

A review of radiation-related malignancy in the pelvis

Lauren C. Linkowski¹, Brandon J. Manley², Peter A. S. Johnstone³, G. Daniel Grass³

¹University of South Florida Morsani College of Medicine, Tampa, FL, USA; ²Department of Genitourinary Oncology, H. Lee Moffitt Cancer Center and Research Institute, Tampa, FL, USA; ³Department of Radiation Oncology, H. Lee Moffitt Cancer Center and Research Institute, Tampa, FL, USA *Contributions:* (I) Conception and design: LC Linkowski, PAS Johnstone, GD Grass; (II) Administrative support: None; (III) Provision of study materials or patients: None; (IV) Collection and assembly of data: LC Linkowski, GD Grass; (V) Data analysis and interpretation: LC Linkowski, GD Grass; (VI) Manuscript writing: All authors; (VII) Final approval of manuscript: All authors.

Correspondence to: G. Daniel Grass, MD, PhD. Department of Radiation Oncology, H. Lee Moffitt Cancer Center and Research Institute, Tampa, FL 33612, USA. Email: daniel.grass@moffitt.org.

Abstract: Radiation therapy is a central treatment modality for the management of various pelvic malignancies and prior data has supported a relationship between radiation exposure and the development of long-term treatment sequelae. One of the most consequential long-term side effects of radiation therapy is the risk of developing a secondary malignancy. With advancements in radiotherapy delivery and a better appreciation of underlying tumor biology, additional considerations are needed when assessing the risk of radiation-mediated malignancies. Also, several adjacent normal structures within the pelvis may be affected by radiation-mediated toxicity, driven in part by acute and chronic inflammation. Depending on treatment modality and primary tumor location, various steps can be taken in radiation planning to reduce the risk of these side effects, which may negatively affect the patient's quality of life. As cancer survivorship continues to increase, it is important to understand both the treatment and biologic variables which influence the risk of developing secondary malignancies in order to minimize the risk for treatment side effects and the late effect of secondary malignancy. Herein, we will provide an overview of secondary malignancies in the context of receiving therapeutic radiation to the pelvis and will highlight biologic considerations that may influence this risk.

Keywords: Secondary malignancy; radiotherapy; pelvic malignancy

Received: 19 October 2020; Accepted: 10 May 2021; Published: 25 March 2022. doi: 10.21037/amj-20-179 View this article at: http://dx.doi.org/10.21037/amj-20-179

Introduction

Rationale/background

An appreciable increase in cancer survivorship has been observed over the last several decades due to advancements in oncologic care. Concomitant with improved survival times, the probability of adverse treatment sequelae occurrence also increases. The development of a secondary malignant neoplasm (SMN) is one of the most consequential late effects following cancer treatment.

SMN is responsible for approximately 19% of the cancers diagnosed in the United States, which is secondary to the aging population and the increase in cancer survivors (1). Radiation therapy contributes to approximately 5% of these SMNs (2). Radiation therapy is the most commonly employed treatment modality in oncology, and given the larger percentage of cancer survivorship, it is important to consider the risk for SMN following radiotherapy. Prior data has established that radiation has carcinogenic potential (3), which has been underscored by the development of cancers in the survivors of the atomic bombs, Chernobyl accident and associated occupational exposures (4,5). In this context, radiation is mostly associated with the development of leukemia, thyroid, and breast cancers (6).

In contrast to the exposure of the spread of radioisotopes released into the atmosphere, targeted ionizing radiation

[^] ORCID: 0000-0002-8417-5611.

directed at patient tissue represents an additional mechanism of radiation-induced malignancies. Clinical data on radiation-induced carcinogenesis in patients without prior malignancy or immunologic defects are rare. Prior data in patients treated for dermatologic, infectious or rheumatologic conditions with radiation have suggested increased risks of SMN in anatomic regions where the radiotherapy was directed (7-10). Data extrapolated from animal studies also support a risk for SMN development following radiation therapy. Combined data in over 100 canines exposed to intraoperative electron beam radiation have shown a significant latency period is required and that non-rapidly dividing tissues require higher cumulative radiation doses to induce a SMN (11,12). Edmondson et al. found 26% of mice developed a radiationinduced SMN, which was dependent on fractionation of radiation, total dose and genetic background (13). These tenets support the clinical rationale to minimize radiation exposure in younger patients, given the longer period of time to develop a SMN, and highlight the minimal consequence of SMN in older patients with anticipated poor survival outcomes.

One of the challenges in surveillance data, is attributing the development of SMNs in the context of other confounding variables, such as various environmental factors, like smoking, which have been associated with malignancy risk (6). Further, surveillance bias could confound results as patients are routinely asked about urinary and bowel symptoms following radiation to the pelvis, therefore warranting a deeper investigation as symptoms arise (14). Thus, in the setting of competing factors influencing risk of carcinogenesis, it is important to define criteria for radiation-induced SMN. In the 1940s, Cahan et al. first proposed criteria to define radiationinduced sarcomas (15). Later, Laskin et al. performed a pathologic assessment of radiation-induced sarcomas and noted most were poorly differentiated and suggested 3 criteria must be met to specify a radiation-induced SMN: (I) tumor must be in an anatomic site exposed to prior radiation, (II) tumor must be a different histology than the treated lesion, and (III) an appropriate duration of time must have elapsed between radiotherapy and SMN formation (16).

Of note, radiation-induced SMNs are usually described as an excess in risk over the estimated risk in the general population for a given tumor. Since radiation-mediated SMN is a relatively rare event, useful analyses must be performed in large study cohorts to identify statistically meaningful differences. For this reason, most studies evaluating radiation-induced SMN is confined to retrospective analyses on epidemiologic data. In order to best understand the impact of the studies included, it is important to consider the difference between absolute and relative risk (RR). Many of the included cohorts have data reported by RR, comparing two groups, one of which was treated by radiation. This is in contrast to the absolute risk, which relays risk over a period of time. Simply reporting RR conceals the absolute risk, which many times is more valuable to an individual patient (17). Risk assessment becomes more complicated when incorporating varying demographics, treatment approaches and polygenic traits, thus most models describe radiation-induced risk in the setting of RR. This distinction is paramount when counseling patients on radiation-induced SMN risk.

Objectives

In this review we provide an overview of the current literature related to radiation-mediated SMN risk in the context of age at exposure, temporality, solid versus hematologic malignancies, chemotherapy, radiation type, and genetic predisposition. Unless otherwise mentioned, radiation therapy refers to external beam radiation therapy.

Methods

Our search was conducted via PubMed with keywords that included "second primary cancer", "subsequent neoplasm", "radiation-induced malignancy", "radiation malignancy", and "childhood cancer survivorship". Additional papers were found via the references from articles related to the original search.

Discussion

Radiation-mediated normal tissue injury

Although the exact mechanisms are unknown, radiationinduced normal tissue injury is a major contributor to treatment-related toxicity. Frequently, radiation oncologists are balancing this therapeutic ratio every day in clinic. The dogma of radiation-mediated cellular injury emphasizes damage to genetic material, which may be induced by double-stranded DNA breaks leading to compromised chromosomal integrity (18). By generation of free radicals and reactive oxygen species, radiation produces an inflammatory response via various cytokine cascades. This inflammation poses a threat to adjacent normal tissue (19). The consequences of these effects are largely based on tissue diversity and their turnover (20). It is hypothesized that the response to injury and interaction between multiple cell types within an organ creates an ongoing process even after the treatment has been delivered (21).

Further detrimental synergy can occur when systemic therapies, which also disrupt cell cycle dynamics and induce further DNA damage, are added to radiation therapy. As both radiation and chemotherapy have been associated with SMN risk, this may create a "perfect storm" in the tissue microenvironment for subsequent SMN formation. These late combinatorial effects have been studied extensively in cardiac tissue with radiation therapy in combination with anthracycline-based chemotherapy regimens (22-24).

Within the pelvis, non-tumor tissues may undergo changes related to the mechanisms described above. The clinical consequences of normal tissue injury are partially dependent on whether the insulted tissue is organized in physiologic serial or parallel units. For example, nerves are considered serial structures as damage to any component can affect both upstream and downstream function. In contrast, a muscle group in the pelvis or a portion of the bladder has parallel units that may compensate if one portion of the tissue is damaged (25). This is important for consideration when assessing the consequences of radiationmediated tissue effects.

The cell turnover characteristics of the tissue also contribute to the radiation response. We encourage further reading on this topic in the recent review by McBride et al. (20). For example, enteritis is acutely driven by endothelium injury and the cytotoxic effects on the crypt stem cells (26), whereas chronic enteritis is characterized by a more prolonged fibrotic process driven by fibrotic signaling pathways and telangiectasia formation (27). Similarly, cystitis also develops from a radiation-induced inflammatory process (28). Cystitis may be complicated by hemorrhage and clinical management is challenging (29). Analysis of peptides in the urine have demonstrated marker expression representing sequential fibrosis and vascular damage resulting in an inflammatory milieu in the bladder (30). Interestingly, acrolein, the byproduct of common chemotherapeutics (e.g., cyclophosphamide), induces robust inflammatory reactions when in contact with the bladder mucosa. This insult is perturbed with the coadministration of antioxidants (31). suggesting that the end result of both radiation or chemotherapeutic tissue damage culminates from uncontrolled inflammation.

The radiation-induced side effects, which are partially attributed to increased reactive oxygen species, may be dampened with the addition of amifostine. Amifostine is a thiol-containing molecule that scavenges free radicals, has additional protective pleiotropic effects, and appears to have selectivity for normal tissue (32). At present, amifostine is FDA approved to prevent xerostomia resulting from head and neck irradiation and in perturbing platinum chemotherapy-induced nephrotoxicity (33-35). Amifostine has also been shown in a randomized trial to reduce acute grade 2-3 toxicities in patients receiving pelvic radiotherapy for a variety of malignancies (36), which may be further improved with amifostine dose escalation (37). Though amifostine appears to decrease radiation-induced side effects to some degree, enthusiasm in the clinic has been tempered by patient intolerance, necessity for dosing right before radiation delivery and risk of hypotension. Whether amifostine decreases the risk of SMN is unknown.

Potential biologic mechanisms associated with SMN

The biologic mechanisms contributing to the development of SMN development are not known. Deleterious cellular effects of radiation have been attributed to induction of chromosomal aberrations and DNA damage in the context of diverse cell autonomous DNA repair capabilities and sustained non-lethal cell division events (38). SMN formation may also be influenced by tissue bystander effects, which are characterized by phenomena of genomic disruption and activation of signaling pathways in cells not directly exposed to radiation (39). Accumulating evidence also suggests non-irradiated cells can reduce the damage to irradiated cell populations through various soluble mediators (40). How these diverse processes influence radiation-induced SMN is a growing area of research.

Age at exposure

The risk of developing a SMN related to age at first radiation exposure has been well documented, which highlights those at younger ages have increased time to develop a SMN. Data from atomic bomb survivors has shown cancer risk is approximately 15% in those under 10 years of age at exposure, which decreases to 1% in those over 60 years old (41). Some studies have cited sensitivity to radiation as a primary mechanism when compared to adult populations (41). Broadly, cancer survivors treated with

Page 4 of 14

radiation at an age above 18 years have a long-term risk of SMN across all body sites of 1.1 to 3 times higher than the general population (42), whereas risk in children are 5-10 times greater (43).

The Childhood Cancer Survivor Study (CCSS) followed 14,359 patients diagnosed between 1970 and 1986 under 21 years old and survived at least 5 years after initial diagnosis (44). Within this cohort, 10.5% received radiation only and about half received chemotherapy and radiation (45). At 30 years of follow-up, 20.5% (95% CI: 19.1-21.8%) developed a SMN, with those treated with radiation having a RR of 2.9 (95% CI: 2.1-4.2) (46). The average time to developing a SMN was 19±6.7 years with the average age being 29.5±9.1 years (45). Meningioma was the most common SMN with a RR of 16.6 (95% CI: 5.2-52.6) (45). When specific risk factors were analyzed, female survivors were at a greater risk for developing secondary malignancy than their male counterparts (RR: 1.46) (47). Breast cancer risk was generally increased in survivors who were treated with chest radiotherapy, but prior radiation to the pelvis was protective with a RR of 0.6 (95% CI: 0.4-0.9) (48). The CCSS cohort is limited in its data of site of radiation-induced SMNs; though, in children other groups have reported an increased risk of thyroid, central nervous system (CNS), and breast cancers (49). This data indicates the most common primary site related to radiation-mediated SMN varies by age, with most data relating to chest radiation and not to the pelvis.

Separate from therapy-related SMN risk, exposure from diagnostic radiology has been attributed with primary malignancy risk. In a study of individuals between 0-19 years and a median follow-up of 9.5 years, the cancer incidence was 24% higher in exposed vs. unexposed individuals; caveats to this data are the imaging eras (e.g., 1980s through 2000s) and how modern low-dose CT approaches in pediatrics now contribute (50). A study in Taiwan observed about a 3-fold increase in benign brain tumor, but not malignant tumor, identification in pediatrics exposed to CT scans of the head vs. those without exposure; this study was limited to lack of data on radiation dose, contribution of other diagnostic radiology exposures and baseline genetic predisposition (51). A similar study from the Netherlands also identified a small excess risk of malignant brain tumors following CT head imaging in the pediatric population. SMN of concern include leukemia, lymphoma, and CNS cancers (PMID, 26882064). Most studies have focused on CT head imaging, though it is possible that similar assertions can be applied to the pelvis

as well. Though there is evidence for a small increased risk of tumor development in pediatrics exposed to radiation from diagnostic radiology, when clinically indicated, the benefit of CT imaging must outweigh the risk of concern for malignancy risk. In doing so, imaging in pediatrics with "as low as reasonably achievable" (ALARA) is a prudent approach to mitigate this risk.

Overall, though there appears to be an increased risk in children, detailed statistics on risk of SMNs related to radiation is limited because malignancy in children is relatively rare, and direct comparisons of SMN risk in anatomically similar tumors between pediatric and adult patients are not robust enough to make definitive conclusions.

Surveillance time to SMN

Surveillance time following cancer therapy varies based on parameters of the individual study and age at treatment. Two national cohorts have been paramount in the analysis of cancer related outcomes, the CCSS and the Surveillance, Epidemiology, and End Results (SEER) Program facilitated by the National Cancer Institute (52). For example, a SEER analysis of children who survived treatment of genitourinary malignancies found the peak risk of developing a SMN was 5 to 9 years after primary diagnosis and treatment (42).

Surveillance data centered in radiation-related malignancies is an additional consideration when choosing primary treatment strategy, especially when the primary diagnosis is at a young age. For example, in the PORTEC-1 trial at a median follow-up of 15 years, 22% and 16% of patients developed SMNs in radiation versus observation groups, respectively. Similarly, long-term surveillance of a cervical cancer cohort (n=107,706) treated with radiation identified an elevated risk of SMN, standardized incidence ratio (SIR) =1.3 (95% CI: 1.28-1.33) at a follow-up time of 40 years (53). Also, the risk of all SMN was higher in those treated with radiation therapy than those who were not, SIR =1.34 (95% CI: 1.31-1.38) and 1.06 (95% CI: 1.02-1.11), respectively (53). In another SEER analysis, the cumulative incidence of developing any SMN 25 years after radiation was 17.5% (95% CI: 17.1-17.9%) in uterine, 15.4% (95% CI: 14.0-16.8%) in vulvar, 13.2% (95% CI: 12.5-13.8%) in cervical, and 9.4% (95% CI: 8.9-9.9%) in ovarian cancers (42). As surveillance time lengthens, the chance for developing SMN increases in most cases, however, it is important to consider the confounding risks that are given additional time to develop as well.

AME Medical Journal, 2022

Primary tumor site treated with radiotherapy	No. of patients	Site of secondary malignant neoplasm	Site of secondary malignant neoplasm	Patient follow-up time (years)	Reference #
Prostate	122,123	RR: 15% & 34%	Bladder, rectum, colon	5 & 10	(54)
	397,416	RR: 1.30 (95% CI, 1.19–1.42)	Bladder	5	(55)
	242,878	RR: 1.94 (95% CI, 1.07–3.50)	Rectum	5	(55)
	34,889	RR: 1.5 (95% Cl, 1.1–2.0)	Bladder	5	(56)
	192,658	SHR: 1.89 (95% CI: 1.66-2.16)		Median: 6 (0–24)	(57)
Bladder	192,658	SHR: 0.67 (95% CI 0.47-0.94)		Median: 6 (0–24)	(57)
Rectal	1,599	RR: 1.85 (95% CI 1.23–2.78)	Adjacent to rectum	20	(58)
	2,554	No SMN risk		Median: 13 (1.8–21.2)	(59)
Gynecologic	568	RR: 2.02 (95% CI, 1.30–3.15)	Not specified	20.5	(60)
	60,949	RR: 1.26 (95% CI, 1.16–1.36)	Solid tumor malignancies	30	(61)
	199,268	HR: 1.72 (95% Cl, 1.37–2.15)	Leukemia	5–15	(62)
	192,658	RR: 1.5 (95% Cl, 1.13–2.00)	Rectum	Median: 6 (0-24)	(57)
Cervical	104,760	RR: 1.30 (95% CI, 1.28–1.33)	Colon, rectum/anus, urinary bladder, ovary, and genital sites	40	(53)
	182,040	RR: 1.1	Bladder, rectum, uterine corpus, ovary, small intestine, bone, and connective tissue	>1	(63)

HR, hazard ratio; RR, relative risk; SHR, sub-hazard ratio; SMN, secondary malignant neoplasm.

Specific pelvic tumor types

It is difficult to accurately define the risk of SMN induced by radiation because most studies do not have an appropriate non-irradiated control group. Some tumors in the pelvis are effectively managed by surgery or radiation, therefore this provides opportunity to have a case and control group for comparisons. Prostate, bladder, rectal, and gynecological (e.g., endometrial, cervical) carcinomas will be discussed with their significance related to development of SMN (*Table 1*).

Prostate cancer

Localized prostate cancer is treated definitively with radical prostatectomy or radiation therapy, which are curative in most instances (64). Several studies have demonstrated prostate cancer patients treated with radiation have an increased risk of developing SMNs, particularly in organs within close proximity to the treated prostate, such as the bladder, rectum, and colon (14,54,55,65,66). Incidence data from the SEER cancer registry compared 51,584 men with prostate cancer who were treated with radiation or surgery without radiation. In this study, radiation therapy was associated with a 6% increase in risk of solid SMN, with this reaching 15% and 34% at 5 and 10 years of follow-up, respectively (54). Similarly, a study in 39,028 men found that at a median follow-up of 5 years, 7.4% of men developed a SMN, which persisted after adjustment of other treatment and clinical confounders (67). An earlier cohort (n=34,889) analysis found a RR of 1.3 (95% CI: 1.0-1.7) for developing bladder cancer at 5 years after radiation compared to those not treated with radiotherapy; interestingly this increased risk peaked at 8 years (RR =1.5; 95% CI: 1.1-2.0) (56). Rombouts et al. also found a significantly increased risk (HR =1.89, 95% CI: 1.66-2.16) for developing a secondary rectal cancer in men treated with radiation for prostate cancer (57), yet no significant risk

was reported for hematologic cancers (54,55). With longer surveillance, the odds of developing bladder or rectal cancer after radiotherapy *vs.* surgery were 1.3 *vs.* 1.9 at 5 years and 1.7 *vs.* 2.2 at 10 years, respectively (55). In contrast, Curtis *et al.* found patients treated with prostate radiation had an increased risk of developing bladder cancer, but no increase in rectal cancer (42). Similarly, Kendal *et al.* also observed no increase in rectal cancers in men treated with prostate radiation after adjusting for confounders (68).

Chrouser et al. found an excess of bladder cancer cases in men treated with prostate bed radiation only (69), whereas a study in the British Columbia Tumor Registry found no excess of risk in irradiated patients over a nonirradiated cohort (66). In a single institution report, Liauw et al. found patients treated with brachytherapy alone had a lower incidence of bladder and colorectal SMNs compared to brachytherapy with supplemental radiation (70). Also, Wang et al. performed a SEER analysis on men treated with radiotherapy, brachytherapy, or prostatectomy and found that both radiotherapy and brachytherapy were associated with an increased risk of SMN compared to surgery, though the absolute increase was 1% after 10 years (71). Also, patients treated with prostatectomy were half as likely to have post-treatment bladder cancer compared to patients who underwent radiation therapy (HR =2.08, 95% CI: 0.29-3.9) (72). Keehn et al. found all radiation modalities increased the risk of bladder cancer, but prostate brachytherapy was associated with the largest increase after 10 years (73). To complicate these observations further, prior data suggests the rate of bladder cancer in patients with prostate cancer is 18 times higher than expected, even in the absence of radiation treatment (74), and that simultaneous diagnosis of both cancers occurs between 0.4-6.6% of the time (75). Overall, there should be some caution in concluding prostate or prostate bed radiation definitively elevates rectal and bladder cancer incidence, as the risk is low, there are competing risks and the data to infer risk are incomplete and conflicting.

Rectal cancer

Treating rectal cancer with radiation therapy carries a risk for structural changes to the bowel wall. Patients may suffer from rectal wall toxicity secondary to an inflammation (i.e., proctitis) and fibrosis sequence, which can manifest as tenesmus, changes in bowel habits and rectal bleeding. Most patients have self-limiting effects during treatment, which resolve with conservative therapy and surveillance. Dose-volume relationships have been evaluated in several settings of pelvic radiation, which includes conventional and hypofractionated approaches (76). From these investigations, "generalized" dose constraints from the population have been employed in radiation planning to minimize rectal toxicity risk. With modern advancements in conformal and inverse-planning radiation delivery, proctitis is significantly lower (77). Chronic proctitis is seen in about 5% of patients and can be complicated by fistula formation, bowel obstruction and permanent changes to bowel habits (78). A systematic review of 21 studies evaluating rectal and anal function following prostate radiation found that radiation may reduce anal resting pressure, decrease compliance of the rectum and induce telangiectasias formation (79). Whether this initial inflammatory cascade induced by radiation predisposes surrounding normal rectal mucosa to malignancy is unknown. A recent study evaluating non-neoplastic rectal mucosa after preoperative rectal radiation noted histologic findings of dysplasia, but next generation sequencing demonstrated no increased risk of genetic aberrations (80). This should be interpreted in the context of tissue analysis immediately following radiation vs. the risk of genetic changes that may occur over several years.

Primary rectal cancer treatment with radiation has also been associated with increased risk for SMN. According to a SEER-based analysis of 77,436 patients followed with primary cancer at the rectosigmoid junction, 28% received a form of radiation therapy in their treatment. Of the patients who received radiation therapy, 7.7% developed a SMN, with 26% being within the digestive tract and 15% specifically within the colon. In the same study, 63% of anal cancer patients were treated with radiation and 10.3% developed a SMN, with no specification for SMN location after radiation (42). Birgisson et al. also found an elevated risk (RR =2.04, 95% CI: 0.97-3.27) of SMN in patients treated with rectal radiation in the Uppsala and Swedish Rectal Cancer Trial, with most instances being in tissue adjacent to the irradiated volume (58). In contrast, Wiltink et al. found no increase in SMN risk in a pooled analysis of >2,500 trial patients treated with or without radiation for pelvic malignancies; they did find that patients with rectal or endometrial primary cancers had increased probability to develop secondary cancers compared to the general population (59). Kendal and Nicholas also found in a population-based analysis that secondary cancers after irradiation for rectal cancers are infrequent and should not be factored into treatment decisions when assessing risk in an older population of patients (81). A modeling study by Zwahlen et al. found that rectal radiation can increase

SMN risk by 2% in older patients, but may increase the risk up to 10% in those less than 30 years of age (82). Overall, the data regarding radiation-induced rectal cancer is also conflicting, but there may be some increased risk compared to not treatment not employing radiotherapy.

Gynecologic cancers

Additional solid tumor malignancies are particularly common in patients with endometrial cancer and several studies have examined the risk of SMN following radiation treatment for this disease. A SEER analysis of patients treated with radiation for uterine cancer who survived at least one year were found to have increased risk for SMN compared to patients who had received surgery alone. In this study, combined radiation and brachytherapy had the strongest association with new solid malignancies in the cohort of patients who developed SMN (61). Additionally, external bream radiotherapy has been associated with an increased risk for SMN, especially in women treated before the age of 60 (HR =2.02, 95% CI: 1.30-3.15) (60).

Women treated for cervical cancer at a younger age sustain a significantly higher risk for SMN due to radiation (53,62). Consistent risk factors among cervical cancer survivors include exposure to radiation, human papillomavirus, and smoking, which are associated with a 40-year cumulative risk of any SMN of 22% (53). It is unclear whether risk factors in cervical cancer are synergistic as in lung cancer, where radiotherapy and smoking have shown increased risk for developing Hodgkin lymphoma (53). In a cohort study of cervical cancer survivors (n=104,760), radiation was a significant risk factor for developing a SMN at any site, as well as locally at sites receiving greater than 3 Gy (53). Interestingly, a reduction in the risk of breast cancer was observed after pelvic radiation, which could be attributed to age of treatment or a change in hormone exposure to breast tissue after hysterectomy and ovarian ablation (53,83). This is underscored by a study demonstrating that more than 6 Gy to the ovaries reduced breast cancer risk by 44% (84).

Hematologic malignancies

Radiation has been associated with an increased risk for developing various hematologic malignancies, such as acute myelogenous leukemia, chronic myelogenous leukemia, and acute lymphoblastic leukemia (6). In contrast, chronic lymphocytic leukemia has not been associated with radiation (6,63,85). The risk for non-lymphocytic leukemia was significant in the first decade of follow-up (SIR =2.74), whereas risk for chronic lymphocytic leukemia was not significantly increased any time after radiation exposure (53,63). Thus, for those patients who are susceptible to developing leukemia following radiation, the time is usually a few years compared to >10 years for the development of most solid tumors (84,86). Single, large doses of radiation are more associated with development of leukemia, as opposed to small doses over a lifetime (87). Further, at low doses (<1 Gy), the predicted leukemia excess RR increases linearly with dose (86).

It is estimated that more than 40% of active bone marrow mass is contained within the pelvic region, which may contribute to the risk of developing leukemias after pelvic radiation therapy (88). More recent estimates have confirmed that the pelvic bones (17.5%), sacrum (9.9%), L5 vertebra (2.5%) are responsible for a large fraction of active bone marrow in adults (86). Due to the enrichment of calcium, the absorbed dose in bone can be much greater than surrounding soft tissue (89). Radiation has been shown to change the bone microenvironment, which may be characterized by altered osteo-clast-blast balance and replacement of marrow with adipose tissue (90). How these altered bone environments may contribute to SMN risk is unknown. It is possible bone marrow stem cells exposed to radiation may acquire genetic abnormalities or that depletion of certain marrow compartments may lead to decreased immune cell surveillance in the host. Understanding the consequences and mechanisms of bone marrow irradiation and SMN risk is an area of needed investigation.

Depending on the treatment scenario, conformal radiation may be planned to spare dose to the bone marrow in the pelvis. Advanced conformal radiation delivery techniques employing intensity-modulated radiation therapy (IMRT) can reduce the volume of pelvic bone marrow receiving a specified radiation dose. This reduction of bone marrow dose has been shown to reduce acute grade \geq 3 hematologic toxicity in patients receiving pelvic radiation and concurrent chemotherapy by approximately 10% (91). Further refinement of this IMRT approach with ¹⁸F-fluorodeoxyglucose positron emission tomography (PET) delineation of active bone marrow has been shown to be feasible (92,93) and is currently being employed in clinical trial development.

In a large study (n=199,268) of patients treated with pelvic radiation for tumors of the vulva, cervix, uterus, anus, and rectosigmoid junction, just over one-third of

patients who received radiation had a 72% increased RR for developing a secondary leukemia. This risk peaked at 5 to 10 years after treatment (HR =1.82, 95% CI: 1.40-2.44) and remained elevated at 15 years of follow-up (HR =1.5, 95% CI: 1.03–2.18); interestingly, there was no significant risk for multiple myeloma (62). Some studies have also found that cervical cancer patients may have an increased risk for leukemia, and treatment with radiation seems to increase this risk further (53). Similar findings have identified an association in the treatment of testicular cancer, citing a greater total volume of bone marrow exposed to radiation is associated with an 11-fold risk of leukemia (6,94,95). At 5 years of follow-up after prostate radiation, Journy et al. found less than 1% of patients developed a leukemia or myelodysplasia (67) and Wang et al. also found an absolute risk increase of 0.5% for hematologic malignancies after prostate radiation at 10 years of follow-up (71).

Chemotherapy risk

A potential confounding variable to consider when examining risk of radiation-mediated malignancy is the use of cytotoxic chemotherapy, which is frequently delivered in combination or in sequence with radiation. The leukemogenic potential of chemotherapy is well established, where the RR of leukemia after radiation is considerably smaller than after chemotherapy alone to the order of approximately 2-fold (6). In a study of ovarian cancer patients (n=28,791), the risk of SMN was differential based on the class of chemotherapy, where the overall risk of leukemia following platinum-based therapy was 4.0 (95% CI: 1.4-11.4) (96). Patients who had received platinumbased chemotherapy in addition to radiation were found to have significantly higher risk of leukemia when compared to platinum-based chemotherapy alone, suggesting different chemotherapies synergize with radiation to induce leukemias (6,96). Also, analysis of the CCSS cohort found 49% of participants received chemotherapy and radiation, 10.5% received radiation alone, and 21.5% received chemotherapy alone. Of these, 61% developed a subsequent neoplasm when treated with both chemotherapy and radiation, compared to 8% and 20% treated with radiation or chemotherapy alone, respectively (45).

Radiation modality and associated risk

Assessment of SMN risk related to radiation approach requires data on patients treated with different radiation modalities with extended follow-up in order to identify the event; unfortunately, such data is currently limited. A general dose-response relationship leading to development of SMN has been documented in both adult and pediatric populations (97,98). Specifically, these studies demonstrated there may be a threshold of the integral radiation dose that non-tumor tissue can be exposed to before a delayed SMN occurs.

Various approaches are employed to deliver tumoricidal radiation doses to patients with the goal to spare normal tissues as much as possible. Though many efforts are made to spare healthy normal tissue from radiation, the dose interface between tumor and normal tissue is never zero. This is a consequence of scattered radiation within the patient, beam-limiting devices, dose leakage and the production of secondary radiation (99). As technology has advanced, radiation delivery now encompasses twodimensional targeting, three-dimensional conformal radiotherapy (3DCRT), IMRT and particle therapy, such as proton beam therapy (PBT).

Previously, investigators postulated IMRT may increase SMN risk due to greater normal tissue volumes receiving low-dose radiation, which is intrinsic to IMRT dose distribution (100). Though IMRT reduces radiation dose to the bladder, rectum and femoral heads compared to 3DCRT, Tao et al. found IMRT increases the volume of non-tumor tissue in the pelvis receiving 5-30 Gy and raises the mean radiation dose outside the pelvis by approximately 7% (101). Diallo et al. analyzed the dosimetric data of 115 young patients who developed SMNs and found that only 12% of SMNs arose within the high dose irradiated volume. Further, they observed two-thirds of SMNs arose in the region <5 cm from the beam border (102). Though this data is not specific to pelvic malignancies, it does highlight the risk of organs in close proximity to targets like the prostate, bladder, rectum and nodal fields, which may receive an intermediate radiation dose. Modeling studies on dose have suggested carcinoma SMN risk decreases by 10% and sarcoma SMN risk decreases by 15% per 1 Gy increase per fraction; this suggests that shorter courses of radiation with higher doses per fraction may decrease SMN risk (103). Thus, in some instances radiation dose distribution can increase SMN risk, but the manner in which the cumulative dose is delivered may counter this risk.

Treatment with PBT can reduce the total integral radiation dose because protons deposit most of their energy at a specified depth (e.g., Bragg peak) with little exit dose (104). Chung *et al.* found that compared to photon therapy, PBT reduces SMN formation by approximately 2%, which remained significant after adjustment for other clinical factors (105). Schneider *et al.* suggested spot-scanning PBT reduces the risk of SMN formation compared to IMRT and this was supported by a study by Fontenot *et al.* who also found PBT reduces the risk of SMN by 26% to 39% depending on patient body habitus (106). In the context of young children and adolescents, PBT may decrease the risk of SMN and is a major reason why this modality is employed in pediatric malignancies (107).

A recent SEER-Medicare analysis of men treated for prostate cancer found no difference in the overall SMN RR between 3DCRT and IMRT; though, there was preliminary data supporting a reduction in rectal and colon cancer with IMRT, but not bladder cancer (67). Most recently, a National Cancer Database study evaluated SMN risk relative to radiation treatment modality in over 450,000 patients spanning 9 tumor types across various anatomic locations. This study found at a median 5 years of followup, the absolute risk of SMN was 1.55/100 patient-years with no differences observed between 3DCRT and IMRT; though, patients treated with PBT had a lower odds (OR =0.31, 95% CI: 0.26–0.36) of developing a SMN compared to IMRT (108). Thus, this data implies there may be advantages to using PBT to reduce secondary malignancy risk compared to other modalities, though with caveats of limited follow-up, incomplete demographics, and genetic predispositions.

Genetic predisposition and risk

Prior data indicates that beyond 10 years, about 9% of cancer survivors develop a SMN, with an estimated 0.5% excess absolute risk at 15 years attributed to radiation therapy (109). Several physical and treatment parameters have implied children are at increased risk of SMNs compared to adults when exposed to radiation. Another factor that may also contribute to increased SMN risk is the underlying genetics of each patient. Whereas many adults develop cancer in the setting of perpetual genomic insults secondary to lifestyle choices or occupational exposures, many children develop cancers due to germline mutations (99). This begs the question of whether intrinsic genetic susceptibilities influence radiation-related risk of SMN development. With the advent and increasing implementation of genetic profiling in cancer patients, more data in this venue is likely to improve our understanding.

Within a patient, the reaction to radiation appears to be mostly ascribed to patient-specific characteristics (110). The general population is assumed to have a uniform sensitivity to radiation, though several rare syndromes have been identified that increase radiation sensitivity and appear to be associated with single genetic aberrations (111). Several "pathogenic" gene variants have been identified which confer an increased risk of cancer development and possibly toxicity to radiation therapy. For instance, patients with alterations in the BRCA1 or BRCA2 genes have an increased risk of developing contralateral secondary breast primary tumors in the absence of radiation (112) or those with bilateral retinoblastomas driven by RB1 aberrations have increased risk of radiationinduced osteosarcomas (113). Although BRCA carriers have an enhanced risk for malignancy, no increased risk of radiation-induced SMN formation has been identified (114). The lack of an increased SMN incidence after radiation in carriers suggests other variables other than maintenance of DNA fidelity contribute to this process. In contrast, patients with TP53 alterations resulting in Li-Fraumeni syndrome have been estimated to have approximately >20% excess risk for radiation-induced SMN (115). A study including prostate cancer patients identified a single nucleotide polymorphism (SNP) in ATM, which increased the risk for acute toxicity, but whether this is related to SMN is unknown (116). Other SNP studies in prostate cancer have found alterations in TANC1 (117), KDM3B and DNAH5 (118) are associated with toxicity to normal organs in the pelvis following radiation. Again, whether this corresponds to SMN risk is unknown. It is also important to appreciate that the majority of somatic mutations in tumors are likely not present in normal tissues, thus relating certain mutations to normal tissue risk of SMN remains unclear.

Summary

The development of SMNs is a significant late sequelae risk following radiotherapy. In general, cancer survivors who were treated with radiation have increased risk to develop SMNs compared to the general public. This risk is determined by age at exposure, radiation dose, volume of irradiated normal tissue, biologic parameters and adequate follow-up time to identify the event. Importantly, most data supporting increased risk for radiation-induced SMNs is based on retrospective epidemiologic data, dose-response models from homogenous exposures of radioisotopes in the atmosphere, and with little consideration of patient-specific biologic predisposition. As the cancer care continuum

Page 10 of 14

continues to advance and patient survival improves, understanding the complexities of what drives SMN risk is of utmost importance.

Acknowledgments

Funding: None.

Footnote

Provenance and Peer Review: This article was commissioned by the Guest Editor (Lucas Wiegand) for the series "Radiation Urologic Reconstruction" published in AME Medical Journal. The article has undergone external peer review.

Conflicts of Interest: All authors have completed the ICMJE uniform disclosure form (available at https://amj.amegroups.com/article/view/10.21037/amj-20-179/coif). The series "Radiation Urologic Reconstruction" was commissioned by the editorial office without any funding or sponsorship. PASJ reports personal fees from Huron Consulting, outside the submitted work. The authors have no other conflicts of interest to declare.

Ethical Statement: The authors are accountable for all aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

Open Access Statement: This is an Open Access article distributed in accordance with the Creative Commons Attribution-NonCommercial-NoDerivs 4.0 International License (CC BY-NC-ND 4.0), which permits the non-commercial replication and distribution of the article with the strict proviso that no changes or edits are made and the original work is properly cited (including links to both the formal publication through the relevant DOI and the license). See: https://creativecommons.org/licenses/by-nc-nd/4.0/.

References

- Morton LM, Onel K, Curtis RE, et al. The rising incidence of second cancers: patterns of occurrence and identification of risk factors for children and adults. Am Soc Clin Oncol Educ Book 2014:e57-67.
- 2. Dracham CB, Shankar A, Madan R. Radiation induced secondary malignancies: a review article. Radiat Oncol J

2018;36:85-94.

- 3. El Ghissassi F, Baan R, Straif K, et al. A review of human carcinogens--part D: radiation. Lancet Oncol 2009;10:751-2.
- Williams D. Radiation carcinogenesis: lessons from Chernobyl. Oncogene 2008;27 Suppl 2:S9-18.
- 5. Doll R. Hazards of ionising radiation: 100 years of observations on man. Br J Cancer 1995;72:1339-49.
- 6. Travis LB. The epidemiology of second primary cancers. Cancer Epidemiol Biomarkers Prev 2006;15:2020-6.
- Ron E, Modan B, Boice JD Jr. Mortality after radiotherapy for ringworm of the scalp. Am J Epidemiol 1988;127:713-25.
- Ron E, Modan B, Boice JD Jr, et al. Tumors of the brain and nervous system after radiotherapy in childhood. N Engl J Med 1988;319:1033-9.
- 9. Smith PG, Doll R. Mortality among patients with ankylosing spondylitis after a single treatment course with x rays. Br Med J (Clin Res Ed) 1982;284:449-60.
- Albright EC, Allday RW. Thyroid carcinoma after radiation therapy for adolescent acne vulgaris. JAMA 1967;199:280-1.
- Johnstone PA, Laskin WB, DeLuca AM, et al. Tumors in dogs exposed to experimental intraoperative radiotherapy. Int J Radiat Oncol Biol Phys 1996;34:853-7.
- Powers BE, Gillette EL, McChesney SL, et al. Bone necrosis and tumor induction following experimental intraoperative irradiation. Int J Radiat Oncol Biol Phys 1989;17:559-67.
- Edmondson EF, Hunter NR, Weil MM, et al. Tumor Induction in Mice After Localized Single- or Fractionated-Dose Irradiation: Differences in Tumor Histotype and Genetic Susceptibility Based on Dose Scheduling. Int J Radiat Oncol Biol Phys 2015;92:829-36.
- Murray L, Henry A, Hoskin P, et al. Second primary cancers after radiation for prostate cancer: a systematic review of the clinical data and impact of treatment technique. Radiother Oncol 2014;110:213-28.
- Cahan WG, Woodard HQ, Higinbotham NL, et al. Sarcoma arising in irradiated bone: report of eleven cases. 1948. Cancer 1998;82:8-34.
- Laskin WB, Silverman TA, Enzinger FM. Postradiation soft tissue sarcomas. An analysis of 53 cases. Cancer 1988;62:2330-40.
- 17. Noordzij M, van Diepen M, Caskey FC, et al. Relative risk versus absolute risk: one cannot be interpreted without the other. Nephrol Dial Transplant 2017;32:ii13-8.
- Jackson SP, Bartek J. The DNA-damage response in human biology and disease. Nature 2009;461:1071-8.
- 19. Schaue D, Kachikwu EL, McBride WH. Cytokines

AME Medical Journal, 2022

in radiobiological responses: a review. Radiat Res 2012;178:505-23.

- 20. McBride WH, Schaue D. Radiation-induced tissue damage and response. J Pathol 2020;250:647-55.
- Robbins ME, Brunso-Bechtold JK, Peiffer AM, et al. Imaging radiation-induced normal tissue injury. Radiat Res 2012;177:449-66.
- 22. Goethals I, Dierckx R, De Meerleer G, et al. The role of nuclear medicine in the prediction and detection of radiation-associated normal pulmonary and cardiac damage. J Nucl Med 2003;44:1531-9.
- 23. Morgan GW, Freeman AP, McLean RG, et al. Late cardiac, thyroid, and pulmonary sequelae of mantle radiotherapy for Hodgkin's disease. Int J Radiat Oncol Biol Phys 1985;11:1925-31.
- 24. Girinsky T, Cordova A, Rey A, et al. Thallium-201 scintigraphy is not predictive of late cardiac complications in patients with Hodgkin's disease treated with mediastinal radiation. Int J Radiat Oncol Biol Phys 2000;48:1503-6.
- Withers HR, Taylor JM, Maciejewski B. Treatment volume and tissue tolerance. Int J Radiat Oncol Biol Phys 1988;14:751-9.
- 26. Harb AH, Abou Fadel C, Sharara AI. Radiation enteritis. Curr Gastroenterol Rep 2014;16:383.
- 27. Kennedy GD, Heise CP. Radiation colitis and proctitis. Clin Colon Rectal Surg 2007;20:64-72.
- 28. Taidi Z, Mansfield KJ, Bates L, et al. Purinergic P2X7 receptors as therapeutic targets in interstitial cystitis/ bladder pain syndrome; key role of ATP signaling in inflammation. Bladder (San Franc) 2019;6:e38.
- 29. Pascoe C, Duncan C, Lamb BW, et al. Current management of radiation cystitis: a review and practical guide to clinical management. BJU Int 2019;123:585-94.
- Zwaans BMM, Nicolai HE, Chancellor MB, et al. Prostate cancer survivors with symptoms of radiation cystitis have elevated fibrotic and vascular proteins in urine. PLoS One 2020;15:e0241388.
- Batista CK, Mota JM, Souza ML, et al. Amifostine and glutathione prevent ifosfamide- and acrolein-induced hemorrhagic cystitis. Cancer Chemother Pharmacol 2007;59:71-7.
- Obrador E, Salvador R, Villaescusa JI, et al. Radioprotection and Radiomitigation: From the Bench to Clinical Practice. Biomedicines 2020;8:461.
- 33. Antonadou D, Pepelassi M, Synodinou M, et al. Prophylactic use of amifostine to prevent radiochemotherapy-induced mucositis and xerostomia in head-and-neck cancer. Int J Radiat Oncol Biol Phys

2002;52:739-47.

- 34. Brizel DM, Wasserman TH, Henke M, et al. Phase III randomized trial of amifostine as a radioprotector in head and neck cancer. J Clin Oncol 2000;18:3339-45.
- Koukourakis MI. Amifostine in clinical oncology: current use and future applications. Anticancer Drugs 2002;13:181-209.
- 36. Athanassiou H, Antonadou D, Coliarakis N, et al. Protective effect of amifostine during fractionated radiotherapy in patients with pelvic carcinomas: results of a randomized trial. Int J Radiat Oncol Biol Phys 2003;56:1154-60.
- Koukourakis MI, Kyrgias G, Panteliadou M, et al. Dose escalation of amifostine for radioprotection during pelvic accelerated radiotherapy. Am J Clin Oncol 2013;36:338-43.
- Huang RX, Zhou PK. DNA damage response signaling pathways and targets for radiotherapy sensitization in cancer. Signal Transduct Target Ther 2020;5:60.
- 39. Marín A, Martín M, Liñán O, et al. Bystander effects and radiotherapy. Rep Pract Oncol Radiother 2014;20:12-21.
- Lam RK, Fung YK, Han W, et al. Rescue effects: irradiated cells helped by unirradiated bystander cells. Int J Mol Sci 2015;16:2591-609.
- 41. Hall EJ. Intensity-modulated radiation therapy, protons, and the risk of second cancers. Int J Radiat Oncol Biol Phys 2006;65:1-7.
- 42. Curtis RE, Freedman DM, Ron E, et al. editors. New Malignancies Among Cancer Survivors: SEER Cancer Registries, 1973-2000. Bethesda, MD: National Cancer Institute. NIH Publ.; 2006.
- 43. de Vathaire F, Hawkins M, Campbell S, et al. Second malignant neoplasms after a first cancer in childhood: temporal pattern of risk according to type of treatment. Br J Cancer 1999;79:1884-93.
- Robison LL, Mertens AC, Boice JD, et al. Study design and cohort characteristics of the Childhood Cancer Survivor Study: a multi-institutional collaborative project. Med Pediatr Oncol 2002;38:229-39.
- 45. Friedman DL, Whitton J, Leisenring W, et al. Subsequent neoplasms in 5-year survivors of childhood cancer: the Childhood Cancer Survivor Study. J Natl Cancer Inst 2010;102:1083-95.
- 46. Armstrong GT, Liu Q, Yasui Y, et al. Late mortality among 5-year survivors of childhood cancer: a summary from the Childhood Cancer Survivor Study. J Clin Oncol 2009;27:2328-38.
- 47. Meadows AT, Friedman DL, Neglia JP, et al. Second neoplasms in survivors of childhood cancer: findings from

Page 12 of 14

the Childhood Cancer Survivor Study cohort. J Clin Oncol 2009;27:2356-62.

- Kenney LB, Yasui Y, Inskip PD, et al. Breast cancer after childhood cancer: a report from the Childhood Cancer Survivor Study. Ann Intern Med 2004;141:590-7.
- 49. Inskip PD, Curtis RE. New malignancies following childhood cancer in the United States, 1973-2002. Int J Cancer 2007;121:2233-40.
- Mathews JD, Forsythe AV, Brady Z, et al. Cancer risk in 680,000 people exposed to computed tomography scans in childhood or adolescence: data linkage study of 11 million Australians. BMJ 2013;346:f2360.
- Huang WY, Muo CH, Lin CY, et al. Paediatric head CT scan and subsequent risk of malignancy and benign brain tumour: a nation-wide population-based cohort study. Br J Cancer 2014;110:2354-60.
- 52. Duggan MA, Anderson WF, Altekruse S, et al. The Surveillance, Epidemiology, and End Results (SEER) Program and Pathology: Toward Strengthening the Critical Relationship. Am J Surg Pathol 2016;40:e94-102.
- Chaturvedi AK, Engels EA, Gilbert ES, et al. Second cancers among 104,760 survivors of cervical cancer: evaluation of long-term risk. J Natl Cancer Inst 2007;99:1634-43.
- Brenner DJ, Curtis RE, Hall EJ, et al. Second malignancies in prostate carcinoma patients after radiotherapy compared with surgery. Cancer 2000;88:398-406.
- 55. Wallis CJ, Mahar AL, Choo R, et al. Second malignancies after radiotherapy for prostate cancer: systematic review and meta-analysis. BMJ 2016;352:i851.
- Neugut AI, Ahsan H, Robinson E, et al. Bladder carcinoma and other second malignancies after radiotherapy for prostate carcinoma. Cancer 1997;79:1600-4.
- 57. Rombouts AJM, Hugen N, Elferink MAG, et al. Increased risk for second primary rectal cancer after pelvic radiation therapy. Eur J Cancer 2020;124:142-51.
- Birgisson H, Pahlman L, Gunnarsson U, et al. Occurrence of second cancers in patients treated with radiotherapy for rectal cancer. J Clin Oncol 2005;23:6126-31.
- 59. Wiltink LM, Nout RA, Fiocco M, et al. No Increased Risk of Second Cancer After Radiotherapy in Patients Treated for Rectal or Endometrial Cancer in the Randomized TME, PORTEC-1, and PORTEC-2 Trials. J Clin Oncol 2015;33:1640-6.
- 60. Onsrud M, Cvancarova M, Hellebust TP, et al. Long-term outcomes after pelvic radiation for early-stage endometrial cancer. J Clin Oncol 2013;31:3951-6.
- 61. Lönn S, Gilbert ES, Ron E, et al. Comparison of second

cancer risks from brachytherapy and external beam therapy after uterine corpus cancer. Cancer Epidemiol Biomarkers Prev 2010;19:464-74.

- 62. Wright JD, St Clair CM, Deutsch I, et al. Pelvic radiotherapy and the risk of secondary leukemia and multiple myeloma. Cancer 2010;116:2486-92.
- Boice JD Jr, Day NE, Andersen A, et al. Second cancers following radiation treatment for cervical cancer. An international collaboration among cancer registries. J Natl Cancer Inst 1985;74:955-75.
- 64. Teo MY, Rathkopf DE, Kantoff P. Treatment of Advanced Prostate Cancer. Annu Rev Med 2019;70:479-99.
- 65. Bostrom PJ, Soloway MS. Secondary cancer after radiotherapy for prostate cancer: should we be more aware of the risk? Eur Urol 2007;52:973-82.
- Pickles T, Phillips N. The risk of second malignancy in men with prostate cancer treated with or without radiation in British Columbia, 1984-2000. Radiother Oncol 2002;65:145-51.
- Journy NM, Morton LM, Kleinerman RA, et al. Second Primary Cancers After Intensity-Modulated vs 3-Dimensional Conformal Radiation Therapy for Prostate Cancer. JAMA Oncol 2016;2:1368-70.
- Kendal WS, Eapen L, Macrae R, et al. Prostatic irradiation is not associated with any measurable increase in the risk of subsequent rectal cancer. Int J Radiat Oncol Biol Phys 2006;65:661-8.
- 69. Chrouser K, Leibovich B, Bergstralh E, et al. Bladder cancer risk following primary and adjuvant external beam radiation for prostate cancer. J Urol 2005;174:107-10; discussion 110-1.
- Liauw SL, Sylvester JE, Morris CG, et al. Second malignancies after prostate brachytherapy: incidence of bladder and colorectal cancers in patients with 15 years of potential follow-up. Int J Radiat Oncol Biol Phys 2006;66:669-73.
- 71. Wang C, King CR, Kamrava M, et al. Pattern of solid and hematopoietic second malignancy after local therapy for prostate cancer. Radiother Oncol 2017;123:133-8.
- 72. Boorjian S, Cowan JE, Konety BR, et al. Bladder cancer incidence and risk factors in men with prostate cancer: results from Cancer of the Prostate Strategic Urologic Research Endeavor. J Urol 2007;177:883-7; discussion 887-8.
- Keehn A, Ludmir E, Taylor J, et al. Incidence of bladder cancer after radiation for prostate cancer as a function of time and radiation modality. World J Urol 2017;35:713-20.
- 74. Chun TY. Coincidence of bladder and prostate cancer. J

AME Medical Journal, 2022

Urol 1997;157:65-7.

- Soga N, Furusawa J, Ogura Y. Long-Term Management of Incidental Bladder Cancer Detected in Patients Undergoing Prostatectomy for Prostate Cancer. Curr Urol 2019;13:145-9.
- Michalski JM, Gay H, Jackson A, et al. Radiation dosevolume effects in radiation-induced rectal injury. Int J Radiat Oncol Biol Phys 2010;76:S123-9.
- 77. Wee CW, Kang HC, Wu HG, et al. Intensity-modulated radiotherapy versus three-dimensional conformal radiotherapy in rectal cancer treated with neoadjuvant concurrent chemoradiation: a meta-analysis and pooledanalysis of acute toxicity. Jpn J Clin Oncol 2018;48:458-66.
- Porouhan P, Farshchian N, Dayani M. Management of radiation-induced proctitis. J Family Med Prim Care 2019;8:2173-8.
- 79. Krol R, Smeenk RJ, van Lin EN, et al. Systematic review: anal and rectal changes after radiotherapy for prostate cancer. Int J Colorectal Dis 2014;29:273-83.
- Zanelli M, Ciarrocchi A, De Petris G, et al. Acute Radiation Colitis after Preoperative Short-Course Radiotherapy for Rectal Cancer: A Morphological, Immunohistochemical and Genetic Study. Cancers (Basel) 2020;12:2571.
- Kendal WS, Nicholas G. A population-based analysis of second primary cancers after irradiation for rectal cancer. Am J Clin Oncol 2007;30:333-9.
- 82. Zwahlen DR, Bischoff LI, Gruber G, et al. Estimation of second cancer risk after radiotherapy for rectal cancer: comparison of 3D conformal radiotherapy and volumetric modulated arc therapy using different high dose fractionation schemes. Radiat Oncol 2016;11:149.
- Kleinerman RA, Boice JD Jr, Storm HH, et al. Second primary cancer after treatment for cervical cancer. An international cancer registries study. Cancer 1995;76:442-52.
- Boice JD Jr, Engholm G, Kleinerman RA, et al. Radiation dose and second cancer risk in patients treated for cancer of the cervix. Radiat Res 1988;116:3-55.
- 85. Curtis RE, Boice JD Jr, Stovall M, et al. Relationship of leukemia risk to radiation dose following cancer of the uterine corpus. J Natl Cancer Inst 1994;86:1315-24.
- Shuryak I, Sachs RK, Hlatky L, et al. Radiation-induced leukemia at doses relevant to radiation therapy: modeling mechanisms and estimating risks. J Natl Cancer Inst 2006;98:1794-806.
- 87. Finch SC. Radiation-induced leukemia: lessons from history. Best Pract Res Clin Haematol 2007;20:109-18.
- 88. Cristy M. Active bone marrow distribution as a function of

age in humans. Phys Med Biol 1981;26:389-400.

- Curi MM, Cardoso CL, de Lima HG, et al. Histopathologic and Histomorphometric Analysis of Irradiation Injury in Bone and the Surrounding Soft Tissues of the Jaws. J Oral Maxillofac Surg 2016;74:190-9.
- Costa S, Reagan MR. Therapeutic Irradiation: Consequences for Bone and Bone Marrow Adipose Tissue. Front Endocrinol (Lausanne) 2019;10:587.
- 91. Mell LK, Sirak I, Wei L, et al. Bone Marrowsparing Intensity Modulated Radiation Therapy With Concurrent Cisplatin For Stage IB-IVA Cervical Cancer: An International Multicenter Phase II Clinical Trial (INTERTECC-2). Int J Radiat Oncol Biol Phys 2017;97:536-45.
- 92. Franco P, Fiandra C, Arcadipane F, et al. Incorporating (18)FDG-PET-defined pelvic active bone marrow in the automatic treatment planning process of anal cancer patients undergoing chemo-radiation. BMC Cancer 2017;17:710.
- 93. Yusufaly T, Miller A, Medina-Palomo A, et al. A Multiatlas Approach for Active Bone Marrow Sparing Radiation Therapy: Implementation in the NRG-GY006 Trial. Int J Radiat Oncol Biol Phys 2020;108:1240-7.
- 94. Travis LB, Curtis RE, Storm H, et al. Risk of second malignant neoplasms among long-term survivors of testicular cancer. J Natl Cancer Inst 1997;89:1429-39.
- 95. Travis LB, Andersson M, Gospodarowicz M, et al. Treatment-associated leukemia following testicular cancer. J Natl Cancer Inst 2000;92:1165-71.
- Travis LB, Holowaty EJ, Bergfeldt K, et al. Risk of leukemia after platinum-based chemotherapy for ovarian cancer. N Engl J Med 1999;340:351-7.
- 97. Tukenova M, Guibout C, Hawkins M, et al. Radiation therapy and late mortality from second sarcoma, carcinoma, and hematological malignancies after a solid cancer in childhood. Int J Radiat Oncol Biol Phys 2011;80:339-46.
- 98. Nguyen F, Rubino C, Guerin S, et al. Risk of a second malignant neoplasm after cancer in childhood treated with radiotherapy: correlation with the integral dose restricted to the irradiated fields. Int J Radiat Oncol Biol Phys 2008;70:908-15.
- Newhauser WD, Durante M. Assessing the risk of second malignancies after modern radiotherapy. Nat Rev Cancer 2011;11:438-48.
- 100. Ruben JD, Davis S, Evans C, et al. The effect of intensitymodulated radiotherapy on radiation-induced second malignancies. Int J Radiat Oncol Biol Phys 2008;70:1530-6.
- 101. Tao Y, Lefkopoulos D, Ibrahima D, et al. Comparison

Page 14 of 14

of dose contribution to normal pelvic tissues among conventional, conformal and intensity-modulated radiotherapy techniques in prostate cancer. Acta Oncol 2008;47:442-50.

- 102. Diallo I, Haddy N, Adjadj E, et al. Frequency distribution of second solid cancer locations in relation to the irradiated volume among 115 patients treated for childhood cancer. Int J Radiat Oncol Biol Phys 2009;74:876-83.
- 103. Schneider U, Besserer J, Mack A. Hypofractionated radiotherapy has the potential for second cancer reduction. Theor Biol Med Model 2010;7:4.
- 104. Tian X, Liu K, Hou Y, et al. The evolution of proton beam therapy: Current and future status. Mol Clin Oncol 2018;8:15-21.
- 105. Chung CS, Yock TI, Nelson K, et al. Incidence of second malignancies among patients treated with proton versus photon radiation. Int J Radiat Oncol Biol Phys 2013;87:46-52.
- 106. Fontenot JD, Lee AK, Newhauser WD. Risk of secondary malignant neoplasms from proton therapy and intensitymodulated x-ray therapy for early-stage prostate cancer. Int J Radiat Oncol Biol Phys 2009;74:616-22.
- 107.Eaton BR, MacDonald SM, Yock TI, et al. Secondary Malignancy Risk Following Proton Radiation Therapy. Front Oncol 2015;5:261.
- 108. Xiang M, Chang DT, Pollom EL. Second cancer risk after primary cancer treatment with three-dimensional conformal, intensity-modulated, or proton beam radiation therapy. Cancer 2020;126:3560-8.
- 109.Berrington de Gonzalez A, Curtis RE, Kry SF, et al. Proportion of second cancers attributable to radiotherapy treatment in adults: a cohort study in the US SEER cancer registries. Lancet Oncol 2011;12:353-60.
- 110. Safwat A, Bentzen SM, Turesson I, et al. Deterministic rather than stochastic factors explain most of the variation

doi: 10.21037/amj-20-179

Cite this article as: Linkowski LC, Manley BJ, Johnstone PAS, Grass GD. A review of radiation-related malignancy in the pelvis. AME Med J 2022;7:3.

in the expression of skin telangiectasia after radiotherapy. Int J Radiat Oncol Biol Phys 2002;52:198-204.

- 111.Bergom C, West CM, Higginson DS, et al. The Implications of Genetic Testing on Radiation Therapy Decisions: A Guide for Radiation Oncologists. Int J Radiat Oncol Biol Phys 2019;105:698-712.
- 112.Menes TS, Terry MB, Goldgar D, et al. Second primary breast cancer in BRCA1 and BRCA2 mutation carriers:
 10-year cumulative incidence in the Breast Cancer Family Registry. Breast Cancer Res Treat 2015;151:653-60.
- 113.He Y, Liu H, Wang S, et al. A nomogram for predicting cancer-specific survival in patients with osteosarcoma as secondary malignancy. Sci Rep 2020;10:12817.
- 114. Bernstein JL, Thomas DC, Shore RE, et al. Contralateral breast cancer after radiotherapy among BRCA1 and BRCA2 mutation carriers: a WECARE study report. Eur J Cancer 2013;49:2979-85.
- 115. Mai PL, Best AF, Peters JA, et al. Risks of first and subsequent cancers among TP53 mutation carriers in the National Cancer Institute Li-Fraumeni syndrome cohort. Cancer 2016;122:3673-81.
- 116. Andreassen CN, Rosenstein BS, Kerns SL, et al. Individual patient data meta-analysis shows a significant association between the ATM rs1801516 SNP and toxicity after radiotherapy in 5456 breast and prostate cancer patients. Radiother Oncol 2016;121:431-9.
- 117.Fachal L, Gomez-Caamano A, Barnett GC, et al. A three-stage genome-wide association study identifies a susceptibility locus for late radiotherapy toxicity at 2q24.1. Nat Genet 2014;46:891-4.
- 118.Kerns SL, Dorling L, Fachal L, et al. Meta-analysis of Genome Wide Association Studies Identifies Genetic Markers of Late Toxicity Following Radiotherapy for Prostate Cancer. EBioMedicine 2016;10:150-63.