9:6255967:G>C	9	*IL33*	LOF	3,933	0.005	−0.43	0.06	8.58E−14	Asthma
9:5073770:G>T	9	*JAK2*	Missense	297	0.000	1.83	0.27	7.14E−12	Pneumonia
8:86556416:T>G	8	*CNGB3*	Intron	69,714	0.082	0.20	0.01	7.48E−78	Asthma
10:7292138:G>A	10	*SFMBT2*	Intron	87,377	0.103	0.18	0.01	6.53E−88	Asthma
11:1167980:C>T	11	*MUC5AC*	Synonymous	34,324	0.040	0.79	0.09	1.98E−20	Fibrosis
11:61781553:G>A	11	*MYRF*	Intron	294,003	0.345	−0.05	0.01	4.28E−09	Asthma
11:73969150:C>T	11	*DNAJB13*	Intron	4,084	0.005	0.34	0.05	2.59E−10	Asthma
11:75727692:G>A	11	*MOGAT2*	Intron	18,052	0.021	0.15	0.03	6.37E−09	Asthma
12:57099944:T>G	12	*STAT6*	Intron	91,006	0.107	−0.10	0.01	6.65E−15	Asthma
14:33367284:AAAG>A	14	*NPAS3*	Intron	2,794	0.003	0.41	0.07	1.27E−09	Asthma
14:94378610:C>T	14	*SERPINA1*	Missense	16,875	0.020	0.83	0.09	1.20E−20	Emphysema
15:67165147:G>C	15	*SMAD3*	Intron	565,282	0.663	0.05	0.01	2.18E−11	Asthma
15:78596440:AT>A	15	*CHRNA5*	Intron	241,793	0.284	−0.13	0.02	1.95E−08	Lung cancer
16:10969079:C>T	16	*CLEC16A*	Intron	172,756	0.203	−0.05	0.01	2.19E−08	Asthma
16:27345038:C>T	16	*IL4R*	Intron	305,939	0.359	0.06	0.01	5.20E−14	Asthma
17:39905964:C>T	17	*GSDMB*	Synonymous	432,680	0.508	−0.07	0.01	2.25E−20	Asthma
17:39990010:GA>G	17	*PSMD3*	Intron	330,442	0.388	0.04	0.01	2.49E−08	Asthma
17:44848191:C>T	17	*HIGD1B*	LOF	61,511	0.072	0.21	0.01	5.22E−72	Asthma
17:76736877:G>A	17	*SRSF2*	Missense	41	0.000	4.40	0.74	2.69E−09	Pneumonia
19:8914908:T>G	19	*MUC16*	Intron	20,753	0.024	0.18	0.02	5.39E−13	Asthma
22:41387655:C>T	22	*TEF*	Missense	195,978	0.230	0.06	0.01	4.76E−11	Asthma

UKB, UK Biobank; CHR, chromosome; MAC, minor allele count; MAF, minor allele frequency; BETA, effect size of allele; SE, standard error of BETA; COPD, chronic obstructive pulmonary disease; LOF, loss-of-function.

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Table S4 Association results of gene-based tests with $P<1\times10^{-5}$

Gene	Genetic model	Max_MAF	BETA	SE	Number_rare	Number_ultra_rare	P	Diseases
SRSF2	Missense; LOF	0.0001	–0.002	0.001	6	52	7.36E–15	Pneumonia
TNXB	Missense; LOF; synonymous	0.01	0.026	0.006	804	2221	2.42E–07	Asthma
TERT	Missense; LOF	0.001	0.010	0.002	56	375	3.85E–07	Fibrosis
HSD3B7	Missense; LOF; synonymous	0.001	0.002	0.001	72	221	7.73E–07	Lung cancer
C6orf10	Missense; LOF; synonymous	0.01	0.010	0.003	48	190	9.40E–07	Asthma
LMNA	Missense; LOF	0.001	0.005	0.001	61	198	1.06E–06	Fibrosis
JAK2	Missense; LOF	0.001	–0.008	0.002	74	455	2.47E–06	Pneumonia
MOSPD3	Missense; LOF; synonymous	0.0001	0.012	0.006	35	137	2.72E–06	Emphysema
LACRT	Missense; LOF; synonymous	0.0001	0.006	0.002	20	75	6.81E–06	COPD
NOTCH4	Missense; LOF; synonymous	0.01	–0.002	0.000	253	999	8.20E–06	Asthma
HLA-DQA2	Missense; LOF; synonymous	0.01	–0.002	0.001	39	95	8.28E–06	Asthma
TTK	Missense; LOF	0.01	–0.005	0.002	62	257	8.95E–06	Asthma
TARM1	Missense; LOF	0.01	0.003	0.002	37	98	9.16E–06	Lung cancer
SPINK7	Missense; LOF; synonymous	0.0001	0.001	0.001	16	52	9.42E–06	Asthma
ZNF274	Missense; LOF; synonymous	0.01	0.000	0.000	77	311	1.01E–05	Emphysema
C10orf71	Missense; LOF	0.0001	0.001	0.001	146	543	1.03E–05	COPD
ZNF330	Missense; LOF	0.01	0.006	0.002	22	106	1.04E–05	Asthma
COL5A2	Missense; LOF	0.01	0.008	0.005	115	487	1.09E–05	Emphysema
DTNA	Missense; LOF; synonymous	0.001	0.012	0.003	111	348	1.14E–05	Fibrosis
SLC39A5	Missense; LOF	0.0001	–0.011	0.003	53	201	1.31E–05	Fibrosis
DPM2	Missense; LOF; synonymous	0.0001	0.002	0.002	15	88	1.45E–05	Emphysema
PHB	Missense; LOF; synonymous	0.0001	0.001	0.001	20	116	1.53E–05	Emphysema
VKORC1	Missense; LOF; synonymous	0.001	0.010	0.003	46	216	1.71E–05	Pneumonia
TSN	Missense; LOF; synonymous	0.0001	0.016	0.004	11	76	1.78E–05	Asthma
HEPACAM2	Missense; LOF; synonymous	0.01	0.003	0.001	53	228	1.83E–05	Lung cancer
LYL1	Missense; LOF; synonymous	0.0001	0.005	0.001	35	178	1.88E–05	Fibrosis
PSMB6	Missense; LOF; synonymous	0.01	0.011	0.003	21	135	2.00E–05	COPD
CPNE1	Missense; LOF	0.01	0.002	0.001	65	198	2.05E–05	Asthma
ELF2	Missense; LOF; synonymous	0.01	0.012	0.005	55	227	2.11E–05	Asthma
CATSPER3	Missense; LOF	0.01	0.021	0.005	39	149	2.19E–05	Pneumonia
CD79B	Missense; LOF	0.0001	–0.005	0.001	16	82	2.29E–05	Lung cancer
D2HGDH	Missense; LOF; synonymous	0.0001	0.011	0.004	101	315	2.59E–05	Pneumonia
SLFN5	Missense; LOF	0.01	0.011	0.003	96	277	2.67E–05	Asthma
LCE2C	Missense; LOF; synonymous	0.01	0.048	0.012	32	75	2.80E–05	Emphysema
OR2T8	Missense; LOF	0.0001	0.032	0.007	28	78	2.87E–05	Lung cancer
ESPL1	Missense; LOF	0.01	0.014	0.003	127	576	3.15E–05	Emphysema
DNAJC17	Missense; LOF; synonymous	0.0001	0.006	0.002	34	146	3.31E–05	Fibrosis
THNSL1	Missense; LOF; synonymous	0.0001	0.045	0.013	70	338	3.33E–05	Asthma
OR6S1	Missense; LOF; synonymous	0.0001	0.045	0.010	55	170	3.42E–05	Lung cancer

Table S4 *(continued)*

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Table S4 *(continued)*

Gene	Genetic model	Max_MAF	BETA	SE	Number_rare	Number_ultra_rare	P	Diseases
RECK	Missense; LOF	0.01	0.010	0.003	71	291	3.80E–05	Emphysema
TMEM241	Missense; LOF; synonymous	0.001	0.021	0.007	31	121	4.00E–05	Lung cancer
CD37	Missense; LOF; synonymous	0.001	0.030	0.009	43	153	4.12E–05	Asthma
OR2M4	Missense; LOF	0.0001	0.022	0.006	27	103	4.15E–05	Pneumonia
IGFN1	Missense; LOF; synonymous	0.0001	0.034	0.008	535	1906	4.15E–05	Pneumonia
WDR19	Missense; LOF	0.01	0.024	0.007	116	391	4.18E–05	Asthma
GET4	Missense; LOF; synonymous	0.001	0.025	0.005	100	259	4.20E–05	Emphysema
HLA-DRB1	Missense; LOF; synonymous	0.01	0.037	0.013	91	53	4.26E–05	Asthma
KLHL11	Missense; LOF	0.01	0.013	0.007	26	207	4.27E–05	Emphysema
GPR45	Missense; LOF	0.001	0.057	0.014	28	119	4.32E–05	COPD
TEX29	Missense; LOF; synonymous	0.0001	0.011	0.004	26	84	4.57E–05	Asthma
OR2T4	Missense; LOF	0.01	0.019	0.004	43	109	4.87E–05	Pneumonia
OR13F1	Missense; LOF	0.001	0.030	0.007	32	105	4.95E–05	COPD
C6orf15	Missense; LOF; synonymous	0.01	0.016	0.004	64	161	4.95E–05	Asthma
DTWD2	Missense; LOF; synonymous	0.001	0.035	0.009	51	216	4.97E–05	Pneumonia
SGMS1	Missense; LOF; synonymous	0.001	0.017	0.005	38	160	5.15E–05	Asthma
PLG	Missense; LOF	0.01	0.047	0.011	84	254	5.15E–05	COPD
ZNF233	Missense; LOF; synonymous	0.01	0.020	0.005	68	254	5.69E–05	Emphysema
FAM163B	Missense; LOF	0.001	0.059	0.014	34	72	5.69E–05	Lung cancer
CD38	Missense; LOF	0.001	0.034	0.010	33	95	6.38E–05	Asthma
NDST3	Missense; LOF	0.001	0.013	0.004	43	237	6.38E–05	Asthma
GPX2	Missense; LOF; synonymous	0.01	0.021	0.007	41	101	6.65E–05	Lung cancer
TPP1	Missense; LOF	0.01	0.045	0.013	63	184	6.68E–05	Asthma
UIMC1	Missense; LOF; synonymous	0.001	0.022	0.006	85	325	6.70E–05	Asthma
S100A13	Missense; LOF	0.0001	0.016	0.004	10	31	6.88E–05	Pneumonia
ELAVL1	Missense; LOF; synonymous	0.01	0.010	0.004	43	116	7.23E–05	Emphysema
PTCHD3	Missense; LOF; synonymous	0.0001	0.035	0.010	103	449	7.28E–05	Pneumonia
SPHK2	Missense; LOF	0.01	0.034	0.009	69	327	7.31E–05	Lung cancer
PTCHD3	Missense; LOF; synonymous	0.0001	0.039	0.010	103	449	7.40E–05	Lung cancer
ARL8A	Missense; LOF; synonymous	0.001	0.052	0.012	14	67	7.42E–05	Pneumonia
PLCB3	Missense; LOF	0.01	0.021	0.010	100	436	7.53E–05	Asthma
RETN	Missense; LOF; synonymous	0.0001	0.029	0.006	17	65	7.83E–05	Lung cancer
YPEL4	Missense; LOF	0.0001	0.052	0.012	5	39	7.85E–05	COPD
ATP2A3	Missense; LOF	0.01	0.025	0.009	115	417	8.18E–05	Asthma
TEDDM1	Missense; LOF	0.0001	–0.004	0.001	18	93	8.20E–05	Lung cancer
CDH8	Missense; LOF	0.01	0.022	0.006	54	285	8.28E–05	Pneumonia
RAD17	Missense; LOF	0.0001	0.020	0.006	49	202	8.43E–05	Emphysema
FAIM2	Missense; LOF	0.01	0.006	0.001	26	104	8.45E–05	Asthma
KCNK17	Missense; LOF	0.0001	0.013	0.003	37	160	8.46E–05	COPD

Table S4 *(continued)*

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Table S4 *(continued)*

Gene	Genetic model	Max_MAF	BETA	SE	Number_rare	Number_ultra_rare	P	Diseases
CCNY	Missense; LOF; synonymous	0.001	0.006	0.003	42	176	8.50E–05	Emphysema
ELL	Missense; LOF	0.01	0.007	0.002	74	238	8.71E–05	COPD
SZT2	Missense; LOF; synonymous	0.01	–0.008	0.002	453	1765	8.79E–05	Asthma
LY6G6C	Missense; LOF	0.01	0.007	0.003	15	39	8.94E–05	Asthma
ANXA8	Missense; LOF	0.0001	0.009	0.003	22	52	8.95E–05	Asthma
ARHGDIG	Missense; LOF	0.001	0.014	0.004	33	93	9.11E–05	Fibrosis
RCAN2	Missense; LOF; synonymous	0.01	0.008	0.002	38	149	9.19E–05	Pneumonia
POT1	Missense; LOF	0.001	0.052	0.008	33	203	9.45E–05	Lung cancer
FBXO27	Missense; LOF	0.001	0.013	0.004	29	99	9.60E–05	Pneumonia

Max_MAF, maximum MAF cutoff; MAF, minor allele frequency; BETA, effect size of allele; SE, standard error of BETA; number_rare, number of markers that are not ultra-rare with MAC >10; number_ultra_rare: number of markers that are ultra-rare with MAC ≤10; MAC, minor allele count; LOF, loss-of-function; COPD, chronic obstructive pulmonary disease.

Table S5 AUC of PGSs used for mediation analyses

PGS based on shared variants	AUC	95% CI	P
PGS_AS	0.536	0.533–0.539	4.70E–207
PGS_COPD	0.543	0.539–0.547	1.46E–278
PGS_EM	0.562	0.553–0.571	2.76E–318
PGS_FI	0.544	0.533–0.556	2.89E–130
PGS_PN	0.524	0.520–0.528	2.83E–149

AUC, area under the receiver operator characteristic curve; PGS, polygenic score; CI, confidence interval; AS, asthma; COPD, chronic obstructive pulmonary disease; EM, emphysema; FI, fibrosis; PN, pneumonia.

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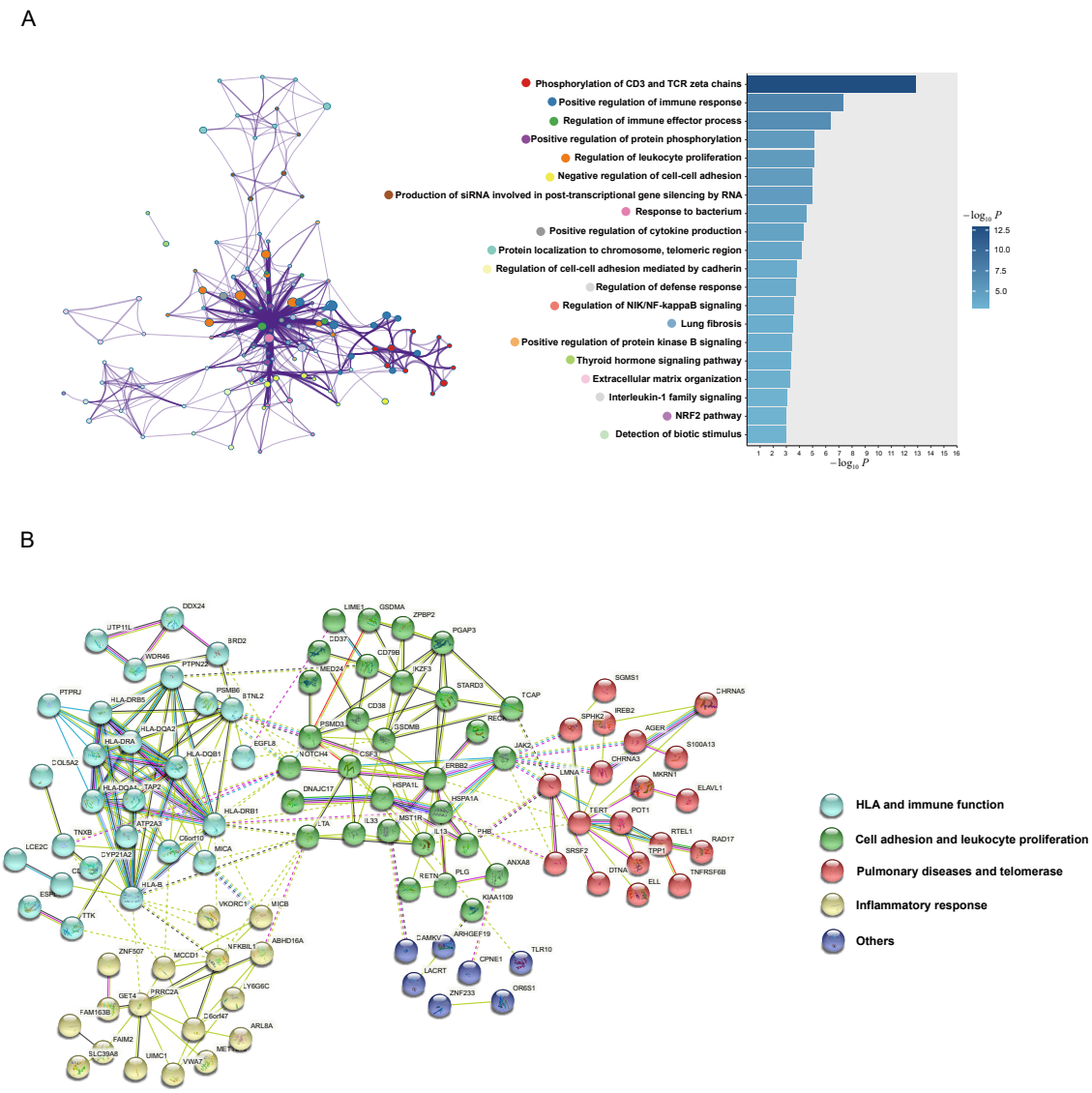

Figure S3 Genetic functional analysis. (A) Left of figure shows the clustering process of the significant enrichment pathway, and those with the same color are grouped into the same class. The most statistically significant term within a cluster is chosen to exhibit in nearby bar plot. (B) Protein-protein interaction network of the signal genes and pleiotropic genes. HLA, human leukocyte antigen.

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