

References

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Table S1 Inclusion and exclusion criteria

Inclusion criteria

- Published in the English language
- Peer-reviewed journals
- Specified radiological method (e.g., radiography, CT, MRI)
- Adult human patients with cervical OPLL
- Use of deep learning model (CNN)

Exclusion criteria

- All non-English languages
- Commentaries, case reports, narrative reviews, letters to editors, books
- Animal studies or lab-based studies
- Studies on children and adolescents (<18 years)
- Patient with previous spinal surgery

CNN, convolutional neural network; CT, computed tomography; MRI, magnetic resonance imaging; OPLL, ossification of the posterior longitudinal ligament.

Table S2 Search strategy conducted on October 1st, 2023, delineating the databases used, search terms employed, publication dates, and the corresponding results retrieved from each database

| Database | Search terms | Publication dates | Results (n) |
|------------------|--|-------------------|-------------|
| Google Scholar | “Deep Learning” AND “Ossification of the Posterior Longitudinal Ligament” | 1998–2023 | 106 |
| ScienceDirect | “Deep Learning” AND “Ossification of the Posterior Longitudinal Ligament” | 2020–2023 | 10 |
| PubMed | (“deep learning”[MeSH Terms] OR (“deep”[All Fields] AND “learning”[All Fields]) OR “deep learning”[All Fields]) AND (“ossification of posterior longitudinal ligament”[MeSH Terms] OR (“ossification”[All Fields] AND “posterior”[All Fields] AND “longitudinal”[All Fields] AND “ligament”[All Fields]) OR “ossification of posterior longitudinal ligament”[All Fields]) | 2021–2023 | 6 |
| BASE | “Deep Learning” AND “Ossification of the Posterior Longitudinal Ligament” | 2021–2023 | 6 |
| Cochrane Library | “Deep Learning” AND “Ossification of the Posterior Longitudinal Ligament” | 2021 | 1 |

Table S3 Level of evidence of each of the included studies based on the Oxford Centre of Evidence-Based Medicine (OCEBM) Levels of Evidence (39)

| Study number | Author, Year | Level of evidence |
|--------------|------------------------------|-------------------|
| 1 | Ogawa <i>et al.</i> (2022) | 3 |
| 2 | Miura <i>et al.</i> (2021) | 3 |
| 3 | Murata <i>et al.</i> (2021) | 3 |
| 4 | Chae <i>et al.</i> (2022) | 3 |
| 5 | Tamai <i>et al.</i> (2022) | 3 |
| 6 | Ito <i>et al.</i> (2023) | 2b |
| 7 | Shemesh <i>et al.</i> (2023) | 3 |

Table S4 Grading of Recommendations, Assessment, Development and Evaluations (GRADE) scoring for all studies (40)

| Study | Risk of Bias | Imprecision | Inconsistency | Indirectness | Publication bias | Overall |
|------------------------------|--------------|-------------|---------------|--------------|------------------|----------|
| Ogawa <i>et al.</i> (2022) | Low | Low | Low | Moderate | Low | Moderate |
| Miura <i>et al.</i> (2021) | Moderate | Moderate | Low | Very low | Low | Moderate |
| Murata <i>et al.</i> (2021) | Low | Low | Low | Moderate | Low | Moderate |
| Chae <i>et al.</i> (2022) | Low | Low | Low | Moderate | Low | Moderate |
| Tamai <i>et al.</i> (2022) | Low | Moderate | Low | Low | Low | Moderate |
| Ito <i>et al.</i> (2023) | Low | Moderate | Low | Low | Low | Moderate |
| Shemesh <i>et al.</i> (2023) | Moderate | Low | Very low | Very low | Low | Moderate |

Table S5 Table of extracted items from both the qualitative synthesis (systematic review) and quantitative synthesis (meta-analysis)

Extracted items for qualitative synthesis

Study, sample size, study type and design, country, deep learning model, model construction, radiological technique, control, patient demographic, indication for radiology, accuracy, AUC, sensitivity, specificity, TP, FN, TN, FP, F1 value, k1 value, recall, precision, NPV, PPV, OPLL subtypes, main conclusion, risk of bias

Extracted items for quantitative synthesis

Accuracy, AUC, sensitivity, OPLL subtypes, human performance

AUC, area under the curve; FN, false negative; FP, false positive; NPV, negative predictive value; OPLL, ossification of the posterior longitudinal ligament; PPV, positive predictive value; TN, true negative; TP, true positive.

Table S6 Risk of bias analysis of all included studies (QUADAS-2 tool) (41)

| Study | Risk of Bias | | | | Applicability Concerns | | |
|------------------------------|-------------------|------------|--------------------|-----------------|------------------------|------------|--------------------|
| | Patient selection | Index test | Reference standard | Flow and timing | Patient selection | Index test | Reference standard |
| Ogawa <i>et al.</i> (2022) | Unclear | Low | Unclear | Low | Low | Unclear | Low |
| Miura <i>et al.</i> (2021) | Low | Low | Low | Low | Low | Unclear | Unclear |
| Murata <i>et al.</i> (2021) | Low | Unclear | Low | Low | Low | Low | Low |
| Chae <i>et al.</i> (2022) | Low | Low | Low | Low | Low | Unclear | Unclear |
| Tamai <i>et al.</i> (2022) | Low | Unclear | Unclear | Low | Low | Low | Low |
| Ito <i>et al.</i> (2023) | Low | Unclear | Low | Low | Low | Low | Low |
| Shemesh <i>et al.</i> (2023) | Low | Low | Low | Low | Unclear | Unclear | Unclear |

Table S7 Table of extracted outcome variables with an definition for each variable (42-44)

| Variable | Definition |
|---------------------|--|
| Accuracy | Accuracy is the ratio of correctly predicted positive and negative observations to the total observations. Formula: $(TP + TN)/(TP + TN + FP + FN)$ |
| Sensitivity | Sensitivity (also known as true positive rate/recall) measures the percentage of actual positive cases that were correctly identified. Formula: $TP/(TP + FN)$ |
| Specificity | Specificity (also known as true negative rate) indicates how well a test correctly identifies negative cases. Formula: $TN/(TN + FP)$ |
| Precision | Ratio of true positive predictions to the total predicted positives. Formula: $TP/(TP + FP)$ |
| AUC | The AUC measures the classifier's performance across all possible threshold values and represents the area under the ROC curve |
| ROC | ROC is a graphical representation of the trade-off between sensitivity and specificity at various thresholds. |
| NPV | The NPV is the percentage of actual negative cases that were correctly identified. Formula: $TN/(TN + FN)$ |
| PPV | The PPV indicates the likelihood of actual positive results being correctly identified. Formula: $TP/(TP + FP)$ |
| K1 score | The F1 score is the harmonic mean of precision and recall. It provides a balance between precision and recall, taking both false positives and false negatives into account. Formula: $2 * (\text{precision} * \text{recall})/(\text{precision} + \text{recall})$, where $\text{precision} = TP/(TP + FP)$ and $\text{recall} = TP/(TP + FN)$ |
| Cohen's kappa score | Cohen's kappa score is a measure of inter-rater agreement for categorical items. It considers the agreement occurring by chance and adjusts the observed agreement accordingly. The formula involves observed and expected agreement between two raters or classifiers |

TP, true positive; TN, true negative; FP, false positive; FN, false negative; AUC, area under the curve; ROC, receiver operating characteristic; NPV, negative predictive value; PPV, positive predictive value.

Table S8 OPLL subtypes definitions (based on Tanaka *et al.* and proposed by the Investigation Committee on OPLL of the Japanese Ministry of Public Health and Welfare by Tsuyama *et al.*). (45,46)

| OPLL subtype | Definition |
|--------------|--|
| Segmental | Involves ossification behind each vertebral body |
| Continuous | Ossified mass that spans several vertebral bodies and the intervening disk spaces |
| Mixed | Mixture of both continuous and segmental types |
| Localized | Ossification is localized to the intervertebral disk space without involvement of the vertebral body |

OPLL, ossification of the posterior longitudinal ligament.

Table S9 The R code utilized in this review

| Graph | Code |
|---------------------------|---|
| World Map (47) | <pre>install.packages("rworldmap") library(rworldmap) studien_daten <- data.frame(Land = c("Japan", "S. Korea", "Israel"), Studies = c(5, 1, 1)) worldprep <- getMap() worldprep\$Studies <- NA for (i in 1:nrow(studien_daten)) { country_name <- studien_daten\$Land[i] worldprep\$Studies[worldprep\$NAME == country_name] <- studien_daten\$Studies[i] } mapCountryData(mapToPlot = worldprep, nameColumnToPlot = "Studies", xlim = c(20, 180), # Begrenzung der Längengrade ylim = c(-10, 100), # Begrenzung der Breitengrade catMethod = c(0:5), colourPalette = "heat", addLegend = TRUE, borderCol = "black", mapTitle = "", # Leere Graph-Überschrift aspect = 1, missingCountryCol = "lightgrey", add = FALSE, nameColumnToHatch = TRUE, lwd = 0.5, oceanCol = NA) title("Number of studies per country", line = -7)</pre> |
| Robvis traffic light (48) | <pre>install.packages("robvis") library(robvis) summary_plot <- rob_traffic_light(data = Risk_of_Bias_assessment, tool = "QUADAS-2", colour = "cochrane", psize = 10) summary_plot</pre> |
| Robvis summary (48) | <pre>library(robvis) summary_plot <- rob_summary(data = Risk_of_Bias_assessment, tool = "QUADAS-2", weighted = FALSE, overall = TRUE) summary_plot</pre> |
| Upset (49) | <pre>install.packages("UpSetR") library(UpSetR) library(readxl) main_bar_color <- "choose color" matrix_color <- "choose color" excel_data <- readxl::read_excel study <- as.data.frame(excel_data) upset(fromList(study), sets = names(study), sets.bar.color = "choose color", main.bar.color = main_bar_color, matrix.color = matrix_color)</pre> |

Table S9 (continued)

Table S9 (continued)

| Graph | Code |
|---------------------|--|
| Funnel (50) | <pre>install.packages("ggplot2") install.packages("meta") library(ggplot2) library(meta) library(readxl) excel_data <- read_excel meta_analysis <- metaprop(event = excel_data\$Sensitivity * excel_data\$SampleSize, n = excel_ data\$SampleSize) funnel(meta_analysis)</pre> |
| Scatter plot matrix | <pre>install.packages("readxl") library(readxl) excel_data <- read_excel plot <- pairs(~ Accuracy + AUC + Sensitivity + Specificity, data = excel_data, main = "Scatterplot Matrix", labels = c("Accuracy", "AUC", "Sensitivity", "Specificity"), cex.labels = 2, font.labels = 2)</pre> |
| Metaanalysis (50) | <pre>library(meta) library(readxl) data <- read_excel(excel_file) overall_meta <- metaprop(event = data\$Events, n = data\$Total, studlab = data\$Study, byvar = data\$OPLL, comb.random = TRUE) forest(overall_meta, fixed = FALSE, random = TRUE, overall = TRUE, col.random = "red", prediction = TRUE, text.fixed = "Test for subgroup differences", rowsize = 2, cex = 0.2)</pre> |