



Impact of *RHCE* variability and complexity in transfusion medicine: a narrative review

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Background and Objective: Rh is one of the most clinically important blood group systems. It comprises five major antigens (D, C, c, E, and e) in addition to several low and high prevalence antigens. RhD and RhCE proteins are encoded by two adjacent genes namely *RHD* and *RHCE*. These genes are highly homologous and polymorphic resulting in numerous variant alleles that encode variant antigens. In this review, we will discuss *RHCE* genetic diversity, frequency of *RHCE* variant alleles and their clinical aspects, complexities, and the importance of *RH* genotyping to prevent alloimmunization.

Methods: A search restricted to English language was performed using the PubMed electronic database to retrieve publications until June 2021, on *RHCE* variability and complexity. The Medical Subject Headings and free terms used were Rh-Hr Blood-Group System/genetics, sickle cell disease (SCD), *RHCE*, *RHCE* variants, and Rh antibodies. Other relevant articles were found by checking the reference list of the articles collected in the initial search.

Key Content and Findings: Common RhCE antigens are highly immunogenic, and their alloantibodies have been involved in delayed hemolytic transfusion reactions (DHTRs) and hemolytic disease of the fetus and newborn (HDFN). Because the distribution of C/c and E/e is different among ethnic groups, several treatment centers adopt prophylactic CE matching protocols to prevent alloimmunization in chronically transfused patients. Despite measures, alloantibody formation against RhCE antigens is still a problem due to numerous variant phenotypes, mainly in individuals of African origin. Elucidation of molecular basis for *RHCE* variants has allowed to determine the frequency of these alleles in patients and donors of African descent, however, the clinical significance of the Rh alloantibodies elicited by most *RHCE* variants is still unclear.

Conclusions: Molecular assays allowed considerable progress in the identification of genetic basis and characterization of *RHCE* variant alleles in patients with SCD and blood donors. Advances in molecular techniques may enable the screening of rare RhCE donors and improve the support for patients carrying variant RhCE phenotype. However, some questions remain to be answered, mainly regarding the clinical significance of the *RHCE* variants.

Keywords: Rh blood group system; *RHCE* variants; Rh antibody; RhCE protein; sickle cell disease (SCD)

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Introduction

Rh is the second most clinically important of all blood group systems after ABO. It encompasses 56 antigens including five major antigens (D, C, c, E, and e) in addition to low and high prevalence antigens identified by their corresponding antibodies (1,2). *RHD* and *RHCE* genes, which encode Rh antigens, are highly homologous and polymorphic with more than 700 *RHD* and 200 *RHCE* alleles reported to date (1,3-5). Many described *RH* alleles have been identified in individuals of African origin, and it is estimated that 87% of patients with sickle cell disease (SCD) and African descent blood donors carry at least one variant *RH* allele (6). This diversity creates clinical challenges and causes significant rates of Rh alloimmunization (7).

Rh antibodies are the most common antibodies identified in transfused patients, and despite the serological matching for D, C, and E antigens and racially matched blood transfusions (6), Rh alloimmunization persists due to variant Rh antigens present either in patients or blood donors (6-8). Importantly, some variant *RH* alleles have been associated with development of clinically significant alloantibodies causing delayed hemolytic transfusion reactions (DHTRs) or hemolytic disease of the fetus and newborn (HDFN) (6,7,9-11). Ideally, patients carrying Rh variants with an antibody to a high-prevalence antigen or with multiple common antibodies need compatible red blood cell (RBC) units. However, providing such compatible units is often a challenge because *RH* genotyping, required to identify the Rh variants, is expensive and is not a routine method; consequently, partial antigens are usually recognized once alloantibodies have already been formed or when a transfusion reaction has occurred (12). In this review, we will discuss the molecular basis for altered RhCE phenotypes, frequency of variant *RHCE* alleles, clinical significance of alloantibodies, SCD and *RHCE* complexities, and the importance of *RH* genotyping to prevent alloimmunization. We present the following article in accordance with the Narrative Review checklist (available at <https://aob.amegroups.com/article/view/10.21037/aob-21-76/rc>).

Methods

The literature search strategy summary and the detailed search strategy used in PubMed database to retrieve publications on *RHCE* variability and complexity are shown in Table 1 and Table S1, respectively. In addition, the *RHCE* allele frequency was collected from the ErythroGene

database in the 1000 Genomes dataset (13) and the nucleotide changes confirmed by checking the Blood Group Terminology Table for *RHCE* at the International Society of Blood Transfusion (ISBT) website (1).

Overview on Rh blood group system

The Rh blood group system encompasses two highly homologous genes, *RHD* and *RHCE*, closely located on the short arm of chromosome 1 (1p36.11), which encode RhD and RhCE proteins differing in only 32–35 of 417 amino acids (14-17). These genes are inherent together as a haplotype, each composed of ten exons, and have more than 90% sequence similarity (18). The *RHD* gene encodes the RhD protein, carrying the D antigen (RH1) while the *RHCE* gene encodes the RhCE protein, carrying C (RH2) or c (RH4) and E (RH3) or e (RH5) antigens. The RhD and RhCE proteins are integral to the red cell membrane and form a complex with the RhAG protein, a chaperone required for Rh antigens expression, encoded by *RHAG* gene on chromosome 6. The absence of RhAG protein causes lack of RhD and RhCE protein expression leading to a rare Rh_{null} phenotype, named regulator Rh_{null}. Individuals with this phenotype can readily form alloantibodies on exposure of Rh antigens (3,15,19).

RhCE antigens

The four main *RHCE* alleles encode the Ce, CE, ce, and cE antigen combinations (3) and changes to the *RHCE* gene can alter their antigen expression and/or generate new antigens (20). The C and c antigens specificity are determined by 4 non-synonymous substitution, c.48G>C (p.Trp16Cys), c.178C>A (p.Leu60Ile), c.203G>A (p.Asn68Ser), and c.307T>C (p.Pro103Ser); and 2 synonymous substitution c.150C>T and c.201A>G (1,21). Among these changes, only the p.Pro103Ser substitution, predicted to reside on the second extracellular loop of the RhCE protein, is associated with the C/c immunogenicity (15,21). As amino acids encoded by exon 2 of the *RHCE* allele are identical to those encoded by exon 2 of the *RHD* gene (22), next-generation sequencing data strongly support that a hybrid allele *RHCE**CE-D(2)-CE is causal for the C+ antigen expression (22,23). The molecular basis for E and e specificities are determined by the nucleotide change c.676G>C in exon 5, resulting in the amino acid substitution, p.Ala226Pro, located on the fourth extracellular loop of the RhCE protein (2).

Table 1 Search strategy summary

Items	Specification
Date of search	July 01, 2021
Databases and other sources searched	PubMed and reference lists of articles identified in the search
MeSH and free terms used	Rh-Hr Blood-Group System/genetics (MeSH), sickle cell disease (MeSH), RHCE, RHCE variants, Rh antibodies
Timeframe	The limit of database until June 30, 2021
Inclusion and exclusion criteria	Articles restricted to English language. No restrictions on publication type
Selection process	Independent literature search was performed by the authors (ES and CPA) using MeSH and free terms separately and in combination. Titles/abstracts of retrieved articles were checked for relevance and selected for further review if addressed <i>RHCE</i> genetics, frequency of <i>RHCE</i> variant alleles, clinical significance of variant <i>RHCE</i> alleles, impact of variant RhCE on patients with SCD, or molecular characterization of variant <i>RHCE</i> alleles. Additional relevant papers were identified by manual searching of reference lists of articles identified in the initial search

SCD, sickle cell disease.

Molecular basis of *RHCE* variants

Molecular mechanisms responsible for altered or null Rh phenotypes have been revealed through molecular typing of patients and donors from different ethnic backgrounds and over 200 *RHCE* alleles have been described to date (1,4,24-28). The genetic diversity of the *RHCE* gene is generated by at least four molecular mechanisms: (I) single nucleotide variations (SNVs), (II) insertions, (III) deletions, and (IV) gene rearrangements (gene conversion), that may cause weaken and/or partial expression of C, c, E, and e, induce expression of low-prevalence antigens, and/or loss of expression of high-frequency antigens. Furthermore, some genetic alterations in the *RHCE* may result in a non-functional RhCE protein (e.g., D⁻, DC^{w-}, Dc⁻ and D^{••}). The inheritance of non-functional *RHCE* alleles in conjunction with deleted *RHD* results in the lack of any Rh proteins on the red cells membrane giving rise to the amorph Rh_{null} phenotype (2,15).

SNV is the main molecular mechanism responsible for RhCE protein alterations. SNVs are often associated with weak RhCE antigen expression when located in the coding regions inducing an amino acid change in the transmembrane or intracellular regions (29). In contrast, an amino acid change in extracellular regions or a change in a transmembrane or intracellular region causing conformational alterations, can alter epitopes and produce partial phenotypes (e.g., partial “e” and “c” due to c.733C>G) which are prone to immunization when exposed to the normal antigens. Because antigen expression is not

always reduced in partial Rh phenotypes and serological methods cannot distinguish *RH* variants, partial antigens are usually recognized after alloantibodies against missing epitopes are formed or when a transfusion reaction has occurred (9). *RHCE* variants may also cause both weak and partial antigen expression; for example, the *RHCE*^{*ceAG} allele is associated with a weak and partial “e” due to an amino acid substitution (p.Ala85Gly) caused by the SNV c.254C>G in exon 2 (30). In addition, SNVs can also produce stop codons that prematurely terminate protein synthesis, generating null phenotypes [e.g., “E⁻” and “c⁻” due to c.221G>A (p.Trp74Ter)] (28).

Single amino acid substitutions in the RhCE protein also can generate new epitopes. These new epitopes are called “low prevalence antigens”, occur in less than 1% of the population and are not routinely typed for, but are potentially immunogenic. Examples include C^w (31), C^x (31), Crawford (32), E^w (33), JAL (34-36), V and VS (37-39). However, in individuals of African origin V and VS antigen are reasonably common (37). Both of them are associated with the SNV c.733C>G (p.Leu245Val) predicted to be located in the eight transmembrane segment of Rhce protein which causes a conformation change within the ce polypeptide leading to partial c and partial e antigens (38,39). The subsequent loss of V expression results from the c.1006G>T (p.Gly336Cys) change on this background arising the V-VS⁺ phenotype (39).

Insertions and deletions are less frequent and generally result in a frameshift and a premature stop codon. For

instance, the *RHCE*ceN.01* allele associated with deletion of five nucleotides at positions 80-84 (c.80_84delTCTTC) introduce a frameshift after p.Leu26 (CTC) causing a premature stop codon. As a result, Rhce antigen expression is completely abolished from the RBCs (40).

Gene rearrangement between *RHD* and *RHCE* is common and associated with the formation of hybrid alleles that is favored by the proximity, homology and tail-to-tail orientation of RH genes (41). For example, the most frequent mechanism associated with partial C in individuals of African origin are the hybrid *RHCE*Ce-D(4)-ce* (*RHCE*CeRN*) identified in R^N individuals, and the *RHD*DIIIa-CEVS(4-7)-D* and *RHD*D-CEVS(4-7)-D* alleles that are part of (C)ce^S haplotype 1 and type 2, respectively (9,39,42-45).

*RHCE*CeRN* encodes weak and partial expression of C and e antigens, absence of the high-prevalence Sec antigen, and expression of low-prevalence R^N and DAK antigens. In addition, R^N erythrocytes slightly overexpress the D antigen (46,47). The R^N haplotype has been described in people of African origin and found to be responsible for partial C antigen expression in 11.8% of C+ patients with SCD in France (48), although in other countries its frequency is lower or absent (12,49,50).

*RHD*DIIIa-CEVS(4-7)-D* and *RHD*D-CEVS(4-7)-D* alleles are linked to *RHCE*ceVS.03* (ce48C, 733G, 1006T) composing (C)ce^S haplotypes, which do not encode the D antigen, instead, they encode partial C, c and e antigens, and loss of highly prevalent hr^B and Hr^B antigens (9,39,43,51). The partial C encoded by (C)ce^S haplotype 1 may have variable expression and, in many cases, go undetected until alloimmunization occurs (38,39,42). In Caucasians, weak C and weak e have been associated with diverse molecular events, for example, *RHCE*CeMA* allele result from the SNV c.340C>T in exon 3 which also cause expression of the low-prevalence Rh antigen, JAL (34,36,52) while *RHCE*CeVA* result from the hybrid *RHCE-D(5)-CE* allele (53).

Overall, altered forms of e and/or c antigens have been associated with numerous *RHCE*ce* variant alleles in people of African origin (Table 2). Importantly, individuals with some homozygous *RHCE*ce* variant alleles may also have a loss of high prevalence antigens on the red cells increasing the alloimmunization risk, for instance, hr^S– (RH–18) in *RHCE*ceAR*, *RHCE*ceEK*, *RHCE*ceBI*, *RHCE*ceMO* and *RHCE*ceSM* carriers; hr^B– (RH–31) in *RHCE*ceAG*, *RHCE*ceMO*, *RHCE*ceS*, *RHCE*ceCF*, and *RHCE*ceVS.02.01* carriers (30,55,56,62). These variants should be well-characterized for transfusion purposes

because their carriers are at risk of alloimmunization. Moreover, finding compatible blood for patients carrying these variants in both alleles can be a challenge since the molecular background of each variant is distinct and they are also often inherited along with *RHD* variant alleles, consequently, rare antigen-negative RBCs will be required for transfusion (37).

E antigen variants are rare and associated with diverse molecular mechanisms encoding weak or partial E antigen and have been mostly identified in Caucasians (29,63,64). One of the most important alleles reported in association with E variants is *RHCE*ceEW* previously reported as E Variant I. This allele is characterized by the amino acid substitution p.Met167Lys (c.500T>A) located at the third extracellular loop of the RhcE protein leading to an E+ partial, weak or negative phenotype (64). In addition, the p.Met167Lys substitution is also the molecular basis for the rare E^w (RH11) antigen (<0.1% in Caucasians) first described in 1955 (65) and associated with few cases of HDFN (33,63,65,66).

RHCE variant alleles can be inherited in combination with specific *RHD* variant alleles creating an additional degree of complexity and a challenge for transfusion. Some combinations are much higher than expected to occur by chance, indicating linkage of *RHD* alleles encoding partial D with specific altered *RHCE*ce*; for example, *RHCE*ceAR* and *RHCE*ceEK* are often in linkage to *RHD*DAR* (54,55), *RHCE*ce^S* is linked to *RHD*DIIIa* (61), *RHCE*ceTI* is linked to *RHD*DIVa-2* (25), and *RHCE*ceMO* is often found with *RHD*DAU0* (56). Frequent RH alleles presenting linkage are listed in Table 2.

Frequency of *RHCE* variants

RHCE variants are more frequent in African descendants and people with mixed ancestry than in Caucasians and Asians (45,49,50,67). Studies on the diversity and frequency of RH alleles in blood donors, and/or patients with SCD who are at high risk of alloimmunization have been conducted using both in-house and commercial genotyping assays (45,49,50,60,67-76). The reported frequencies of *RHCE* variant alleles are inconsistent among reports likely due to several reasons, including differences in study design, molecular strategies used for *RHCE* characterization, and population ethnicity. Additional bias includes lack of consensus on the clinical significance of *RHCE* variants resulting from c.48G>C and c.733C>G [*RHCE*ce.01* (ce48C), *RHCE*ceVS.01* (ce733G), and *RHCE*ceVS.02*

Table 2 Summary of the most frequent variant *RHCE* alleles

<i>RHCE</i> allele [common name (1)]	<i>RHCE</i> allele [ISBT name (1)]	References	Nucleotide change	Predicted phenotype			Often linked to <i>RHD</i> allele	Population frequency [†]
				HFA	LFA	PA		
<i>RHCE</i> *ceTi	<i>RHCE</i> *01.02.01	(25)	c.48G>C, c.1025C>T			e ⁺ , c ⁺	<i>RHD</i> *D1Va-2	Africa: 2.27%, America: 0.43%, East Asia: 0%, Europe: 0%
<i>RHCE</i> *ceAR	<i>RHCE</i> *01.04.01	(54)	c.48G>C, c.712A>G, c.733C>G, c.787A>G, c.800T>A, c.916A>G	Hi ^{r-} , hr ^{s-}	V ⁺ , VS ⁻	e ⁺ , c	<i>RHD</i> *DAR	NR
<i>RHCE</i> *ceEK	<i>RHCE</i> *01.05.01	(55)	c.48G>C, c.712A>G, c.787A>G, c.800T>A	Hi ^{r-} , hr ^{s-}		e ⁺ , c	<i>RHD</i> *DAR	NR
<i>RHCE</i> *ceAG	<i>RHCE</i> *01.06.01	(30)	c.254C>G	hr ^{B-} , CEAG ⁻		e ⁺	<i>RHD</i> deletion	Africa: 5.60%, America: 0.72%, East Asia: 0%, Europe: 0%
<i>RHCE</i> *ceMO	<i>RHCE</i> *01.07.01	(56)	c.48G>C, c.667G>T	hr ^{s-} , hr ^{B-} , CEVF ⁻		e ⁺ , c	<i>RHD</i> *DAU0	Africa: 1.44%, America: 0.43%, East Asia: 0.20%, Europe: 0.10%
<i>RHCE</i> *ceBI	<i>RHCE</i> *01.08	(57-59)	c.48G>C, c.712A>G, c.818C>T, c.1132C>G	Hi ^{r-} , hr ^{s-}	STEM ⁺	e ⁺	<i>RHD</i> *DOL1 or <i>RHD</i> *DOL2	Africa: 0.08%, America: 0%, East Asia: 0%, Europe: 0%
<i>RHCE</i> *ceSM	<i>RHCE</i> *01.09	(57)	c.48G>C, c.712A>G, c.818C>T	Hi ^{r-} , hr ^{s-}	STEM ⁺ [†]		<i>RHD</i> *DOL1 or <i>RHD</i> *DOL2	NR
<i>RHCE</i> *ceVS.01	<i>RHCE</i> *01.20.01	(38)	c.733C>G	hr ^{B+} [†]	V ⁺ , VS ⁺	e, c		Africa: 15.28%, America: 2.31%, East Asia: 0%, Europe: 0.30%
<i>RHCE</i> *ceVS.02.01	<i>RHCE</i> *01.20.02.01	(38,60)	c.48G>C, c.733C>G	hr ^{B-}	V ⁺ , VS ⁺	e, c	<i>RHD</i> *DAR3	Africa: 2.87%, America: 0%, East Asia: 0%, Europe: 0%
<i>RHCE</i> *ceS	<i>RHCE</i> *01.20.03	(38,51,61)	c.48G>C, c.733C>G, c.1006G>T	hr ^{B-}	V ⁻ , VS ⁺	e, c	<i>D-CE</i> (4-7)-D or <i>DIIIa-CE</i> (4-7)-D	Africa: 4.46%, America: 0.29%, East Asia: 0%, Europe: 0%
<i>RHCE</i> *ceCF	<i>RHCE</i> *01.20.06	(32,62)	c.48G>C, c.697C>G, c.733C>G	hr ^{S-} , hr ^{B-} , Crawford ⁺ , CELO ⁻	VS ⁺	e, c		Africa: 0.08%, America: 0%, East Asia: 0%, Europe: 0%, South Asia: 0%
<i>RHCE</i> *CeRN	<i>RHCE</i> *02.10.01	(46,47)	c.505C>A, c.509G>T, c.514T>A, c.544A>T, c.577A>G, c.594T>A, c.602G>C	Sec ⁻	DAK ⁺	C ⁺ , e ⁺		NR

[†], from Erythrocyte database (13); [†], weakened expression. HFA, high frequency antigen; LFA, low frequency antigen; PA, partial antigen; NR, not reported.

(ce48C, 733G)].

In a study from France, including blood donors and patients of African origin, showed that among individuals with altered expression of RhCE antigens and/or with anti-RhCE alloantibodies in the presence of the corresponding antigen 83% had variant *RHCE* alleles and *RH* haplotypes, and the most frequent were R^N and (C)ce^s haplotypes, *RHCE*ceMO*, and *RHCE*ceAR* alleles (75). The same group reported in a later study, in French blood donors of African origin that 14.2% of that population had a variant *RHCE* allele, being (C)ce^s type 1 haplotype the most frequent followed by *RHCE*ceTI*, *RHCE*ceMO*, and *RHCE*ceAR* alleles (45); suggesting that systematic screening of donors for *RHCE* increases the chances of finding rare *RHCE* variants and may help to fulfill the transfusion needs of patients requiring an *RHCE* genotype matching in France (45).

In the US, approximately 85% of patients with SCD carry at least one variant *RH* allele (6). Independent studies reported similar frequencies of *RHCE* variants in African-American blood donors and patients with SCD, where *RHCE*ce.01* (ce48C), *RHCE*ceVS.01* (ce733G), *RHCE*ceTI*, *RHCE*ceAG*, and *RHCE*ceMO* were the most frequent *RHCE* alleles reported; indicating that they probably would be able to provide transfusion support to patients with SCD, although a large number of genotyped blood donors would be needed (37,49).

In Brazil, where the population is highly admixed, the presence of at least one clinically relevant *RHCE* variant allele has been found in approximately 45% of patients with SCD (12,50) and 53% of blood donors self-declared as of African origin (12,60). However, the frequency of homozygous *RHCE* variant alleles or compound heterozygous in patients with SCD and donors who self-declared as of African origin in Brazil is considerably lower, ranging from 1.4% to 16.9% (12,50,67). Although *RHCE* variant alleles between patients and donors are similar to those found in the US, Brazilian patients with SCD and self-declared African origin donors have a high frequency of R1r phenotype which demonstrates the genetic influence of Caucasian origin (12,50). Nevertheless, characterization of donors self-declared as of African origin is the best choice for finding compatible blood for patients with SCD, since both groups have similar frequencies of RhCE phenotypes and *RHCE* variant alleles (12,77).

Clinical aspects

Alloimmunization is a major adverse effect of blood

transfusion, increases the risk of DHTRs and reduces the availability of compatible RBC units. Alloimmunization rates vary depending on antigen profile disparity between blood donors and patient, level of antigen immunogenicity, patient age, medical conditions, and frequency of transfusion events (52,78-80).

Rh antigens are highly immunogenic and can induce not only alloantibodies but also autoantibodies. Rh alloantibodies are the most frequent antibodies in chronically transfused patients (52,78,80). Epidemiological study performed using the “Recipient Epidemiology and Donor Evaluation Study-III” (RED-III) database showed that antibodies against RhCE antigens comprised 47.5% of the clinically significant antibodies detected in the 6597 alloimmunized patients (80). From these, 61.3% were anti-E, 18.5% anti-C, 13.2% anti-c, and 2.5% anti-e (80). To prevent alloimmunization, it has been recommended the use of prophylactic Rh (C/c, E/e) and K antigen matched transfusion for chronically transfused patients, especially for patients with SCD who are at high risk of alloimmunization and have the highest rates of Rh antibodies (81,82). Implementation of this practice has demonstrated to reduce alloimmunization rate and DHTRs (78,83,84).

In warm autoimmune hemolytic anemia, about 80% of patients have in their serum autoantibodies that react optimally at 37 °C (2). Although most of these autoantibodies appear to be “nonspecific”, many of them have a specificity, and anti-e is the most common followed by anti-c, -E, -D, and -C. Noteworthy, when investigating an apparent autoantibody with Rh specificity or unexplained Rh antibodies, RhCE variants should be considered since they can result in partial antigens that elicit alloantibody formation.

The most common *RHCE* variant alleles found among African descent individuals are *RHCE*ce.01* (ce48C), *RHCE*ceVS.01* (ce733G), and *RHCE*ceVS.02* (ce48C, 733G) but the clinical impact of these variants is questionable. Although the Rhce protein encoded by *RHCE*ce.01* (ce48C) allele does not lack epitopes and the e antigen is not recognized as partial (85), there are studies reporting clinical significance of anti-e in patients who carry *RHCE*ce.01* (ce48C) allele (6,7,86). Analysis of hemoglobin (Hb) and hemoglobin S (HbS) levels in pre- and post-transfusion events showed a reduction of Hb and an increase in HbS levels in a SCD patient carrying anti-e and *RHCE*ce.01*/(C)ce^s genotype who was transfused with RBC e+ (7); and an improved response to transfusion was observed in another SCD patient homozygous for

*RHCE*ce.01* with anti-e, who received a genotyped-matched transfusion (86). Despite these findings, the clinical significance of this variant is still controversial and unclear, and it is currently recognized that patients with these alleles seem to have a lower risk of Rh alloimmunization than patients with other altered alleles (49,85).

*RHCE*ceVS.01* (ce733G) and *RHCE*ceVS.02* (ce48C, 733G) also have questioned clinical significance although both alleles are predicted to generate partial c and e antigens and have also been associated with alloantibodies (6,7,50,87). For example, a study including 16 e+ SCD patients with anti-e, reported that 12 of them (75%) presented *RHCE*ce.01*, *RHCE*ceVS.01* or *RHCE*ceVS.02* alleles, and 4 of those 12 (33%) had evidence of DHTR due to anti-e (6). In contrast, some experts do not consider that *RHCE*ceVS.01* (ce733G) and *RHCE*ceVS.02* (ce48C, 733G) encode partial antigens because in their experience the anti-e antibodies associated with these molecular backgrounds are auto-antibodies (48).

The clinical importance for most antibodies formed in patients carrying variant RhCE phenotypes is not easy to establish because the individual variability to alloimmunization remains poorly understood and the reports are often incomplete, particularly regarding serology data (88,89). Furthermore, the classification of allo- or auto-antibody and the role of the antibodies in a DHTR may be difficult to ascertain because allo- as well as auto-antibodies can lead to DHTRs (11,90-92). Ideally, to discriminate allo- and auto-antibody an auto-adsorption assay should be performed. Nevertheless, auto-adsorption assay cannot be performed in a recently transfused patient and results may be inconclusive for very weakly expressed antigens (93).

Besides partial e, partial c also arises from variant *RHCE* alleles, but fewer cases are reported compared with anti-e. Alteration on c antigen expression is rarely found in variant phenotypes probably because the structure of c epitope(s) involves two adjacent proline residues that might form a more stable structure resistant to perturbations induced by changes in upstream or downstream (94). Nevertheless, some cases of partial c involving different variant *RHCE* alleles have been reported; for example, *RHCE*ceAR* (95,96), *RHCE*ceMO* (55), *RHCE*ceJAL* (35), *RHCE*ceCF* (62), *RHCE*ceEK* (10), *RHCE*ceTI* (25) and also (C)ce^s haplotype (51).

Anti-C elicited by partial C expression is also commonly reported, mostly in patients with SCD. Studies in a cohort of SCD patients showed that 20–30% of patients with C+ phenotype have partial C, mostly as a result of (C)

ce^s and R^N haplotypes, and have a high risk of anti-C alloimmunization if transfused with conventional C+ units (9,48,49). Analysis of the clinical significance of this antibody showed heterogeneous results; however, some reports revealed DHTR after transfusion with C+ RBCs (6,9).

Patients carrying variant *RHCE* alleles with lack of expression of high-frequency Rh antigens including Hr^B (RH34), hr^B (RH31), Hr (RH18), and/or hr^S (RH19) are at risk for alloimmunization. Anti-Hr^B (RH34) and anti-hr^B (RH31) are found mainly in African descent individuals carrying (C)ce^s haplotypes (97,98), and anti-Hr and anti-hr^S are commonly found in patients carrying *RHCE*ceAR*, *RHCE*ceEK* and *RHCE*ceBI*. Anti-Hr^B and anti-Hr react with all RBCs of common RhCE phenotype, but react strongly with e+ RBCs (98), while anti-hr^B and anti-hr^S react with e+ RBCs, preferentially with Ce and ce haplotypes, respectively, but do not react with e- (DccEE) RBCs (97). Hence, when Hr^B- individuals have an antibody reacting against all RBCs of common Rh phenotype, identification of anti-Hr^B associated with anti-hr^B is possible through adsorption studies with e- (DccEE) RBCs. Anti-Hr^B is adsorbed on e- RBCs and the remaining reactivity in the serum is an anti-hr^B. Similarly, when anti-Hr is adsorbed on e- RBCs, the remaining reactivity in the serum is an anti-hr^S (55,99,100). The clinical significance of anti-Hr^B and anti-Hr is well established for both RBC transfusion and HDFN (55,94,96), and some studies have reported an association of anti-hr^S and anti-hr^B with adverse effects on the fetus or DHTR (6,7,9,10,35,56,101). Once anti-hr^B or anti-hr^S is identified, compatible transfusion can be achieved by providing e- RBCs. However, these patients may develop anti-E (if E-), and anti-Hr^B or -Hr which may lead to complications in antibody identification and provision of suitable blood.

The mechanism for loss of expression of hr^S and hr^B has not been fully elucidated, and specific epitope(s) and residues involved have not been definitively localized on the Rh proteins (30,56). The inconsistency in serologic results and the lack of antisera support the use of *RH* genotyping for classification of RBCs with altered Rh antigens. In addition, multiple molecular backgrounds encode similar phenotype as shown in Table 2; however, patients with antibodies elicited by those variants are not always compatible with donor's RBCs with the same RhCE phenotype but different molecular background (37,96). For example, anti-c developed by a patient carrying *RHCE*ceAR*/Ce reacts with RBCs with *RHCE*ceEK* and *RHCE*ceBI*, but not with *RHCE*ceMO* and *RHCE*ceJAL*, suggesting that

the c antigen encoded by *RHCE*ceAR* allele is different than that encoded by *RHCE*ceEK* and *RHCE*ceBI* and may express common epitopes with the c antigens encoded by *RHCE*ceMO* and *RHCE*ceJAL* (96). Therefore, for efficient and safe blood transfusion, *RH* genotyping and molecular matching is recommended.

SCD and *RHCE* complexities

Patients with SCD are chronically transfused and usually highly immunized for Rh antigens. The difference in frequency of RhCE antigens among ethnic groups greatly contributes to alloimmunization because in many countries blood donors are mostly of European descent, while SCD is prevalent in African descendants (102–104). Selection of blood donors self-declared as African descent for patients with SCD is a good transfusion strategy adopted by several centers to provide a more similar phenotypic profile and avoid RBC alloimmunization without overuse of D– RBC units, since in individuals from African origin the haplotype Dce is more frequent, while in Caucasians DCe haplotype is more common (49).

A common strategy for reducing the alloimmunization risk in patients with SCD has been to provide prophylactic matching RBC units for C, E, and K antigens (105,106). Regardless of this strategy, alloimmunization against Rh antigens continues to occur due to Rh complexities. The presence of variant *RHCE* alleles in the SCD population has been shown to range from 27% to 58% (6,12,37,67) and patients with SCD carrying variant alleles have a higher risk for alloimmunization (7,48). In the US, 13% of patients with SCD who developed RhCE-alloantibodies were carriers of partial antigen (49), and in France the presence of anti-C was detected in 14.3–30% of patients with partial C antigen (9,88).

Recent evidence about the impact of variant RhCE on patients with SCD has expanded the application of *RH* genotyping since the variants are not distinguished by serological techniques. Special attention to RhCE phenotyping may indicate an altered antigen expression, and further genotyping can inform if the patient is at risk for alloimmunization or to provide insight to determine if Rh antibodies are allo- or auto-antibodies, predict clinical significance, and aid in transfusion decisions (49).

RH genotyping is a great strategy to provide superior matching, reduce alloimmunization and improve red cell utilization (107). Genotype-matching can be achieved by high-throughput genotyping, which offers significant cost

savings in both labor and reagents compared with antigen typing by serologic methods, and expands testing to detect genetic variation of antigen expression (107). However, genotyping is still of high cost, fact that leads to the development and use of selection strategies to screen donors with rare RhCE phenotypes, to increase the probability to find them at an affordable cost. The recruitment of donors self-declared as African descent to provide *RH* genetic matching has also been the best choice, as indicated by reports that showed similar frequency of *RHCE* variant alleles among African descendant donors and patients with SCD, even in countries with ethnic admixture (12,49,77). Patient classification in responders and non-responders could be important to restrict molecular matching to patients with higher chance to develop alloantibody (48,105). Centralization of genotyping tests in larger centers has been suggested as alternative that would give support to the smaller centers that frequently transfuse patients with SCD (105). Alternatively, prioritizing some conventional molecular tests (allele-specific PCR or PCR-RFLP) targeting specific SNVs, such as c.733G>C, c.254C>G and c.667G>T to screen the most common variants can also be a useful strategy (108). Additionally, selection of donors with Fy(a–b–) or weak D phenotype may increase the chances of identifying donors lacking high-frequency RhCE antigens (12,77).

Conclusions

RhCE antigens have a significant role in transfusion medicine due to their high immunogenicity and the hemolytic power of their antibodies. In the last two decades, studies have been conducted to elucidate the presence of unexplained RhCE antibodies. Molecular assays allowed considerable progress in the identification of genetic basis and characterization of *RHCE* variant alleles in patients with SCD and blood donors. Current knowledge about variant allele frequencies in different ethnicities allows the blood centers to define target variants for screening aiming to provide matched RBCs to most patients. However, some questions remain to be answered, mainly regarding the clinical significance of the *RHCE* variants. Except for anti-Hr^B and anti-Hr, the clinical importance of antibodies elicited by variant phenotypes is unclear. The interpretations of events that define clinical significance are inconsistent and require comprehensive studies with extensive serological tests and clear association with clinical observations. Additionally, although more than one genetic

variant can predict the same phenotype some of those can be incompatible for transfusion. Therefore, studies that evaluate cross-matching among genetic variants predicting similar phenotype but with different genetic backgrounds are paramount to use genotyping for expanding the availability of donors.

Finally, transfusion of patients carrying variant RhCE phenotype is still a significant challenge in transfusion medicine. Although genotyping revolutionized the knowledge of RhCE variants, its application in routine immunohematology, for donor screening and molecular-matching, is still cost-prohibitive. We expect that technological advances, such as next-generation sequencing or large-scale genotyping microarray platforms, allow screening of rare RhCE donors to improve transfusion care for patients with variant RhCE phenotype.

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Footnote

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aspects of the work in ensuring that questions related to the accuracy or integrity of any part of the work are appropriately investigated and resolved.

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References

1. International Society of Blood Transfusion (ISBT) [Internet]. Red Cell Immunogenetics and Blood Group Terminology [cited 2021 Nov 01]. Available online: <https://www.isbtweb.org/working-parties/red-cell-immunogenetics-and-blood-group-terminology>
2. Daniels GL. Human Blood Groups. Third Edition ed. Oxford, UK: Wiley-Blackwell, 2013.
3. Flegel WA, Wagner FF. Molecular genetics of RH. Vox Sang 2000;78 Suppl 2:109-15.
4. Floch A, Téletchéa S, Tournamille C, et al. A Review of the Literature Organized Into a New Database: RHeference. Transfus Med Rev 2021;35:70-7.
5. Wagner FF, Flegel WA. The Rhesus Site. Transfus Med Hemother 2014;41:357-63.
6. Chou ST, Jackson T, Vege S, et al. High prevalence of red blood cell alloimmunization in sickle cell disease despite transfusion from Rh-matched minority donors. Blood 2013;122:1062-71.
7. Sippert E, Fujita CR, Machado D, et al. Variant RH alleles and Rh immunisation in patients with sickle cell disease. Blood Transfus 2015;13:72-7.
8. Waldis SJ, Uter S, Kavitsky D, et al. Rh alloimmunization in chronically transfused patients with thalassemia receiving RhD, C, E, and K matched transfusions. Blood Adv 2021;5:737-44.
9. Tournamille C, Meunier-Costes N, Costes B, et al. Partial C antigen in sickle cell disease patients: clinical relevance and prevention of alloimmunization. Transfusion 2010;50:13-9.
10. Walters TK, Lightfoot T. A delayed and acute hemolytic transfusion reaction mediated by anti-c in a patient with variant RH alleles. Immunohematology 2018;34:109-12.

11. Pirenne F, Yazdanbakhsh K. How I safely transfuse patients with sickle-cell disease and manage delayed hemolytic transfusion reactions. *Blood* 2018;131:2773-81.
12. Arnoni CP, Vendrame T, Muniz J, et al. RHCE diversity among Brazilian patients with sickle cell disease (SCD) and selected groups of blood donors. *Transfusion* 2021;61:3473-82.
13. Möller M, Jöud M, Storry JR, et al. ErythroGene: a database for in-depth analysis of the extensive variation in 36 blood group systems in the 1000 Genomes Project. *Blood Adv* 2016;1:240-9.
14. Flegel WA. The genetics of the Rhesus blood group system. *Blood Transfus* 2007;5:50-7.
15. Avent ND, Reid ME. The Rh blood group system: a review. *Blood* 2000;95:375-87.
16. Chérif-Zahar B, Bloy C, Le Van Kim C, et al. Molecular cloning and protein structure of a human blood group Rh polypeptide. *Proc Natl Acad Sci U S A* 1990;87:6243-7.
17. Le van Kim C, Mouro I, Chérif-Zahar B, et al. Molecular cloning and primary structure of the human blood group RhD polypeptide. *Proc Natl Acad Sci U S A* 1992;89:10925-9.
18. Okuda H, Suganuma H, Kamesaki T, et al. The analysis of nucleotide substitutions, gaps, and recombination events between RHD and RHCE genes through complete sequencing. *Biochem Biophys Res Commun* 2000;274:670-83.
19. Cartron JP. Defining the Rh blood group antigens. *Biochemistry and molecular genetics. Blood Rev* 1994;8:199-212.
20. Westhoff CM, Silberstein LE, Wylie DE, et al. 16Cys encoded by the RHce gene is associated with altered expression of the e antigen and is frequent in the R0 haplotype. *Br J Haematol* 2001;113:666-71.
21. Mouro I, Colin Y, Chérif-Zahar B, et al. Molecular genetic basis of the human Rhesus blood group system. *Nat Genet* 1993;5:62-5.
22. Westhoff CM. The Rh blood group system in review: a new face for the next decade. *Transfusion* 2004;44:1663-73.
23. Wheeler MM, Lannert KW, Huston H, et al. Genomic characterization of the RH locus detects complex and novel structural variation in multi-ethnic cohorts. *Genet Med* 2019;21:477-86.
24. Kulkarni S, Mishra G, Maru H, et al. Molecular characterization of rare D--/D-- variants in individuals of Indian origin. *Blood Transfus* 2022;20:59-65.
25. Westhoff CM, Vege S, Halter Hipsky C, et al. RHCE*ceTI encodes partial c and partial e and is often in cis to RHD*DIVa. *Transfusion* 2013;53:741-6.
26. Chen YX, Peng J, Novaretti M, et al. Deletion of arginine codon 229 in the Rhce gene alters e and f but not c antigen expression. *Transfusion* 2004;44:391-8.
27. Zhao FY, Li Q, Zhang DM, et al. A novel silent RHCE allele in Chinese population. *Transfus Med* 2019;29:430-3.
28. Ochoa-Garay G, Moulds JM, Cote J, et al. New RHCE variant alleles encoding the D- - phenotype. *Transfusion* 2013;53:3018-23.
29. Bugert P, Scharberg EA, Geisen C, et al. RhCE protein variants in Southwestern Germany detected by serologic routine testing. *Transfusion* 2009;49:1793-802.
30. Westhoff CM, Vege S, Hipsky CH, et al. RHCE*ceAG (254C>G, Ala85Gly) is prevalent in blacks, encodes a partial ce-phenotype, and is associated with discordant RHD zygosity. *Transfusion* 2015;55:2624-32.
31. Mouro I, Colin Y, Sistonen P, et al. Molecular basis of the RhCW (Rh8) and RhCX (Rh9) blood group specificities. *Blood* 1995;86:1196-201.
32. Flegel WA, Wagner FF, Chen Q, et al. The RHCE allele ceCF: the molecular basis of Crawford (RH43). *Transfusion* 2006;46:1334-42.
33. Strobel E, Noizat-Pirenne F, Hofmann S, et al. The molecular basis of the Rhesus antigen Ew. *Transfusion* 2004;44:407-9.
34. Westhoff CM, Vege S, Wylie D, et al. The JAL antigen (RH48) is the result of a change in RHCE that encodes Arg114Trp. *Transfusion* 2009;49:725-32.
35. Ong J, Walker PS, Schmulbach E, et al. Alloanti-c in a c-positive, JAL-positive patient. *Vox Sang* 2009;96:240-3.
36. Hustinx H, Poole J, Bugert P, et al. Molecular basis of the Rh antigen RH48 (JAL). *Vox Sang* 2009;96:234-9.
37. Reid ME, Halter Hipsky C, Hue-Roye K, et al. Genomic analyses of RH alleles to improve transfusion therapy in patients with sickle cell disease. *Blood Cells Mol Dis* 2014;52:195-202.
38. Faas BH, Beckers EA, Wildoer P, et al. Molecular background of VS and weak C expression in blacks. *Transfusion* 1997;37:38-44.
39. Daniels GL, Faas BH, Green CA, et al. The VS and V blood group polymorphisms in Africans: a serologic and molecular analysis. *Transfusion* 1998;38:951-8.
40. Kato-Yamazaki M, Okuda H, Kawano M, et al. Molecular genetic analysis of the Japanese amorph rh(null) phenotype. *Transfusion* 2000;40:617-8.
41. Wagner FF, Flegel WA. RHCE represents the ancestral RH position, while RHD is the duplicated gene. *Blood* 2002;99:2272-3.

42. Silvy M, Granier T, Beley S, et al. Identification of novel polymorphism restricted to the (C)ces type 1 haplotype avoids risk of transfusion deadlock in SCD patients. *Br J Haematol* 2013;160:863-7.
43. Pham BN, Peyrard T, Juszczak G, et al. Heterogeneous molecular background of the weak C, VS+, hr B-, Hr B- phenotype in black persons. *Transfusion* 2009;49:495-504.
44. Flegel WA, Wagner FF. Two molecular polymorphisms to detect the (C)ce(s) type 1 haplotype. *Blood Transfus* 2014;12:136-7.
45. Kappler-Gratias S, Auxerre C, Dubeaux I, et al. Systematic RH genotyping and variant identification in French donors of African origin. *Blood Transfus* 2014;12 Suppl 1:s264-72.
46. Rouillac C, Gane P, Cartron J, et al. Molecular basis of the altered antigenic expression of RhD in weak D(Du) and RhC/e in RN phenotypes. *Blood* 1996;87:4853-61.
47. Le Pennec PY, Rouger P, Klein MT, et al. A serologic study of red cells and sera from 18 Rh:32,-46 (RN/RN) persons. *Transfusion* 1989;29:798-802.
48. Floch A, Tournamille C, Chami B, et al. Genotyping in Sickle Cell Disease Patients: The French Strategy. *Transfus Med Hemother* 2018;45:264-70.
49. Chou ST, Evans P, Vege S, et al. RH genotype matching for transfusion support in sickle cell disease. *Blood* 2018;132:1198-207.
50. Gaspardi AC, Sippert EA, De Macedo MD, et al. Clinically relevant RHD-CE genotypes in patients with sickle cell disease and in African Brazilian donors. *Blood Transfus* 2016;14:449-54.
51. Pham BN, Peyrard T, Juszczak G, et al. Alloanti-c (RH4) revealing that the (C)ce s haplotype encodes a partial c antigen. *Transfusion* 2009;49:1329-34.
52. Aygun B, Padmanabhan S, Paley C, et al. Clinical significance of RBC alloantibodies and autoantibodies in sickle cell patients who received transfusions. *Transfusion* 2002;42:37-43.
53. Noizat-Pirenne F, Le Pennec PY, Mouro I, et al. Molecular background of D(C)(e) haplotypes within the white population. *Transfusion* 2002;42:627-33.
54. Hemker MB, Ligthart PC, Berger L, et al. DAR, a new RhD variant involving exons 4, 5, and 7, often in linkage with ceAR, a new Rhce variant frequently found in African blacks. *Blood* 1999;94:4337-42.
55. Noizat-Pirenne F, Lee K, Pennec PY, et al. Rare RHCE phenotypes in black individuals of Afro-Caribbean origin: identification and transfusion safety. *Blood* 2002;100:4223-31.
56. Westhoff CM, Vege S, Horn T, et al. RHCE*ceMO is frequently in cis to RHD*DAU0 and encodes a hr(S) -, hr(B) -, RH:-61 phenotype in black persons: clinical significance. *Transfusion* 2013;53:2983-9.
57. Reid ME, Halter Hipsky C, Hue-Roye K, et al. The low-prevalence Rh antigen STEM (RH49) is encoded by two different RHCE*ce818T alleles that are often in cis to RHD*DOL. *Transfusion* 2013;53:539-44.
58. Roussel M, Poupel S, Nataf J, et al. RHD*DOL1 and RHD*DOL2 encode a partial D antigen and are in cis with the rare RHCE*ceBI allele in people of African descent. *Transfusion* 2013;53:363-72.
59. Halter Hipsky C, da Costa DC, Omoto R, et al. Prevalence of RHD*DOL and RHCE*ce(818T) in two populations. *Immunohematology* 2011;27:66-7.
60. Prisco Arnoni C, Guilhem Muniz J, de Paula Vendrame TA, et al. RHCE variants inherited with altered RHD alleles in Brazilian blood donors. *Transfus Med* 2016;26:285-90.
61. Westhoff CM, Vege S, Halter-Hipsky C, et al. DIIIa and DIII Type 5 are encoded by the same allele and are associated with altered RHCE*ce alleles: clinical implications. *Transfusion* 2010;50:1303-11.
62. Hipsky CH, Lomas-Francis C, Fuchisawa A, et al. RHCE*ceCF encodes partial c and partial e but not CELO, an antigen antithetical to Crawford. *Transfusion* 2011;51:25-31.
63. Döschner A, Vogt C, Bittner R, et al. RHCE alleles detected after weak and/or discrepant results in automated Rh blood grouping of blood donors in Northern Germany. *Transfusion* 2009;49:1803-11.
64. Noizat-Pirenne F, Mouro I, Gane P, et al. Heterogeneity of blood group RhE variants revealed by serological analysis and molecular alteration of the RHCE gene and transcript. *Br J Haematol* 1998;103:429-36.
65. Greenwalt TJ, Sanger R. The Rh antigen Ew. *Br J Haematol* 1955;1:52-4.
66. Grobel RK, Cardy JD. Hemolytic disease of the newborn due to anti-EW. A fourth example of the Rh antigen, EW. *Transfusion* 1971;11:77-8.
67. Cruz BR, de Souza Silva TC, de Souza Castro B, et al. Molecular matching for patients with haematological diseases expressing altered RHD-RHCE genotypes. *Vox Sang* 2019;114:605-15.
68. Dezan MR, Oliveira VB, Conrado MCAV, et al. Variant genotypes associated with reduced expression of RhCE antigens among Brazilian blood donors. *Transfusion* 2021;61:1923-31.
69. Dezan MR, Ribeiro IH, Oliveira VB, et al. RHD and

- RHCE genotyping by next-generation sequencing is an effective strategy to identify molecular variants within sickle cell disease patients. *Blood Cells Mol Dis* 2017;65:8-15.
70. Fichou Y, Le Maréchal C, Scotet V, et al. Insights into RHCE Molecular Analysis in Samples with Partial D Variants: the Experience of Western France. *Transfus Med Hemother* 2015;42:372-7.
 71. Flores-Bello A, Mas-Ponte D, Rosu ME, et al. Sequence diversity of the Rh blood group system in Basques. *Eur J Hum Genet* 2018;26:1859-66.
 72. Granier T, Beley S, Chiaroni J, et al. A comprehensive survey of both RHD and RHCE allele frequencies in sub-Saharan Africa. *Transfusion* 2013;53:3009-17.
 73. Jia S, Chen J, Wen J, et al. Serological screening and genetic analysis of RhCE variants in the Chinese Southern Han donors. *Transfus Med* 2021;31:271-6.
 74. Ouchari M, Jemni Yacoub S, Houissa B, et al. System RH: screening of partials D with RHD/RHCE hybrid gene. *Transfus Clin Biol* 2013;20:35-9.
 75. Pham BN, Peyrard T, Juszczak G, et al. Analysis of RhCE variants among 806 individuals in France: considerations for transfusion safety, with emphasis on patients with sickle cell disease. *Transfusion* 2011;51:1249-60.
 76. Souza Silva TC, Cruz BR, Costa SS, et al. RHD and RHCE molecular analysis in weak D blood donors and in patients with Rh antibodies against their own corresponding Rh antigen. *Blood Transfus* 2020;18:295-303.
 77. de Almeida FAA, Dezan MR, Oliveira VB, et al. Effectiveness of strategies to screen for blood donors with RH variants in a mixed population. *Transfus Apher Sci* 2020;59:102720.
 78. Castro O, Sandler SG, Houston-Yu P, et al. Predicting the effect of transfusing only phenotype-matched RBCs to patients with sickle cell disease: theoretical and practical implications. *Transfusion* 2002;42:684-90.
 79. Muniz JG, Arnoni C, Medeiros R, et al. Antigen matching for transfusion support in Brazilian female patients with sickle cell disease to reduce RBC alloimmunization. *Transfusion* 2021;61:2458-67.
 80. Karafin MS, Westlake M, Hauser RG, et al. Risk factors for red blood cell alloimmunization in the Recipient Epidemiology and Donor Evaluation Study (REDS-III) database. *Br J Haematol* 2018;181:672-81.
 81. Davis BA, Allard S, Qureshi A, et al. Guidelines on red cell transfusion in sickle cell disease Part II: indications for transfusion. *Br J Haematol* 2017;176:192-209.
 82. Chou ST, Alsawas M, Fasano RM, et al. American Society of Hematology 2020 guidelines for sickle cell disease: transfusion support. *Blood Adv* 2020;4:327-55.
 83. Vichinsky EP, Luban NL, Wright E, et al. Prospective RBC phenotype matching in a stroke-prevention trial in sickle cell anemia: a multicenter transfusion trial. *Transfusion* 2001;41:1086-92.
 84. Sakhalkar VS, Roberts K, Hawthorne LM, et al. Allosensitization in patients receiving multiple blood transfusions. *Ann N Y Acad Sci* 2005;1054:495-9.
 85. Chou ST, Flanagan JM, Vege S, et al. Whole-exome sequencing for RH genotyping and alloimmunization risk in children with sickle cell anemia. *Blood Adv* 2017;1:1414-22.
 86. Lapadat R, Meyer E, Josephson C, et al. Exon Sequencing Improves Red Blood Cell Phenotype Matching in a Patient Homozygous for the RHCE*ce48C Allele. *Transfusion* 2015;55:152A.
 87. MA Keller PM, J Maurer, P Ligthart, et al. Identifying compatible blood for RH alloimmunized patients: an international case 29th Regional Congress of ISBT: Vox Sanguinis, 2019:85.
 88. Silvy M, Tournamille C, Babinet J, et al. Red blood cell immunization in sickle cell disease: evidence of a large responder group and a low rate of anti-Rh linked to partial Rh phenotype. *Haematologica* 2014;99:e115-7.
 89. Tormey CA, Hendrickson JE. Transfusion-related red blood cell alloantibodies: induction and consequences. *Blood* 2019;133:1821-30.
 90. Noizat-Pirenne F, Tournamille C. Relevance of RH variants in transfusion of sickle cell patients. *Transfus Clin Biol* 2011;18:527-35.
 91. Thonier V. Immuno-hematological findings in Delayed Hemolytic Transfusion Reaction (DHTR). *Transfus Clin Biol* 2019;26:102-8.
 92. Noizat-Pirenne F, Bachir D, Chadebech P, et al. Rituximab for prevention of delayed hemolytic transfusion reaction in sickle cell disease. *Haematologica* 2007;92:e132-5.
 93. Floch A. Molecular genetics of the Rh blood group system: alleles and antibodies—a narrative review. *Ann Blood* 2021;6:29.
 94. Westhoff CM, Silberstein LE, Wylie DE. Evidence supporting the requirement for two proline residues for expression of c. *Transfusion* 2000;40:321-4.
 95. Hipsky CH, Lomas-Francis C, Fuchisawa A, et al. RHCE*ceAR encodes a partial c (RH4) antigen. *Immunohematology* 2010;26:57-9.
 96. Peyrard T, Pham BN, Poupel S, et al. Alloanti-c/ce in a c+ceAR/Ce patient suggests that the rare RHCE ceAR

- allele (ceAR) encodes a partial c antigen. *Transfusion* 2009;49:2406-11.
97. Pham BN, Peyrard T, Tourret S, et al. Anti-HrB and anti-hrb revisited. *Transfusion* 2009;49:2400-5.
 98. Shapiro M, Le Roux M, Brink S. Serology and genetics of a new blood factor: hr B . *Haematologia (Budap)* 1972;6:121-8.
 99. Moores P. Rh18 and hrS blood groups and antibodies. *Vox Sang* 1994;66:225-30.
 100. Shapiro M. Serology and genetics of a new blood factor: hrS. *J Forensic Med* 1960;7:96-105.
 101. Coleman S, Westhoff CM, Friedman DE, et al. Alloimmunization in patients with sickle cell disease and underrecognition of accompanying delayed hemolytic transfusion reactions. *Transfusion* 2019;59:2282-91.
 102. Moreira Júnior G, Bordin JO, Kuroda A, et al. Red blood cell alloimmunization in sickle cell disease: the influence of racial and antigenic pattern differences between donors and recipients in Brazil. *Am J Hematol* 1996;52:197-200.
 103. Garratty G. Severe reactions associated with transfusion of patients with sickle cell disease. *Transfusion* 1997;37:357-61.
 104. Vichinsky EP, Earles A, Johnson RA, et al. Alloimmunization in sickle cell anemia and transfusion of racially unmatched blood. *N Engl J Med* 1990;322:1617-21.
 105. Castilho L, Dinardo CL. Optimized Antigen-Matched in Sickle Cell Disease Patients: Chances and Challenges in Molecular Times - the Brazilian Way. *Transfus Med Hemother* 2018;45:258-62.
 106. Van Buren NL, Gorlin JB, Corby SM, et al. How do I incorporate red cell genotyping to improve chronic transfusion therapy? *Transfusion* 2020;60:16-25.
 107. Chou ST, Westhoff CM. Molecular biology of the Rh system: clinical considerations for transfusion in sickle cell disease. *Hematology Am Soc Hematol Educ Program* 2009;178-84.
 108. Arnoni CP, Latini FR, Muniz JG, et al. A simple approach to screen rare donors in Brazil. *Immunohematology* 2015;31:20-3.

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Table S1 Detailed search strategy used in PubMed database

Search	Query	Results
#7	Filters: English language, publication dates from 1000/1/1 [‡] -2021/6/30	325
#6	#4 OR #5	352
#5	#2 AND #3	43
#4	#1 AND #2	336
#3	cell disease, sickle[MeSH Terms]	24,283
#2	("RHCE") OR (" RHCE variants") OR ("Rh antibodies")	959
#1	Rh-Hr Blood-Group System/genetics*[MeSH Terms]	1,647

[‡], corresponds to the limit of the database.